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# Molecular characterization of 18S rDNA partial sequence in *Microcosmus* (Stolidobranchiata, Pyuridae)

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#### Abstract

We present a 18S rDNA based molecular phylogeny of two species of the genus Microcosmus (<u>M.</u> <u>sulcatus</u> and <u>M. claudicans</u>) sampled in the Mediterranean, to investigate their phylogenetic position relative to species of the order Stolidobranchiata. The analysis is based on partial sequences (739 bp) of the 18S rDNA. Among the 18 variable sites found between the two species, 4 correspond to transitions (ts), 14 to transversions (tv) and 4 to deletions/insertions. In the considered Stolidobranchiata, we found 4.3% overall mean number of nucleotide differences and 0.06 (S.E.  $\pm 0.01$ ) Kimura 2-parameter distance. The mean number of nucleotide differences between Microcosmus spp. and other Stolidobranchiata species was of 6% and 0.08 (S.E.  $\pm 0.01$ ) Kimura 2-parameter distance. A molecular phylogeny obtained by Maximum Parsimony corroborates results of the traditional taxonomy.

Keywords: Microcosmus; 18S r DNA; Tunicata; phylogenesis.

#### Introduction

Tunicata represents a subphylum of Chordata widely distributed in the Mediterranean Sea subdivided into three classes: Appendicularia, Ascidiacea and Thaliacea.

Ascidiacea, or ascidians, are by far the largest and most diverse extant group, with over 2500 described species in 14 families (Satoh 1994).

The order Stolidobranchia contains three families of ascidians, the Molgulidae, the Styelidae, and the Pyuridae (HADFIELD *et al.* 1995; HUBER *et al.* 2000; SWALLA *et al.* 2000; SWALLA 2001). According to traditional taxonomy the genus *Microcosmus*  (Heller 1877) is placed in the family of Pyuride.

Though *Microcosmus* formed a sister group of *Pyura* in traditional taxonomy, no molecular phylogenetic tree was created to confirm or contradict this relationship.

Recent extensive molecular phylogenetic reconstruction was developed by STACH and TURBEVILLE (2002), who confirmed the monophyly of Stolidobranchiata as regards the families Styelydae, Molgulidae, and Pyuridae, but there have been no data on any species of the *Microcosmus* genus. Therefore, it is interesting to resolve the molecular phylogeny of *Microcosmus* by analysing the same markers used by Stach and Turbeville.

In the present study we aim to characterise, through 18S rDNA gene sequences, two morphologically distinct species of *Microcosmus* found in the Mediterranean sea i.e. *M. sulcatus* and *M. claudicans*, and to clarify their relative phylogenetic position within tunicates.

*Microcosmus sulcatus* (Coquebert 1797) is exclusively found in the Mediterranean Sea and it is an edible species well known among fishermen. *Microcosmus claudicans* (Savigny 1816), on the other hand, is quite small and rare. Below are presented some distinctive morphological and ecological traits of the two species:

*M. sulcatus* is a species of relatively large dimensions with a hard tunic with many folds, tawny red or brilliant red in colour, siphons with four red lobes, branchial chamber with 7 folds on every side and sexseparated gonads (SALFI 1931; RIEDL *et al.* 1991).

*M. claudicans* has an ovoid body, covered by sand or other deposit materials.Its tunic is thick and hard, with a wrinkled surface and reddish brown in colour. It has a hermaphrodite, undivided gonad for each side of the body; the dorsal folds are fused in one unique membrane, siphons have reddishyellow longitudinal bands and the branchial cambers are supplied with 7–8 and 8–9 folds on the right and the left respectively (SALFI 1931; RIEDL *et al.* 1991).

#### **Materials and Methods**

Ten samples of each species were collected in the Tyrrhenian Sea (Gulf of Naples) at a depth of about 10 metres.

*Microcosmus* individuals in this area can be found on rocky or on organic substrates.

Specimens were randomly collected by snorkelling immersion in the upper intertidal zone and immediately frozen in liquid nitrogen. Once in the laboratory, they were transferred at -80° C until further analysis.

Total DNA was extracted from 1g of mus-

cular tissue, taken from each individual, with a standard proteisane K and phenol/chloroform extraction protocol (SAMBROOK *et al.* 1989).

18S rDNA sequence is the most widely used in previous molecular phylogenetic analyses of Urochordates (HADFIELD *et al.* 1995; SWALLA *et al.* 2000; WADA 1998; WADA *et al.* 1992; WADA & SATOH 1994; STACH & TURBEVILLE 2002), therefore allowing straightforward data integration.

The partial 18S rDNA was amplified by polymerase chain reaction (PCR) using the following primers (STACH & TURBEV-ILLE, 2002):

18S 607-626, 5' TCTGGTCCCAG-CAGCCGCGG 3' 18S 1338-11324r, 5' GAACGGCCAT-GCACCACC 3'.

All PCR reactions were performed in a 50 $\mu$ l volume with 1.7 u of Taq polymerase (Sigma), 100 ng of DNA, 5  $\mu$ l 10x thermal polymerase buffer (Sigma), 0.2 mM MgCl<sub>2</sub>, 200  $\mu$ M dNTPs, and 300 pmol of each primer. The cycling conditions were 35 cycles of 1 min at 94° C, 1 min at 59° C and 1 min 30 s at 72° C, followed by a final step of 5 min at 72° C.

PCR products were purified with a Qiaquick PCR purification kit (Quiagen) and submitted to direct sequencing by use of BigDye<sup>TM</sup> Terminator Cycle Sequencing chemistry (Applied Biosystem product), following the manufacturer's protocols. The sequences were run and analysed with an ABI3100 automated sequencer (Perkin-Elmer).

Sequence alignment and phylogenetic analyses were conducted using the software MEGA, version 2.1 (KUMAR *et al.* 2001).

#### **Results and Discussion**

The utilised primers amplified 739 nt of 18S rDNA for *M. sulcatus* (GeneBank Accession Number DQ149330) and 735 nt for *M. claudicans* (GeneBank Accession Number

[		111	1111111111	1111111111	1111111111	]
[	237788888	99999999001	2222223333	3333444444	4445555667	]
[	7570324678	0235678073	0137890345	6789023456	7890678123	]
#M.sulcatus	CTCTTCTGCT	GGCCCGCCTA	TTAGCGGCGC	TCTCCCCGGT	TTTCGCCATG	
#M.claudicans	TG.AA.G		T			
#Pyura	GC	A.T	.CTT	G.CTTACG	.CTG	
#Botryllus				GNN.NT.ACC		
#Botrylloides				GNN.NTAG		
#Cnemidocarpa				G.CTC-		
#Dendrodoa				G.CTC-		
#Halocynthia				G.CTCC		
#Herdmania				G.CTTA		
#Pelonaia				G.CT.ACC		
#Polyandrocarpa				G.CTGA.C		
#Polycarpa				G.CT.ACC		
#Styela				G.CT.ACC		
#Symplegma				G.CTCC		
#Bostrichobranchus						
				GGAGTTTACC		
#Eugyra #Malawla						
#Molgula	G.1AIG.		C	GAC.TT.TT.	GT.GG	
r	1111111111	1000000000		2222222222	224444444	1
[				33333333333 0011222445		
[ 						-
[				7901369237		1
#M.sulcatus				AGTTTGGCG-		
#M.claudicans						
#Pyura				GCGCC		
#Botryllus				.CGAC		
#Botrylloides				.CGNT.G		
#Cnemidocarpa				GCGC		
#Dendrodoa				GCGC		
#Halocynthia				GCGC		
#Herdmania				GCACA		
#Pelonaia				GCGC		
#Polyandrocarpa				GCGC.AT-		
#Polycarpa				GCGC		
#Styela				GCAC		
#Symplegma				GCG		
#Bostrichobranchus						
#Eugyra				.TGA-		
#Molgula	TG.CGC.G	TTTTAGTT	CGAGGGAT	CCA-	.T-T	
[	4445555555	55555555555	555567]			
[		3333446777	-			
[	5682457363	4568166025	793420]			
#M.sulcatus	GATGGAGCTC	TTC-CT-GTC	ACGTTA			
#M.claudicans	$\texttt{AC}\ldots \texttt{T} \cdot \texttt{T}$	c	CA			
#Pyura		CGTCG	GATGA.			
#Botryllus						
#Botrylloides						
#Cnemidocarpa	C.	CGTCG	GATG			
#Dendrodoa		CGTCG	GATG			
#Halocynthia		CCTCG	GATG			
#Herdmania		CGT.G	GATGA.			
#Pelonaia		TCGTCG				
#Polyandrocarpa		CGTCG				
#Polycarpa		CGTCG				
#Styela		TCGTCG				
#Symplegma		CGTCG				
#Bostrichobranchus						
#Eugyra		CGAGGCGTCG				
#Molgula		CGGGGCGTCG				
····						

Fig. 1: Variable sites in 18S rDNA partial sequences of some Stolidobranchiata species.

DQ149331). From the 10 samples collected, not all tissue gave good quality sequences that could be considered for a comparative analysis. In fact, the results referred to only three sequences for species, and in these we did not observe any significant intraspecific variation. Further investigations into this feature, also using new markers such as Cox1, are in progress.

The nucleotide composition of the sequences averaged 21.2% C, 25.1% T, 23.2% A and 30.5% G. There were 16.6% polymorphic sites, 10% Parsimony informative sites and 6% singleton mutations. Alignment identified 4 deletions/insertions and 18 variable sites between the two species, corresponding to 4 transitions (ts) and 14 transversions (tv) with ts:tv =  $\kappa \approx 0.3$ . This ratio did not deviate significantly from neutral expectation (1:2,  $\chi^2 = 0.0072$ , d.f. = 1, P > 0.05), suggesting that there is no selection acting on the *M. sulcatus* and *M. claudicans* 18S rDNA genes.

The sequences of the two Microcosmus species, were analysed with sequences of 17 Stolidobranchiata species, representative of all genuses of the families: a) Molgulidae. with L12379 (GeneBank Accession Number) Bostrichobranchus digonas, L12414 Eugyra arenosa. L12420 Molgula citrina. AB013016, b) Pyuridae, with Halocynthia roretzi, AF165827 Herdmania curvata, AJ250772 Pyura vittata, and Styelidae, with AF008422 Botrylloides fuscus, AF008424 Botryllus scalaris, AJ250775 Cnemidocarpa clara, AJ250774 Dendrodoa aggregata, L12440 Pelonaia corrugata, AF165825 Polyandrocarpa misakiensis, L12441 Polycarpa pomaria, L12442 Styela clava, AF165826 Symplegma reptans.

In the considered Stolidobranchiata, we found 4.3% overall mean number of nucleotide differences and a 0.06 (S.E.  $\pm 0.01$ ) Kimura 2-parameter distance. The mean number of nucleotide differences between *Microcosmus* spp. and other Stolidobranchiata species was of 6% and 0.08 (S.E.  $\pm 0.01$ ) Kimura 2-parameter distance (Fig. 1).

The phylogenetic analysis was carried

out by Maximum Parsimony using as outgroups two Phlebobranchiata species (Gene Bank Accession Number: AF165820 Ascidia zara, AJ250778 Ciona intestinalis).

The strict consensus of the 42 most parsimonious trees obtained suggests that *Microcosmus* genus is placed within Stolidobranchiata (Fig. 2). Differences between *M. claudicans* and *M. sulcatus* are supported.

In order to fully resolve the phylogenetic relationship of the *Microcosmus* species we provided some molecular information that might be integrated with morphological and ecological knowledge. This is a preliminary study, but further intra- and inter-specific investigations of the other *Microcosmus* species, will be useful to describe the diversity patterns in this group, with important implications in developing appropriate conservation management strategies for coastal ecosystems.

#### Ackowledgements

This short article is devoted to the memory of Mario Milone, recently disappeared.

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*Fig. 2*: Phylogenetic relationship of Stolidobranchiata. Strict consensus of 42 most parsimonious trees. Bootstrap values were calculated for 1,000 replicates using heuristic search strategy with n = 1,000 random addition sequence replicates.

Ascidia zara and Ciona intestinalis represent two species of Phlebobranchiata used as outgroups.

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