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*D. PESSANI, T. TIRELLI, S. FLAGELLA*

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## Key for the identification of Mediterranean brachyuran megalopae

D. PESSANI, T. TIRELLI & S. FLAGELLA

Dipartimento di Biologia Animale e dell'Uomo,  
Via Accademia Albertina 17, 10123 Torino, Italy

e-mail: daniela.pessani@unito.it

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### Abstract

*Based on larval literature, an identification key has been constructed for the megalopae of 55 species of Mediterranean Brachyura. This key is based mainly on external morphological characteristics visible, by using a microscope, limiting the necessity for dissection of specimens. Characteristics used include presence/absence of ornamentation on the carapace, number of abdominal somites, number and position of setae and/or spines on antennula, antenna, pereopods and structure of uropods, etc. Partial dissection is only required to count the setae on the scaphognathite margin. Using the above characters, it is also possible to gather almost all the families into groups. However, the megalopae of Portunidae as well as those of the three species of *Brachynotus* genus are similar to each other and their identification at a specific level requires the use of features somehow variable and difficult to count. In the Majidae, the megalopae of 14 species are already known. The complexity of morphological characters typical of this family makes it difficult to define characters common to all spider crab megalopae. Nevertheless, the key may be an aid for carcinology studies, especially those including sorting and identification of megalopae from plankton samples.*

**Keywords:** Larval development; Larval morphology; Key; Decapoda; Brachyura; Mediterranean Sea.

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### Introduction

The larval development of Brachyura (Crustacea, Decapoda) includes two or more planktonic stages of zoea and the plankto-benthonic stage of megalopa. The identification of megalopae is difficult because:

- there is a lack of adequate descriptions and their usual scarce presence in plankton samples prevents their assignment to known zoeal series;
- during laboratory larval rearing experiments,

obtaining this phase usually results in a high zoeal mortality.

Of the 138 species of Brachyura totally present in the Mediterranean (d'Udekem d'Acoz, 1999), one or more zoeal stages are known for 78 species (PESSANI *et al.*, 1998; GONZALES-GORDILLO *et al.*, 2001), while the complete larval development (zoeal stages plus megalopa) has been described for 65 species. For a few species only the megalopa is known.

A key for the identification of the Mediterranean crab zoeae exists already

(PESSANI *et al.*, 1998); a key for the identification of the zoeae and megalopae of the Northeastern Atlantic crabs was provided by INGLE (1992) while a key for the identification of the zoeae from the Atlantic coast of Europe was provided by PAULA (1996). The aim of the present work is to create a dichotomic key fit for the identification of the megalopa stage of the Mediterranean *Brachyura* species.

## Materials and Methods

To create the key, the literature regarding crabs' larval stages has been taken into account, selecting the works which had provided the complete description and the most accurate illustrations.

The key is based mainly on external morphological characters that are visible, by using an optical microscope, without the necessity of dissecting specimens. Though for some species first antenna requires removal for an adequate observation, the only included character requiring dissection is the number of setae on the scaphognathite (Fig. 1).

## Results

The key includes all the 55 Mediterranean crab species, whose megalopa had been previously described in detail (Table 1).

The key has been constructed by subdividing the species into groups, based first of all on the most evident characters (presence of spines on the carapace, number of abdominal somites, structure of the uropods, etc.), then on characters gradually less easily identifiable or countable. For a correct identification, all the characters indicated in each point must be present in the specimen under examination.

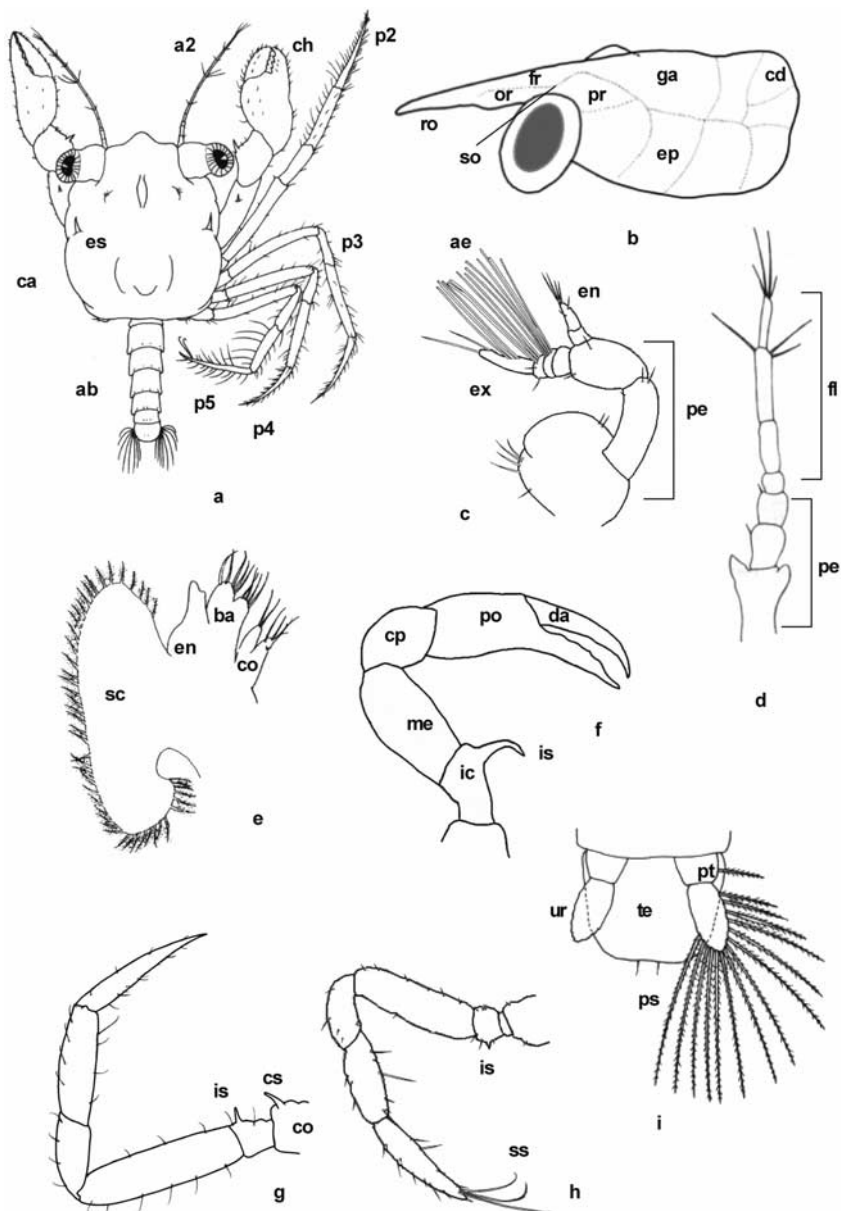
## Considerations

Although the used features are almost all simple and visible, however, they are numerous

and, in some cases, detailed. As a consequence, the incomplete description existing for 12 species does not allow them to be inserted in the key: *Medorippe lanata* (Linnaeus, 1767) by GILET (1952); *Ethusa mascarone* (Herbst, 1785) and *Ilia nucleus* (Linnaeus, 1758) by CANO (1892); *Ebalia cranchii* Leach 1817 and *Xaiva biguttata* (Risso, 1816) by LEBOUR (1928); *Herbstia condyliata* (J. C. Fabricius, 1787), *Lissa chiragra* (J. C. Fabricius, 1775) and *Maja crispata* Risso, 1827 by CANO (1893); *Sirpus zariquieyi* Gordon 1953 by BOURDILLON-CASANOVA (1960); *Asthenognathus atlanticus* Monod, 1933 by BOCQUET (1965); *Planes minutus* (Linnaeus, 1758) by LEBOUR (1944); *Pachygrapsus transversus* (Gibbes, 1850) by ROSSIGNOL (1957). Generally, the use of the key enables an easy identification of the megalopae; but for most of the *Portunidae* species, as well as for the megalopae of the three species of *Brachynotus* genus, it has been necessary to use characters in some way variable and often difficult to count.

The features used in this key do not reflect any systematic arrangement of the *Brachyura* families; nevertheless, with the aid of few characters, it is possible to separate different families or to gather the families into groups (Table 2). For the families not reported in the table, a combination of many different features is necessary. The megalopae of 14 *Majidae* species are already known: the complexity of morphological characters typical of this family – already pointed out by PESSANI & SALTON (1998) as regards planktonic zoeae – prevents us from finding characteristics common to the megalopae of the whole family.

Though the number of Mediterranean species, whose megalopa is actually known, is still small (about 50%), this key may be a useful aid for carcinology studies, especially those including sorting and identification of megalopae in plankton samplings.



**Fig. 1:** Schematic representation of the characters used in the key. *a* = megalopa, dorsal view (from KURATA & MATSUDA, 1980, modified); *b* = carapace, left lateral view (from INGLE, 1992, modified); *c* = antennula; *d* = antenna; *e* = maxilla; *f* = cheliped; *g* = pereiopod 2; *h* = pereiopod 5; *i* = telson and uropods, ventral view (*c*-*i* = from various authors, modified). Abbreviations: *a2* = antenna; *p2*-*p5* = pereiopod 2-5; *ab* = abdomen (6 abdominal somites, telson and uropods; pleopods not visible); *ae* = aesthetascs; *ba* = basis; *ca* = carapace; *cd* = cardiac region; *ch* = cheliped; *co* = coxa; *cp* = carpus; *cs* = coxal spine; *da* = dactyl; *en* = endopod; *ep* = epibranchial region; *es* = epibranchial spine; *ex* = exopod; *fl* = flagellum; *fr* = frontal region; *ga* = gastric region; *ic* = ischium; *is* = ischial spine; *me* = merus; *or* = orbital region; *pe* = peduncle; *po* = propodus; *pr* = protogastric region; *ps* = posterior setae; *pt* = protopod; *ro* = rostrum; *sc* = scaphognathite; *so* = supraorbital region; *ss* = subterminal setae; *te* = telson; *ur* = uropod.

**Table 1**  
**List of the known Mediterranean species of brachyuran megalopae.**

<b>Family</b>	<b>Species</b>	<b>Author</b>
Dromiidae	<i>Dromia personata</i>	RICE <i>et al.</i> , 1970
Homolidae	<i>Homola barbata</i>	WEAR & FIELDER, 1985
Latreillidae	<i>Latreillia elegans</i>	RICE, 1982
Calappidae	<i>Calappa granulata</i>	GUERAO <i>et al.</i> , 1998
Leucosiidae	<i>Ebalia tuberosa</i>	SALMAN, 1982a; INGLE, 1992
Majidae	<i>Acanthonyx lunulatus</i>	GUERAO & ABELLÓ, 1996
	<i>Achaeus cranchii</i>	BOCQUET, 1954; INGLE, 1992
	<i>Dorhynchus thomsoni</i>	WILLIAMSON, 1982; INGLE, 1992
	<i>Eurynome aspera</i>	SALMAN, 1982b; INGLE, 1992
	<i>Eurynome spinosa</i>	INGLE, 1992
	<i>Inachus dorsettensis</i>	CLARK, 1983; INGLE, 1977; INGLE, 1992
	<i>Inachus leptochirus</i>	CLARK, 1983; INGLE, 1992
	<i>Inachus phalangium</i>	CLARK, 1983; INGLE, 1992
	<i>Macropodia rostrata</i>	INGLE, 1982
	<i>Macropodia tenuirostris</i>	SALMAN, 1981; INGLE, 1992
	<i>Maja goletziana</i>	PAULA, 1988A
	<i>Maja squinado</i>	CLARK, 1986
	<i>Pisa armata</i>	INGLE & CLARK, 1980
	<i>Pisa tetraodon</i>	RODRÍGUEZ, 1997
	<i>Rochinia carpenteri</i>	INGLE, 1979
Parthenopidae	<i>Parthenope massena</i>	THIRIOT, 1973
Atelecyclidae	<i>Atelecyclus rotundatus</i>	HONG & INGLE, 1987
	<i>Atelecyclus undecimdentatus</i> (?)	PAULA, 1987; INGLE, 1992
Cancridae	<i>Cancer pagurus</i>	INGLE, 1981
Corystidae	<i>Corystes cassivelaunus</i>	INGLE & RICE, 1971
Pirimelidae	<i>Pirimela denticulata</i>	PAULA, 1987; FLORES & PAULA, 2000
Thiidae	<i>Thia scutellata</i>	INGLE, 1984
Geryonidae	<i>Geryon longipes</i>	GUERAO <i>et al.</i> , 1996
Portunidae	<i>Bathynectes longipes</i>	INGLE, 1985
	<i>Callinectes sapidus</i>	ROFF <i>et al.</i> , 1984; Ingle, 1992
	<i>Carcinus aestuarii</i>	RICE & INGLE, 1975
	<i>Carcinus maenas</i>	RICE & INGLE, 1975
	<i>Liocarcinus arcuatus</i>	INGLE, 1992
	<i>Liocarcinus corrugatus</i>	KIM & HONG, 1999
	<i>Liocarcinus depurator</i>	INGLE, 1992
	<i>Liocarcinus marmoreus</i>	INGLE, 1992
	<i>Liocarcinus pusillus</i>	INGLE, 1992
	<i>Necora puber</i>	INGLE, 1992
	<i>Polybius henslowi</i>	INGLE, 1992
	<i>Portumnus latipes</i>	PAULA, 1988B
	<i>Portunus pelagicus</i>	YATSUKA & SAKAI, 1980
Goneplacidae	<i>Goneplax rhomboides</i>	INGLE & CLARK, 1983
Xanthidae	<i>Eriphia verrucosa</i>	LUMARE & GOZZO, 1972
	<i>Monodaeus couchi</i>	INGLE, 1983
	<i>Pilumnus hirtellus</i>	INGLE, 1983
	<i>Rhithropanopeus harrisi</i>	ROFF <i>et al.</i> , 1984
	<i>Xantho incisus</i>	INGLE, 1983
	<i>Xantho poressa</i>	RODRIGUEZ & MARTIN, 1997
Grapsidae	<i>Brachynotus atlanticus</i>	RODRIGUEZ <i>et al.</i> , 1992
	<i>Brachynotus gemellari</i>	GUERAO <i>et al.</i> , 1995
	<i>Brachynotus sexdentatus</i>	CUESTA <i>et al.</i> , 2000
	<i>Pachygrapsus marmoratus</i>	GUERAO <i>et al.</i> , 1997
Pinnotheridae	<i>Nepinnotheres pinnotheres</i>	INGLE, 1992
	<i>Pinnotheres pisum</i>	INGLE, 1992
Euryplacidae	<i>Eucrate crenata</i>	KURATA & MATSUDA, 1980

**Table 2**  
**Characters which enable separation or grouping of some brachyuran families.**

Main character	Secondary character	Family	
Many long spines on carapace		Latreillidae Majidae (partly)	
5 abdominal somites		Majidae (partly) Pinnotheridae	
6 abdominal somites	biramous uropod	Dromiidae Homolidae	
	long cardiac region spine	Atelecyclidae Cancridae Majidae (partly) Parthenopidae	
	7-segmented antenna	Majidae (partly)	
	20-segmented antenna	Corystidae	
	setae on telson posterior margin		Calappidae Majidae (partly) Geryonidae Portunidae (partly) Xanthidae (partly)
			Grapsidae
		no coxal or ischial spine on cheliped and pereopod 2-5	
		pereopod 5 dactyl with 2 subterminal setae	Thiidae

**Key for the identification of Mediterranean brachyuran megalopae**

(The couplet in brackets indicates the previous point of the key)

- 1a. – 2 pairs of long spines on frontal region; 1 pair of long spines on protogastric region; cheliped and pereopod 2-5 with long and prominent spines ..... *Dorhynchus thomsoni*
- 1b. – not as above ..... 2
- 2a. (1b) – 1 pair of very long and diverging spines on the supraorbital region; a prominent spine, with a small anterior spine, on the gastric region; 1 pair of hook-shaped spines on the epibranchial region ..... *Latreillia elegans*
- 2b. (1b) – not as above ..... 3
- 3a. (2b) – 5 abdominal somites; 4 pairs of pleopods ..... 4
- 3b. (2b) – 6 abdominal somites; 5 pairs of pleopods ..... 11
- 4a. (3a) – 1 pair of long spines on the protogastric region; 1 long spine on the cardiac region ..... (*Macropodia* spp.) 5
- 4b. (3a) – not as above ..... 6
- 5a. (4a) – antennula exopod 4-segmented, with resp. 0, 0, 2, 3 aesthetascs; 23-26 setae on scaphognathite margin ..... *Macropodia tenuirostris*

- 5b. (4a) – antennula exopod 2-segmented, with resp. 2, 4 aesthetascs; 19 setae on scaphognathite margin ..... *Macropodia rostrata*
- 6a. (4b) – 1 pair of spines on the frontal region; spines on abdominal somites ..... 7
- 6b. (4b) – absence of spines on the carapace and abdominal somites ..... 10
- 7a. (6a) – abdominal somites 2-5 resp. with 1, 2, 3, 4 pairs of postero-dorsal spines, some of them very small ..... *Achaeus cranchii*
- 7b. (6a) – abdominal somites 2-5 with 1 pair of small postero-dorsal spines .....  
.....(*Inachus* spp.) 8
- 8a. (7b) – 4 spines on cheliped merus; 16 setae on pereopod 5 propodus .....  
..... *Inachus leptochirus*
- 8b. (7b) – 3 spines on cheliped merus; 15 setae on pereopod 5 propodus ..... 9
- 9a. (8b) – pereopod 2-4 propodus resp. with 16, 16, 17 setae ..... *Inachus dorsettensis*
- 9b. (8b) – pereopod 2-4 propodus resp. with 17, 17, 16 setae ..... *Inachus phalangium*
- 10a. (6b) – tubercle on the cardiac region; 2 setae on antennula terminal flagellum segment .....  
..... *Nepinnotheres pinnotheres*
- 10b. (6b) – no tubercle on the cardiac region; 4 setae on antennula terminal flagellum segment .....  
..... *Pinnotheres pisum*
- 11a. (3b) – uropod endopod present and setose ..... 12
- 11b (3b) – uropod endopod absent ..... 13
- 12a. (11a)– carapace dorsal surface with numerous conical spines; antenna 15-segmented .  
..... *Dromia personata*
- 12b. (11a)– carapace dorsal surface without conical spines; antenna 30-segmented .....  
..... *Homola barbata*
- 13a. (11b)– rostrum and cardiac region spine long and acute ..... 14
- 13b. (11b)– not as above ..... 18
- 14a. (13a)– cardiac region spine dorsally directed; antenna 7-segmented .....  
..... *Rochinia carpenteri*
- 14b. (13a)– cardiac region spine posteriorly directed; antenna 8-9-11-segmented ..... 15
- 15a. (14b)– antenna 8-9-segmented; pereopod 5 dactyl without terminal or subterminal setae .....  
..... *Parthenope massena*
- 15b. (14b)– antenna 11-segmented; pereopod 5 dactyl with 3 terminal or subterminal setae ..... 16
- 16a. (15b)– cheliped with ischial spine; uropod exopod with 8 setae ..... *Cancer pagurus*
- 16b. (15b)– cheliped without ischial spine; uropod exopod with 10 setae .....  
..... (*Atelecycclus* spp.) 17

17a. (16b)– antennula exopod 4-segmented with resp. 0, 5-8, 6-8, 6-7 aesthetascs	.....	<i>Atelecyclus rotundatus</i>
17b. (16b)– antennula exopod 4-segmented with resp. 0, 12, 9, 7 aesthetascs	.....	<i>Atelecyclus undecimdentatus</i> (?)
18a. (13b)– antenna 20-segmented	.....	<i>Corystes cassivelaunus</i>
18b. (13b)– antenna max. 11-segmented	.....	19
19a. (18b)– spines on the protogastric or epibranchial region	.....	20
19b. (18b)– not as above	.....	21
20a. (19a)– protogastric region with a spine; antennula exopod 4-segmented; antenna 10-segmented	.....	<i>Goneplax rhomboides</i>
20b. (19a)– epibranchial region with a spine; antennula exopod 5-segmented; antenna 11-segmented	.....	<i>Eucrate crenata</i>
21a. (19b)– antenna max. 7-segmented; pereopod 5 dactyl without terminal or subterminal setae	.....	22
21b. (19b)– antenna at least 7-segmented; pereopod 5 dactyl with terminal or subterminal setae	.....	29
22a. (21a)– cheliped with a prominent ischio-meral process; 42-44 marginal setae on scaphognathite	.....	<i>Ebalia tuberosa</i>
22b. (21a)– cheliped without ischial process or spine	.....	( <i>Majidae</i> spp.) 23
23a. (22b)– uropod exopod with 3 setae	.....	( <i>Eurynome</i> spp.) 24
23b. (22b)– uropod exopod with 5 setae	.....	25
24a. (23a)– antennula exopod indistinctly segmented; pereopod 3 with an obtuse coxal process	.....	<i>Eurynome aspera</i>
24b. (23a)– antennula exopod 3-segmented with resp. 5, 3, 1 aesthetascs; pereopod 2-4 with a minute coxal process	.....	<i>Eurynome spinosa</i>
25a. (23b)– antennula exopod 4-segmented; telson posterior margin with 2 setae	.....	<i>Acanthonyx lunulatus</i>
25b. (23b)– antennula exopod 3-segmented; telson posterior margin without setae	.....	26
26a. (25b)– an ischial spine only on pereopod 2	.....	( <i>Pisa</i> spp.) 27
26b. (25b)– an ischial spine on pereopod 2-5	.....	( <i>Maja</i> spp.) 28
27a. (26a)– telson posterior margin concave; pleopod 1-4 exopod each with 10 setae	.....	<i>Pisa armata</i>
27b. (26a)– telson posterior margin straight; pleopod 1-4 exopod resp. with 12, 12, 11, 9 setae	.....	<i>Pisa tetraodon</i>
28a. (26b)– 48 setae on scaphognathite margin; pereopod 2-4 with a coxal spine	.....	<i>Maja goltziana</i>



28b. (26b)–34-36 setae on scaphognathite margin; pereiopod 2-4 without a coxal spine .....	<i>Maja squinado</i>
29a. (21b)– uropod exopod with 3 setae .....	<i>Rhithropanopeus harrisi</i>
29b. (21b)– uropod exopod with more than 3 setae .....	30
30a. (29b)– pereiopod 5 dactyl with 5 terminal or subterminal setae; uropod exopod with 10-11 setae .....	31
30b. (29b)– pereiopod 5 dactyl with 2, 3, or 4 terminal or subterminal setae .....	32
31a. (30a)– antennula exopod 3-segmented; pereiopod 4 with a very long coxal spine; no setae on telson posterior margin .....	<i>Portunus pelagicus</i>
31b. (30a)– antennula exopod 4-segmented; pereiopod 4 without coxal spine; telson posterior margin with 6-8 setae .....	<i>Callinectes sapidus</i>
32a. (30b)– pereiopod 5 dactyl with 4 long terminal setae; uropod exopod with 16 setae; telson posterior margin with 3 setae .....	<i>Eriphia verrucosa</i>
32b. (30b)– pereiopod 5 dactyl with 2, or 3 terminal or subterminal setae; other characters not combined .....	33
33a. (32b)– pereiopod 5 dactyl with 2 subterminal setae; antennula exopod 3-segmented; no setae on telson posterior margin; uropod protopod without setae .....	<i>Thia scutellata</i>
33b. (32b)– pereiopod 5 dactyl with 3 terminal or subterminal setae; other characters not combined .....	34
34a. (33b)– telson posterior margin with setae .....	35
34b. (33b)– telson posterior margin without setae .....	39
35a. (34a)– telson posterior margin with 6 setae; 126 setae on scaphognathite margin .....	<i>Calappa granulata</i>
35b. (34a)– not as above .....	36
36a. (35b)– telson posterior margin with 4 setae; 90 setae on scaphognathite margin .....	<i>Monodaeus couchii</i>
36b. (35b)– telson posterior margin with 2, or 3 setae; 30– 80 setae on scaphognathite margin .....	37
37a. (36b)– telson posterior margin with 2-3 setae; 76-80 setae on scaphognathite margin; uropod exopod with 13-15 setae .....	<i>Geryon longipes</i>
37b. (36b)– telson posterior margin with 3 setae; 34-58 setae on scaphognathite margin; uropod exopod with 9-12 setae .....	( <i>Xantho</i> spp.) 38
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38b. (37b)– 34-36 setae on scaphognathite margin; uropod exopod with 9 setae .....	<i>Xantho poressa</i>

- 39a. (34b)– antennula exopod 3-4-segmented; cheliped and pereopod 2-5 without ischial or coxal spines ..... (Grapsidae) 40
- 39b. (34b)– antennula exopod 4-segmented; at least cheliped ischial spine, or pereopod 2 coxal spine, or both ..... 43
- 40a. (39a)– antennula exopod 4-segmented; uropod protopod with 2 setae; uropod exopod with 17 setae ..... *Pachygrapsus marmoratus*
- 40b. (39a)– antennula exopod 3-segmented; uropod protopod with 1 seta; uropod exopod with 9 setae ..... (*Brachynotus* spp.) 41
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- 43a. (39b)– cheliped with an ischial spine; pereopod 2-4 without ischial or coxal spines . 44
- 43b. (39b)– cheliped with or without ischial spine; pereopod 2 with coxal spine ..... 46
- 44a. (43a)– antennula exopod 4-segmented with resp. 0, 6, 5-7, 3-7 aesthetascs; scaphognathite margin with more than 45 setae ..... *Portumnus latipes*
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- 45a. (44b)– telson posterior margin straight or convex ..... *Carcinus aestuarii*
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- 46a. (43b)– cheliped with ischial spine; pereopod 3-4 without coxal spine .....  
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- 46b. (43b)– cheliped with or without ischial spine; pereopod 3-4 with coxal spine ..... 47
- 47a. (46b)– cheliped with ischial spine; pereopod 2-3 with ischial spine .....  
..... *Pilumnus hirtellus*
- 47b. (46b)– cheliped with or without ischial spine; pereopod 2-3 with ischial spine ..... 48
- 48a. (47b)– cheliped without ischial spine; uropod protopod with 2 setae .....  
..... *Liocarcinus corrugatus*
- 48b. (47b)– cheliped with ischial spine; uropod protopod with 0, 1, or 2 setae ..... 49
- 49a. (48b)– antenna 9 (apparently 8) -segmented; scaphognathite margin with 64-68 setae; uropod protopod with 2 setae ..... *Polybius henslowii*
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- 50a. (49b)– antennula exopod with resp. 0, 11, 10, 7 aesthetascs; uropod exopod without seta .....  
..... *Liocarcinus depurator*
- 50b. (49b)– antennula exopod not as above; uropod exopod with a seta ..... 51

- 51a. (50b) .– antennula exopod with resp. 0, 8, 9, 6 aesthetascs; scaphognathite margin with 44-46 setae ..... *Liocarcinus arcuatus*
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- 52a. (51b)– antennula exopod with resp. 0, 10, 7, 5 aesthetascs; antenna peduncle with resp. 3, 2, 2 setae; uropod exopod with 8 – 11 setae ..... *Necora puber*
- 52b. (51b)– antennula exopod and antenna peduncle not as above ..... 53
- 53a. (52b)– antennula exopod with resp. 0, 11, 8, 6 aesthetascs; antenna peduncle with resp. 0(?), 2, 1 setae; uropod exopod with 8-9 setae ..... *Liocarcinus pusillus*
- 53b. (52b)– antennula exopod not as above; antenna peduncle with resp. 4, 2, 2 setae ... 54
- 54a. (53b) – antennula exopod with resp. 0, 7, 7, 7 aesthetascs; uropod exopod with 9 setae .  
..... *Liocarcinus marmoreus*
- 54b. (53b)– antennula exopod with resp. 0, 10, 8, 6 aesthetascs; uropod exopod with 11 setae  
..... *Bathynectes longipes*

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## References

- BOCQUET, C., 1954. Développement larvaire d'*Achaeus cranchii* Leach (Décapode Oxyrhynque). Bulletin de la Société Zoologique de France 79: 50-56.
- BOCQUET, C., 1965. Stages larvaire et juvéniles de *Tritodynamia atlantica* (Th. Monod) (= *Asthenognathus atlanticus* Th. Monod) et position systématique de ce crabe. Cahiers de Biologie Marine 6: 407-418.
- BOURDILLON-CASANOVA, L., 1960. Le méroplancton du golfe de Marseille. Les larves des crustacés décapodes. Recueil des Travaux de la Station Marine d'Endoume 30: 1-286.
- CANO, G., 1892. Sviluppo postembrionale dei Dorippidei, Leucosiadi, Corystoidei e Grapsidi. Memorie della Società Italiana di Scienze Naturali 8: 1-14.
- CANO, G., 1893. Sviluppo e morfologia degli Oxyrhynchi. Mitteilungen aus der Zoologischen Station zu Neapel 10: 527-583.
- CLARK, P. F., 1983. The larval and first crab stages of three *Inachus* species (Crustacea: Decapoda: Majidae): a morphological and statistical analysis. Bulletin of the British Museum (Natural History): Zoology 44: 179-190.
- CLARK, P. F., 1986. The larval stages of *Maja squinado* (Herbst, 1788) (Crustacea: Brachyura: Majidae) reared in the laboratory. Journal of Natural History 20: 825-836.
- CUESTA, J. A., SCHUBART, C. D. & RODRÍGUEZ, A., 2000. Larval development of *Brachynotus sexdentatus* (Risso, 1827) (Decapoda, Brachyura) reared under laboratory conditions, with notes on larval characters of the Varunidae. Invertebrate Reproduction and Development 38: 207-223.
- FLORES, A. A. V. & PAULA, J., 2000. Larval and early juvenile stages of *Pirimela denticulata* (Montagu, 1808) (Crustacea, Brachyura, Pirimelidae) obtained in the laboratory. Journal of Natural History 34: 2123-2143.
- GILET, R., 1952. Métazoé de *Dorippe lanata* (Linné) et sa mégalope. Vie et Milieu 3: 415-420.
- GONZALES-GORDILLO, J. J., DOS SANTOS, A. & RODRIGUEZ, A., 2001. Checklist and annotated bibliography of decapod crustacean larvae from the Southwestern European coast (Gibraltar Strait area). Scientia Marina 65: 275-305.
- GUERAO, G. & ABELLÓ, P., 1996. Larval development of *Acanthonyx lunulatus* (Brachyura,

- Majidae, Epialtinae). *Journal of Plankton Research* 18: 1185-1200.
- GUERAO, G., ABELLÓ, P. & CARTES, J., 1998. Morphology of the megalopa and first crab instar of the shamefaced crab *Calappa granulata* (Crustacea, Brachyura, Calappidae). *Miscellanea Zoologica* 21: 37-47.
- GUERAO, G., ABELLÓ, P. & CASTEJÓN, M. R., 1996. Morphology of the larval stages of the deep-sea crab *Geryon longipes* (Brachyura, Geryonidae). *Journal of Natural History* 30: 505-521.
- GUERAO, G., ABELLÓ, P. & CUESTA, J. A., 1997. Morphology of the megalopa and first crab stage of the mediolittoral crab *Pachygrapsus marmoratus* (Brachyura, Grapsidae, Grapsinae). *Zoosystema* 19: 437-447.
- GUERAO, G., CUESTA, J. A., ABELLÓ, P. & GONZÁLEZ-GORDILLO, J. I., 1995. Larval development of *Brachynotus gemmellari* (Rizza, 1839) (Brachyura, Grapsidae) reared under laboratory conditions. *Journal of Plankton Research* 17: 1143-1161.
- HONG, S. Y. & INGLE, R. W., 1987. Larval development of the circular crab, *Atelecyclus rotundatus* (Olivi) (Crustacea: Brachyura: Atelecyclidae) reared in the laboratory. *Journal of Natural History* 21: 1539-1560.
- INGLE, R. W., 1977. The larval and post-larval development of the scorpion spider crab, *Inachus dorsettensis* (Pennant) (Family: Majidae) reared in the laboratory. *Bulletin of the British Museum (Natural History): Zoology* 30: 329-348.
- INGLE, R. W., 1979. The larval development of the spider crab *Rochinia carpenteri* Thomson (Oxyrhyncha: Majidae) with a review of majid subfamilial larval features. *Bulletin of the British Museum (Natural History): Zoology* 37: 47-66.
- INGLE, R. W., 1981. The larval and post-larval development of the edible crab, *Cancer pagurus* Linnaeus (Decapoda: Brachyura). *Bulletin of the British Museum (Natural History): Zoology* 40: 211-236.
- INGLE, R. W., 1982. The larval and post-larval development of the slender-legged spider crab, *Macropodia rostrata* (Linnaeus) (Oxyrhyncha: Majidae: Inachinae) reared in the laboratory. *Bulletin of the British Museum (Natural History): Zoology* 42: 207-225.
- INGLE, R. W., 1983. A comparative study of the larval development of *Monodaeus couchii* (Couch), *Xantho incisus* (Leach) and *Pilumnus hirtellus* (Linnaeus) (Crustacea: Brachyura: Xanthidae). *Journal of Natural History* 17: 951-978.
- INGLE, R. W., 1984. The larval and post-larval development of the thumb-nail Crab, *Thia scutellata* (Fabricius), (Decapoda: Brachyura). *Bulletin of the British Museum (Natural History): Zoology* 47: 53-64.
- INGLE, R. W., 1985. The larval development of the red swimming crab *Bathynectes longipes* (Risso, 1816) (Crustacea: Decapoda: Portunidae). *Bulletin of the British Museum (Natural History): Zoology* 49: 239-255.
- INGLE, R. W., 1992. Larval Stages of Northeastern Atlantic Crabs. An Illustrated Key. Natural History Museum Publication and Chapman & Hall, London.
- INGLE, R. W. & CLARK, P. F., 1980. The larval and post-larval development of Gibbs's spider crab, *Pisa armata* (Latreille) (family Majidae: subfamily Pisinae) reared in the laboratory. *Journal of Natural History* 14: 723-735.
- INGLE, R. W. & CLARK, P. F., 1983. The larval development of the angular crab, *Goneplax rhomboides* (Linnaeus) (Decapoda: Brachyura). *Bulletin of the British Museum (Natural History): Zoology* 44: 163-177.
- INGLE, R. W. & RICE, A. L., 1971. The larval development of the masked crab, *Corystes cassivelaunus* (Pennant) (Brachyura, Corystidae), reared in the laboratory. *Crustaceana* 20: 271-284.
- KIM, K. B. & HONG, S. Y., 1999. Larval development of the wrinkled swimming crab *Liocarcinus corrugatus* (Decapoda: Brachyura: Portunidae) reared in the laboratory. *Journal of Crustacean Biology* 19: 792-808.
- KURATA, H. & MATSUDA, T., 1980. Larval stages of a goneplacid crab, *Eucrate crenata*, reared in the laboratory. *Bulletin of Nansei Regional Fisheries Research Laboratory* 12: 43-49.
- LEBOUR, M. L., 1928. Larval stages of the Plymouth Brachyura. *Proceedings of the Zoological Society of London* 1928: 473-560.
- LEBOUR, M. L., 1944. Larval crabs from Bermuda. *Zoologica* 29: 113-128.
- LUMARE, F. & GOZZO, S., 1972. Sviluppo larvale del crostaceo Xanthidae *Eriphia verrucosa* (Forsk.) in condizioni di laboratorio. *Bollettino della Pesca, Piscicoltura e Idrobiologia* 27: 185-200.
- PAULA, J., 1987. Planktonic stages of brachyuran crabs from the south-western Iberian coast

- (Crustacea, Decapoda, Brachyura). *Journal of Natural History* 21: 717-756.
- PAULA, J., 1988a. The larval development of the spider crab *Maja goltziana* d'Oliveira, 1888 (Crustacea, Decapoda, Majidae) reared in the laboratory. *Journal of Natural History* 22: 1697-1708.
- PAULA, J., 1988b. The larval and post-larval development of Pennant's swimming crab, *Portumnus latipes* (Pennant) (Brachyura, Portunidae), reared in the laboratory. *Crustaceana* 55: 202-216.
- PAULA, J., 1996. A key and bibliography for the identification of zoeal stages of brachyuran crabs (Crustacea, Decapoda, Brachyura) from the Atlantic coast of Europe. *Journal of Plankton Research* 18: 17-27.
- PESSANI, D. & SALTON, L., 1998. Planktonic larval stages of Brachyura in the Gulf of Tigullio (Ligurian Sea, Italy). *Invertebrate Reproduction and Development* 33: 201-208.
- PESSANI, D., BURRI, R. & SALTON, L., 1998. A key for the identification of the known larval stages of the Mediterranean Brachyura. *Invertebrate Reproduction and Development* 33: 191-199.
- RICE, A. L., 1982. The megalopa stage of *Latreillia elegans* Roux (Decapoda, Brachyura, Homolidea). *Crustaceana* 43: 205-210.
- RICE, A. L. & INGLE, R. W., 1975. The larval development of *Carcinus maenas* (L.) and *C. mediterraneus* Czerniavsky (Crustacea, Brachyura, Portunidae) reared in the laboratory. *Bulletin of the British Museum (Natural History): Zoology* 28: 101-119.
- RICE, A. L., INGLE, R. W. & ALLEN, E., 1970. The larval development of the sponge crab, *Dromia personata* (L.) (Crustacea, Decapoda, Dromiidea), reared in the laboratory. *Vie et Milieu* 21: 223-240.
- RODRÍGUEZ, A., 1997. Larval and postlarval development of *Pisa tetraodon* (Pennant, 1777) (Decapoda: Majidae) reared in the laboratory. *Journal of Plankton Research* 19: 29-41.
- RODRÍGUEZ, A. & MARTIN, J. W., 1997. The larval development of the crab *Xantho poressa* (Decapoda, Xanthidae) reared in the laboratory. *Journal of Crustacean Biology* 17: 98-110.
- RODRÍGUEZ, A., GONZÁLES-GORDILLO, J. I. & CUESTA, J. A., 1992. Larval stages of *Brachynotus atlanticus* Forest, 1957 (Crustacea: Decapoda: Grapsidae) reared under laboratory conditions. *Journal of Plankton Research* 14: 867-883.
- ROFF, J. C., DAVIDSON, K. G., POHLE, G. & DADSWELL, M. J., 1984. Guide to the marine flora and fauna of the Bay of Fundy and Scotian Shelf: Larval Decapoda: Brachyura. Canadian Technical Report of Fisheries and Aquatic Sciences 1322: 1-57.
- ROSSIGNOL, M., 1957. Crustacés décapodes marines de la région de Pointe-Noire. In Collignon, J., M. Rossignol & C. Roux (eds), *Mollusques, Crustacés, Poissons Marins des Côtes d'AEF en Collection au Centre d'Océanographie de l'Institut d'Études Centrafricaines de Pointe Noire*. Paris: 71-136.
- SALMAN, S. D., 1981. Larval development of *Macropodia tenuirostris* (Leach) (Crustacea, Brachyura, Majidae), reared in the laboratory. *Journal of Natural History* 15: 931-938.
- SALMAN, S. D., 1982a. Observations on the larvae of North European crabs of the genus *Ebalia* (Brachyura, Leucosiidae). *Crustaceana* 42: 256-269.
- SALMAN, S. D., 1982b. Larval development of the spider crab *Eurynome aspera* (Pennant), reared in the laboratory, with a key to the known larvae of the subfamily Pisinae (Brachyura, Majidae). *Crustaceana* 43: 78-88.
- THIRIOT, A., 1973. Stades larvaires de Parthenopidae Méditerranéennes: *Heterocrypta maltzani* Miers et *Parthenope massena* (H. Milne-Edwards). *Cahiers de Biologie Marine* 14: 111-134.
- UDEKEM D'ACCOZ, C. D', 1999. Inventaire et distribution des crustacés décapodes de l'Atlantique nord-oriental, de la Méditerranée et des eaux continentales adjacentes au nord de 25° N. *Patrimoines Naturels (Muséum National d'Histoire Naturelle / Service du Patrimoine Naturel)* 40: 1-383.
- WEAR, R. G. & FIELDER, D. R., 1985. The marine fauna of New Zealand: larvae of the Brachyura (Crustacea, Decapoda). *New Zealand Oceanographic Institute Memoirs* 92: 1-90.
- WILLIAMSON, D. I., 1982. The larval characters of *Dorhynchus thomsoni* Thomson (Crustacea, Brachyura, Majoidea) and their evolution. *Journal of Natural History* 16: 727-744.
- YATSUZUKA, K. & SAKAI, K., 1980. The larvae and juvenile crabs of Japanese Portunidae (Crustacea, Brachyura). I. *Portunus pelagicus* (Linné). *Reports of the Usa Marine Biological Institute* 2: 25-41.