Molecular, chemical and morphological evidence for hematite biogenicity at the Quaternary Cape Vani Mn-(Ba-Fe) deposit, Milos, Greece

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MOLECULAR, CHEMICAL AND MORPHOLOGICAL EVIDENCE FOR HEMATITE BIOGENICITY AT THE QUATERNARY CAPE VANI MN-(BA-FE) DEPOSIT, MILOS, GREECE

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Abstract

Many aspects of the biotic or abiotic origin of iron-rich sedimentary rocks consisting mainly of hematite, an important indicator for exobiology, remain unresolved. Here, we use combined optical microscopy, scanning electron microscopy and energy dispersive X-ray spectrometry (SEM-EDS), Raman spectroscopy and time-of-flight secondary ion mass spectrometry (TOF-SIMS), to image the spatial distribution, and determine the composition, of potential biogenic markers (microbial microfossils, trace elements, organic ion species) in hematite-microstromatolites and oncolite-like microstructures. These structures are identified in iron-rich material cementing a Quaternary fossil-beach conglomerate deposit in the Cape Vani area, NW Milos Island, Greece. The combined detection of morphological, chemical, and molecular organic-ion, biomarkers closely associated with possible hematite microfossils within microstromatolite laminae, strongly supports microbial mediation for their formation, and indicates that hematitic microlamination may be used as a biosignature for Fe-rich biomats on Earth and for their detection in extra-terrestrial materials. The Cape Vani hematite-microstromatolites may contribute to growing our understanding of the function of microorganisms in the genesis of modern and ancient Fe deposits.

Key words: TOF-SIMS, microfossils, organic biomarkers, microstromatolites.

Περίληψη

Η γένεση ιζηματογενών πετρωμάτων πλούσιων στην σίδηρο, κυρίως σε σίδηρη, ιζηματογενής ή αβιοτικός αρχής, ή κάποιος συνδυασμός αυτών, παρατίθεται αλήθεια επιστημονικού προβλήματος. Στην εργασία αυτή, χρησιμοποιήσαμε συνδυασμό αναλυτικών μεθόδων που συμπεριλαμβάνουν οπτική μικροσκοπία, ηλεκτρονική μικροσκοπία σάρωσης, μικροανάλυση (SEM-EDS), φασματοσκοπία Raman και φασματομετρία μάζας χρόνου πτήσης δευτερογενών ηλεκτρονίων (time-of-flight secondary ion mass spectrometry, TOF-SIMS), για να προσβάσουμε την γεωοργική κατανομή και την σύσταση εν δυνάμει γεωβιολογικών δεικτών (π.χ. μικροβιακά
μικροαπολιθώματα, ιχνοστοιχεία, οργανικές ιοντικές ενώσεις) σε αιματιτικούς μικροστρωματόλιθους και μικροδομές από αιματίτη που μοιάζουν με oncolites (ονκόλιθους). Οι μικροδομές αυτές προσδιορίστηκαν σε υλικό πλούσιο σε σίδηρο που αποτελεί συνδετικό υλικό παράλιου κροκαλοπαγούς Τεταρτογενούς ηλικίας από την ευρύτερη περιοχή του ακρωτηρίου Βάνι, στη ΒΔ Μήλο. Ο προσδιορισμός μορφολογικών, χημικών και οργανικών μοριακών ιοντικών βιοδομών σε συνδυασμό με πιθανά μικροαπολιθώματα από αιματίτη, υποδηλώνουν μικροβιακή μεσολάβηση στον σχηματισμό των τελευταίων, και εποδεικνύουν ότι μικροστρωματόλιθοι από αιματίτη μπορεί να χρησιμοποιηθούν ως βιοδείκτες για τη ύπαρξη σιδηρούχων βιοδείκτες για την ανίχνευση σιδηρούχων βιοδείκτες στη Γη, και για την ανίχνευση τους σε εξωγήινα υλικά. Η μελέτη των αιματιτικών μικροστρωματόλιθων του ακρωτηρίου Βάνι μπορεί να συμβάλει στην κατανόηση των λειτουργιών των μικροοργανισμών στη γένεση τόσο σύγχρονων όσο και παλαιών κοιτασμάτων σιδήρου.

Αύξεις κλειδία: TOF-SIMS, μικροαπολιθώματα, οργανικοί βιοδείκτες, μικροστρωματόλιθοι.

1. Introduction

Many aspects of the sedimentary origin of iron-rich rocks and soils, consisting mainly of hematite, an important indicator for exobiology, remain unresolved (Bekker et al., 2010; Bishop, 1998). Most prismatic or platy hematite in an iron-ore formation is the result of post depositional or diagenetically late upgrade processes; however, the Fe oxidation mechanisms, i.e. biological versus non-biological, currently are not determined with certainty (Ohmoto et al., 2006; Beukes and Gutzmer, 2008). Primary micron-sized hematite spheroids are scarce in the geological record, and they have only been reported from modern iron oxide deposits of the Red Sea (Taitel-Goldman and Singer, 2002), and Archean banded iron formations (BIF) of the Hamersley Province, Australia (Ayres et al., 1972); these hematite spheroids have been interpreted as aqueous deposited low-temperature early diagenetic products (Bekker et al., 2010). Moreover, nanophase (nm-sized) grey spherulitic hematite, known to be formed by both biotic and abiotic aqueous-fluid processes on Earth, has been spectroscopically identified on the Martian surface, and raised the possibility of liquid water and biological activity on Mars (Bishop, 1998).

Here, we are undertaking a laboratory study of laminated iron oxide-bearing cement samples from a fossil-beach conglomeratic deposit, NW Milos Island, Greece. The conglomerate forms an interbed in Early Quaternary volcanioclastic sandstones/sandy tufts which host the Cape Vani Mn-Ba-Fe oxide deposit that is known as a distinct littoral to tidal-flat metallogenic environment related to the activity of ancient microbial mats and white smoker-type seafloor hydrothermal venting (Kilias, 2012). The scope of the paper is to document micron-sized spheroid hematite occurrences, and morphologic (i.e. fossil microbes, filaments, or biofilm) and chemical (i.e. organic) biomarkers, in an attempt to decipher the origin of hematite.

2. Materials and Methods

Polished sections of the Vani fossil-beach conglomeratic ferruginous cement is prepared for study. These were sputtered with either carbon or gold for avoiding charging in insulating areas, for EPA and ToF-SIMS analysis. Especially for the ToF-SIMS analysis, a smaller sample was cut to allow its easy insertion into the sample holder and the ultra-high vacuum chamber. This was also required to allow quick vacuum generation, since the sample contains a high number of pores. Optical, scanning electron microscopy and analysis (SEM/EDS) (National and Kaposdistrian University of Athens and National Technical University of Athens), micro-Raman (National Technical University of Athens) and Time-of-Flight mass spectrometry (University of Manchester) techniques were used to investigate the ferruginous cement. Reflected optical

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microscopy and imaging was performed with a Carl Zeiss Axioskop 40 microscope at various magnifications. Scanning electron microscopy and electron probe analysis were performed on a Jeol 6380LV electron microscope. On this instrument, a liquid nitrogen cooled Energy Dispersive Spectrometer (EDS) from Oxford Systems is installed. Acquisition is controlled through the INCA software. To achieve high-resolution secondary electron images and backscattered electron images, the acceleration voltage was set to 30kV and the smallest slit size (1) available was used. Chemical analysis was performed at 20kV and with a beam current on the sample of about 1nA. Raman spectra were acquired with a confocal Renishaw Ramascope RM1000 Raman micro-spectrometer, attached to a Leica DMLM optical microscope. A 632.8nm He-Ne laser is probed on the sample, on a spot size of less than 1.5μm when using the ×100 objective lens and with energy, at the spot, of less than 5mW. Rayleigh scattering is reduced by an edge filter whose cut-off is at about 180cm⁻¹. The entrance slit to the spectrometer was set to 50μm. The spectra acquisition system is composed of a grating of 1800lines/mm and a 576×384pixel Peltier-cooled CCD camera. Spectra acquisition was made in continuous scanning mode. Final spectra are accumulated scans of a few individual spectra, each of 10 seconds integration time. The TOF-SIMS instrument used in this study is the “IDLE” instrument, and it is described in detail in Henkel et al. (2006). Mass-resolved spectra were acquired by sputtering with a pulsed gold (197Au⁺) primary ion beam set at 25kV acceleration voltage. The beam size was about 1μm, defining the spatial resolution of the acquired ion maps. The ion map manipulation and handling software is the “spaceTOF” (Chatzitheodoridis et al., 2005). The surface of the sample was always thoroughly cleaned prior to analysis by sputtering with a direct current ion beam in scanning mode. Positive and negative secondary ion spectra were acquired using both low mass and high mass resolution (about 500 and 2500, respectively).

3. Results
3.1. Morphologic and Mineralogical Biosignatures

The main volume of the sample is a ferruginous deposit, which glues together various detrital crystals, and partly fossilising biological material (cyanobacterial cells and their EPS). The detrital crystals are various common minerals, such as feldspar, Ti-magnetite, zircon and manganese minerals. Optical and scanning electron microscope (SEM) investigation of the ferruginous cement has revealed that it occurs as finely (micrometre scale) laminated colloform or botryoidal structures that bear morphological similarities to microstromatolites (e.g. Allwood et al., 2006; Lazăr et al., 2012) (Figure 1). The microstromatolites consist mainly of a remarkable cyclic repetition of growth laminae consisting of chainlike concentrations of spheroidal/coccoidal structures of variable size, aligned parallel to the microlamination, enclosed in a gel-like mucoid film (Figures 1, 3). Oncoidal microstromatolites also occur, forming the cortices of rounded and bulbous oncoid-like structures, and are developed as closely packed build-ups surrounded by colloform or botryoidal microstromatolites; they range in size from 50 to 200 μm and have a globular to elongated/flattened shape. In general, microlamination is a result of a combination of differential porosity, and weathering of the film material, as well as size variability of the spheroids/coccolids. The laminae are composed mainly of hematite and organics (see below). Detrital silicates (adularia, chlorite) represent a minor component of some laminae and when present, are mainly found in specific detrital layers. This indicates that stromatolite growth was mainly controlled by in situ precipitation.

Hematite in the microstromatolites was identified by Raman spectroscopy (Figure 2). This hematite compares very well to a metamorphic hematite used as reference and to hematite measured by Shim and Duffy (2001), with only minor offsets of the peak positions. One exception is a wide peak centred at 660.6cm⁻¹, which is normally a forbidden band for Raman but IR-active. This peak has been interpreted to represent either surface resonance of incomplete symmetry of hematite, due to stress-induced defects or the first overtone of the 1318.3cm⁻¹ peak (Shim and Duffy, 2001), or, due to impurities (Zoppi et al., 2005). Partial transformation to magnetite due to
the effect of the laser beam has been also suggested (Ulubey et al., 2008; Bouchard and Smith, 2003), however, it seems not applicable here due to the very low energy of the beam used, and its presence to all performed analyses irrespectively of conditions, except for our standard hematite spectrum. A peak at 658 cm$^{-1}$ is reported by Bikiaris et al. (1999) and is attributed to the Al-O-Si bond of clays, e.g. kaolinite. Vani hematite samples do contain traces of clay, however, this peak in our spectra, is interpreted as a result of disorder (Shim and Duffy, 2001) due to its high intensity.

Figure 1 - Backscattered electron (BSE) image of micrometre-scale ferruginous microstromatolites (mic) and oncoidal microstromatolites (onc) cementing a fossil-beach conglomerate, NW Milos Island, Greece.

Figure 2 - Raman spectra of Vani hematite (spectra 1 and 2) compared to a reference highly ordered hematite of metamorphic origin. The Vani hematite 2 spectrum shows additionally a feldspar peak (Fdsp; 513 cm$^{-1}$).
3.2. Molecular Biomarkers

In order to set the inorganic frame on which organics sit, firstly a set of ToF-SIMS ion maps of the major elements is given (Figure 4), together with the total ion image which is the sum for all counts measured in each pixel. Iron ion map coincides with the stromatolitic texture. Dark areas on Figure 4 indicate voids or silicate mineral composition, i.e. such as feldspars, or chlorite. The results obtained by TOF-SIMS demonstrate the presence of Mn, Mg, Si, Na, Ca, Al and lesser K, within the microstromatolitic lamination (Figure 4). This elemental distribution may indicate detrital silicate minerals along the specific lamina, most probably fine distributions of clays, micas and feldspars. Especially on the bright spots that concentrate on the bottom-right of the images, mainly feldspars are located, as evidenced also from electron probe investigations. This zone is also composed of more diffused Mn, Mg, Na and Ca. Probably these elements are present in the matrix of hematite as well. The very limited occurrence of K in this layer compared to the rest of the area, together with the enrichment in Na is intriguing. This might be resolved if the molecular ion maps of hydrocarbons are considered, as shown on Figure 5.

Figure 5 demonstrates that the hematitic lamina already described is preferentially enriched in carbonaceous material in the form of hydrocarbon fragments. Hydrocarbon fragments with odd $(C_{2n+1}H_{x})$ and even $(C_{2n}H_{x})$ number of carbon atoms, as well as organic molecules containing oxygen, seem to be especially rich in this layer. Mono-carbonic chains $(C_n)$ show an even distribution coinciding with the iron-rich area. These chains probably have a different origin e.g. contamination. To investigate the distribution of hydrocarbons as well, a detailed analysis of the mass spectrum was performed and the intensities of the peaks were measured. The result is shown in Figure 6, which demonstrates a systematic abundance of hydrocarbon fragments.
Figure 4 - ToF-SIMS ion maps of major elements. “Total Image” is the total ion image, which is composed of all ions of the spectrum acquired from each pixel.

Figure 5 - ToF-SIMS molecular ion maps of the carbonaceous matter in hematite lamina of the Vani microstromatolites. Note the good correspondence between hematitic coccooidal microfossils and the distribution of globular carbonaceous matter, i.e. $C_5O_2H_3$, $C_7H_7$, and $C_3H_7$. 

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4. Discussion-Conclusions

The biogenicity and syngenicity of the morphological and chemical signatures were evaluated according to a series of published criteria (Westall et al., 2011; Schopf et al., 2010). The Vani hematite-rich conglomeratic cement shows typical features of microstromatolitic microbialites, i.e. laminated organosedimentary deposits resulting from benthic microbial community trapping and binding clastic sediment particles and/or inducing mineral precipitation (Burne and Moore, 1987; Burkhalter, 1995). The microbialites exhibit two main morphological fabrics: (i) hematitic microstromatolites (Burkhalter, 1995), and (ii) hematitic oncoidal microstromatolites (Cavalazzi et al., 2007) (Figure 1). The Vani Fe-microstromatolites show features extremely similar to typical biogenic stromatolites, such as non-isopachous micrometer-scale alternations of smooth to wrinkly growth laminae, linked domical pseudocolumns with continuous bridging laminae between them, hemispherical to bulbous 3-D laminae shape, and thickening of laminae over the crests of the domes (Allwood et al., 2006; Rossi et al., 2006). Hematitic oncoidal microstromatolites form the cortices of rounded and bulbous, oncoid-like structures and occur as closely packed build-ups surrounded by colloform banded microstromatolites; they range in size from 50 to 200 μm and have a globular to elongated/flattened shape. The strongest evidence that these structures are stromatolitic microbialites, however, is the outstanding profusion of fossil microbes in the form of hematitic spheroids, in the stromatolitic laminae. The hematitic spheroids have morphological characteristics consistent with colonies of coccoidal fossil microbial cells (Schopf et al., 2010), and the polymer film maintains morphological details identical to fossil extracellular polymeric substances (EPS) produced by the cells (Westall et al., 2000). This, combined with the alignment of the coccoidal fossil microbes parallel to the convex outward microlamination, offer strong evidence that microbes were active and not fortuitous in the development of such microstructures (Rossi et al., 2006).

Carbonaceous material, i.e. odd and even chained 
\[ C_xH_y \] ionic molecules, 
\[ C_xO_yH_z \] molecules and 
mono-carbolic chains 
\[ C_x \], was identified in the hematite laminae using TOF-SIMS in a highly systematic abundance (Figure 6). Carbonaceous material occurs as micrometer-sized heterogeneously distributed chain-like globular clusters, as well as diffuse film-like material that coats both the hematite microfossils and their substrate, and it is exclusively associated with the hematite laminae. Moreover, according to Siljeström et al. (2010) hopanes and steranes constitute organic lipid biomarkers, indicative of cyanobacteria. Their diagnostic molecular ions are at 191.19amu for hopanes and 217.2amu for steranes. Both peaks exist in the spectra of the studied section of hematite and they follow the same pattern with the rest of the organic fragments (Figure 6). Therefore, it can be inferred that the carbonaceous material is indicative of the existence of biological material trapped in and/or with hematite.

According to its organic nature and in conjunction with the biomorphological evidence presented, the organic-rich hematite spheroidal and filmy material is thus interpreted as Fe- bio-mineralised fossil bacterial cells and associated fossil EPS, likely produced by iron-oxidising micro-organisms (Westall et al., 2000; Heim at al., 2012). Along with the presence of Fe, local selective enrichments of microstromatolitic laminae with inorganic bio-essential elements such as Mn, Na, Ca, ±K detected by ToF-SIMS, further support microbial mediated reactions. The combined detection of morphological, chemical and molecular ion biomarkers in the Cape Vani hematitic microstromatolites and oncoid-like structures clearly supports microbial mediation for their formation, and indicates that this microlamination may be used as a biosignature for Fe-rich biomats on Earth and for their detection in extraterrestrial materials. Moreover, the Cape Vani microstromatolites may contribute to growing our understanding of the function of microorganisms in the genesis of modern and ancient Fe and associated Mn deposits.
Figure 6 - Distribution patterns of mono-carbonic chains ($C_n$), and hydrocarbon fragments with even ($C_{2n}H_x$) and odd ($C_{2n+1}H_x$) number of carbon atoms.

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6. References


