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Mycotoxins occurrence in food commodities, their associated hazards and control strategies

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ABSTRACT: Globally, the food is contaminated by various means, but microorganisms are predominant factor in contaminating the food and agriculture commodities. Among microorganisms, fungi are mainly involved in the spoilage of food due to their diversified nature and minimal requirement for growth. The toxigenic fungi associated with mycotoxins, can grow during any stage of food chain including harvesting, handling, distribution and storage. Mycotoxins are fungal secondary metabolites and their production is influenced by various factors such as environmental conditions, crop type and storage conditions. Mycotoxins in agriculture commodities expose serious health hazards. This review entails different types of mycotoxins involved in the spoilage of food and agriculture commodities, their potential health hazard, maximum allowable limits of mycotoxins in different food commodities and possible control strategies. In developing countries, regulatory authorities need to establish quality control strategies and limits of mycotoxins in food, in order to ensure the consumer safety.

Keywords: mycotoxins, secondary metabolites, food spoilage, aflatoxins, maximum allowable limits

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INTRODUCTION

Food contamination is a problem since ages and fungal contamination has been identified as main cause of food spoilage (Ashiq, 2015). Since ancient times, spoilage of crops and other food commodities have shown the presence of fungi and molds (Umesha et al., 2017). There is a high chance of fungal contamination if agricultural commodities are not stored properly and are exposed to high moisture content. Once agricultural commodities are infected by fungi, the infection spreads through the different stages i.e. harvesting, processing, and storage due to favorable conditions for the fungal growth (da Rocha et al., 2014).

The population of the world is expected to reach 8.2 billion by 2030 and 842 million people were estimated undernourished during 2011-2013. In coming decades, the food supply chains will be associated with growing challenges related to urbanization, family size, population aging and consumer concerns for healthy and sustainable food production. These trends will have a significance influence on food supply which will need to be more efficient in order to meet the demands. As a result, the food supply will need to grow within the domain of available natural resources and technologies (FAO, Food and Agriculture Organization of the United Nations, 2014).

The word “mycotoxin” means a poisonous substance that is produced by fungi. Food contamination by mycotoxins is a major food safety threat and possess several health risks to the users depending on the specific type of mycotoxin consumption, exposure level and a person’s health status (Reddy et al., 2010). Mycotoxins are natural contaminants and present a serious challenge due to their diversified nature in terms of chemical structure and symptoms in humans and animals (Zychowski et al., 2013). The predominant effects of mycotoxins include carcinogenicity and neurotoxicity (Kolpin et al., 2014). The presence of mycotoxins not only poses a threat to human and animal health but it also accounts for significant economic losses, these losses can arise at any step during the food supply chain from farm to fork level (Rodrigues et al., 2011). The prevention can be best solution to overcome the mycotoxin contamination due to diversified nature of mycotoxins to contaminate wide variety of food products along the supply chain and difficulties associated with its detection (Anater et al., 2016).

Generally, most of the mycotoxins are low mo-

lecular weight secondary metabolites that apparently have no function in the metabolism of fungi (da Rocha et al., 2014). Mycotoxin chemical structure ranges mostly from simple C₄ compounds to complex compounds (Paterson and Lima, 2010). Many species of fungi produce mycotoxins and few mycotoxins have been reported to exhibit carcinogenic potential in humans and animals (Huffman et al., 2010). Among mycotoxin producing fungi, *Fusarium*, *Aspergillus*, and *Penicillium* species are main mycotoxin producers and are called field fungi due to their ability to contaminate various food commodities (Jajić et al., 2019; Ashiq, 2015). Mycotoxins including aflatoxins (AFs), fumonisins (FMN), ochratoxin A (OTA), trichothecenes (include deoxynivalenol (DON) and T-2 toxin), and zearalenone (ZEN) gained more awareness because of their high frequency of occurrence and adverse health effects to humans and animals (Bhat et al., 2010). The consumption of mycotoxin contaminated food can result in carcinogenic, immunosuppressive, and teratogenic effects (Binder et al., 2007). The mycotoxins mainly target kidneys, liver, immune and nervous systems and in humans’ general manifestations of mycotoxicosis are diarrhea, gastrointestinal distress and vomiting (Bhat et al., 2010).

The ingestion, inhalation and absorption of mycotoxins can cause mortality in humans and animals (Bankole and Adebajo, 2003). The term mycotoxicosis associated with the ingestion of mycotoxin contaminated food by animals or humans (Binder et al., 2007). The mycotoxicosis can be experienced through indirect exposure to products of animals (meat or milk) which are contaminated with mycotoxin (Bankole and Adebajo, 2003).

The foods contamination by mycotoxins can be avoided by maintaining the higher quality of food during the entire food supply chain. The high-income developed countries have less exposure to high mycotoxins level due their food safety standards and regulations (Ashiq, 2015). In hot and humid areas of the world, food spoilage thorough fungi are more common (Sabahat et al., 2010; Thompson and Henke, 2000). The developing countries with high temperature and relative humidity need to adopt the modern food safety standards and regulations in the entire food supply chain to minimize the fungal and mycotoxin food contamination. The present review summarizes different types of food contaminating mycotoxins, their influence on human and animal health and possible control strategies.

MYCOTOXINS

The word mycotoxin is derived from Greek words, i.e. mykes indicates fungi or molds and toxicum means “poison”. In 1960s the word “mycotoxin” was first used to explain the toxin in animal feed related to contaminated peanuts and the death of turkeys in England called Turkey-X-disease (Bennett, 1987). Normally the word ‘mycotoxin’ is used for relatively low molecular weight toxic chemicals (Mw<500 Da) (da Rocha et al. 2014). Generally, the effects of mycotoxins on humans and animals vary with change in their molecular structures (Miller, 1995).

FACTORS AFFECTING MYCOTOXINS PRODUCTION

Fungi are dependent on oxygen due to their aerobic nature; therefore, fungi have to face the consequences of oxygenic conditions such as presence of reactive oxygen species. The reactive oxygen species are produced during metabolic processes and their production can be influenced by environmental stress (Halliwell and Gutteridge, 2007). The accumulation of reactive species can potentiate morphological and metabolic transitions in fungi which in turn can result in toxin synthesis (Reverberi et al., 2010).

Miller (2001) reported that secondary metabolites are produced from one of the primary metabolites due to limitation of one or more nutrient. Proline, asparagine and tryptophan can increase the biosynthesis of AFs in *A. parasiticus* (Reverberi et al., 2010) however, their presence can reduce the production of AFs in *A. flavus* (Wilkinson et al., 2007). Temperature, pH, water activity and various other environmental factors significantly affect the production of mycotoxins such as OTA and AFs (Chein et al., 2019a). The environment-based factors influence the mycotoxin synthesis at transcription level and even the exposure of suboptimal quantities of fungicides can potentiate the biosynthesis of mycotoxins (Schmidt-Heydt et al., 2007).

The nature and production quantity of mycotoxins are mainly influenced by synergies of various factors: available nutrition, temperature, types of substrate, moisture content conditions, humidity, colony maturity, co-occurrence of mycotoxins with other fungi, and competing with other microbes and stress factors (Rao, 2001). The major contributing factors in the production of mycotoxins and consequences of their consumption are summarized in Figure 1.

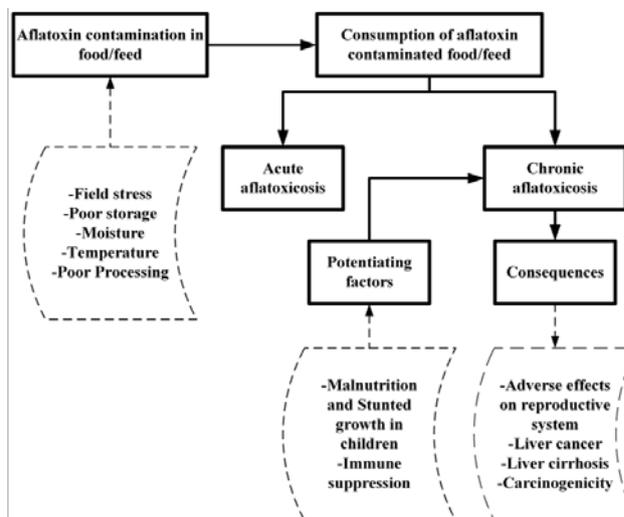


Figure 1. Contributing factors for mycotoxins contamination and consequences of their consumption (adopted from Bbosae et al. 2013)

MYCOTOXIGENIC FUNGI

Several fungal genera produce mycotoxins including *Aspergillus*, *Fusarium*, *Penicillium*, *Alternaria* and *Claviceps* spp. (Majeed et al., 2021; Reddy et al., 2010). These different mycotoxigenic fungi genera are most abundant and have a strong environmental relation with human food materials (Samsudin and Abdullah, 2013). *Fusarium* spp is reported as mycotoxigenic fungi in cereal crops as well as other food commodities. *Aspergillus* spp and *Penicillium* spp are most common pathogens for plants and food commodities during drying and storage periods (Mohale et al., 2013).

TYPES OF MYCOTOXINS

AFLATOXINS

Aflatoxins (AFs) are well-known mycotoxins and in 1960s, for the first time reported in the UK when turkey poults 100, 000 suffered and died (known Turkey X disease) after consuming AFs contaminated peanut meal (CAST, 1989). AFs are derivatives of difuranocoumarin produced by species of *A. flavus* and *A. parasiticus* via polyketide mechanism (Ellakany et al., 2018; Turner et al., 2009). Among the 18 AFs categories, (AFB1 and AFB2) B series, (AFG1 and AFG2) G series and (AFM1 and AFM2) M series have been identified as the most important AFs affecting humans and animals (Figure 2). B and G series of AFs are characterized on the basis of their fluorescence under ultraviolet light (B = blue, G = green) (da Rocha et al. 2014). M series is associated with B series hydroxylated derivatives, reported in dairy

cattle, milk, meat and various mammals which consumed contaminated food and feed with AFs (Acaroz et al., 2019a; Chen et al., 2005). AFB1 and AFG1 dihydroxy derivatives are AFB2 and AFG2, respectively (Chunet al., 2007). The main reason that AFB1 and AFG1 are generally more toxic and carcinogenic than AFB2 and AFG2 is the presence of a double bond in the form of vinyl ether in their terminal furan ring, which is the active site and intensifies their fluorescence. Thus, their active site can experience a reaction of reduction leading to a change in activity (Turner et al., 2009). AFs are stable at constant high temperatures with minimum loss through cooking or pasteurization. AFs are unstable with UV, intense pH (< 3 or >10) values and oxidizing agents in the presence of oxygen (Herzallah, 2009).

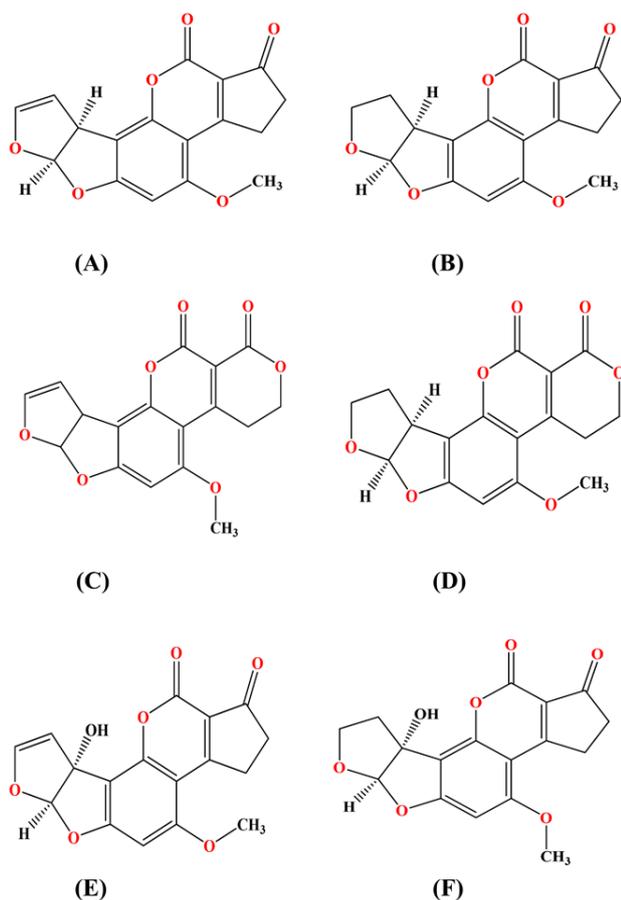


Figure 2. Chemical structure of (A) Aflatoxin B1 (B) Aflatoxin B2 (C) Aflatoxin G1 (D) Aflatoxin G2 (E) Aflatoxin M1 (F) Aflatoxin M2

A. flavus only produces B AFs (B1 and B2) and around 40% of *A. flavus* species produce AFs. *A. parasiticus* produces G AFs (G1 and G2) (Umesha et al., 2017). Morphologically *A. nomius* is like *A. flavus* but it produces both B and G AFs. *A. flavus* commonly

occurs in crops in the world's warm temperature regions (Pepeljnjak et al., 2004).

AFs are mainly produced by *A. Parasiticus* and *A. flavus*, the optimum growth temperature range are 25 to 35 °C and 28 to 30 °C, respectively (Bhat et al., 2010). AFs are of main concern in hot and humid areas, as the optimum temperature in warm areas of the world are favorable for fungal growth (Fernandez-Cruz et al., 2010). AFs are typically found in agriculture products like cereals (barley, sorghum, wheat, maize and rice), spices (ginger, turmeric, coriander, black pepper, and chili), tree nuts (nuts, walnuts, peanuts, pistachios and almonds) and oilseeds (cotton soybean, sunflower and sesame) (Acaroz et al., 2019b; Firdous et al., 2012). Whereas AFM1 and AFM2 are mainly present in milk, milk products and meat (da Rocha et al., 2014).

Health Hazards of Aflatoxin

In tropical and subtropical regions, including Africa and Asia, various studies have been reported that indicated the adverse effects of AFs food contamination (Acaroz et al., 2019a, 2019b). The ingestion of food heavily contaminated with AFs can cause death in various cases (Ashiq, 2015). A wide variety of animals are affected by AFs including rodents, cattle, poultry fish and swine. Though the AFs response depends on the level of exposure, exposure duration, nutrition status, health, age, and environmental factors (Wagacha and Muthomi, 2008).

AFs are carcinogenic, teratogenic, hepatotoxic, mutagens, immunosuppressants and can induce various other serious hazards in animals and humans (da Rocha et al., 2014; Eaton and Gallagher, 1994). AFs are categorized as group 1 carcinogens by International Agency for Research on Cancer (IARC, 1993). AFs interfere with the protein synthesis due to their DNA binding capacity, hence, effect various essential cellular metabolisms and immune system (da Rocha et al., 2014). Generally, aflatoxicoses is recognized as disease associated with the AFs consumption. The death can be encountered in acute aflatoxicoses, whereas the chronic conditions can induce cancer, immunosuppression and hepatotoxicity (Zain, 2011).

FUMONISINS (FMNS)

FMNs were first discovered and reported in 1988 (Bennett and Klich, 2003). FMNs are mainly produced by *Fusarium* genera (*Fusarium proliferatum*, *F. verticillioides*, and *F. nygamai*) and 28 different

types of FMNs have been isolated and classified in four groups (A, B, C and P). The different species of *Fusarium* genera produce FMNs, particularly *F. verticillioides* formerly *Fusarium moniliforme*, *F. proliferatum*, *F. anthophilum*, as well as *Alternaria alternata* (Omurtag 2008). *Aspergillus niger* has been reported to produce FMNs like B2 and B4, and a new B series of FMNs (FB6) was recognized from *A. niger* (Huffman et al., 2010).

The major types of FMNs (FB1, FB2, and FB3) are contaminants of natural cereals (Omurtag, 2008). Among FMNs, the most important and abundant mycotoxin family member is FB1 (Figure 3) (Reddy et al., 2010). FMNs are polyketide metabolites derived from repetitive condensation of acetate units or other short carboxylic acids by a similar mechanism of enzyme bound for fatty acid synthesis (Huffman et al., 2010).

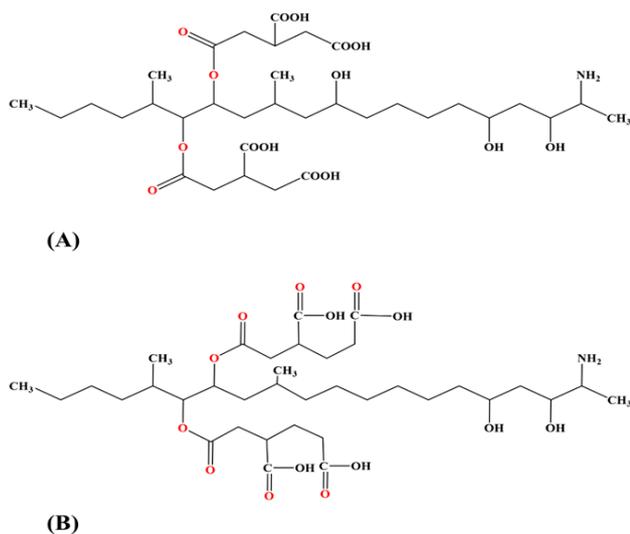


Figure 3. Chemical structure of (A) Fumonisin B1 (B) Fumonisin B2

Temperature and humidity are important factors for *Fusarium* contamination and synthesis of mycotoxins (Omurtag, 2008). The optimum temperature and water activity for the production of FMNs were reported in the range of 15-30 °C and 0.9-0.995, respectively (Sanchis and Magan, 2004). The presence of FMNs was found in agriculture and food commodities including bovine milk, dried figs, corn, products of corn, medicinal plants, and herbal tea (Omurtag et al., 2010). FMNs were observed as common contaminant of feeds and food in Philippines, South America, China, USA, Africa, France, Italy, Indonesia, and Thailand (Kumar et al., 2008).

Health Hazards of Fumonisin

Human intake of FMNs contaminated foods has been associated with esophageal cancer in Asia, South Africa, and Central America, (Alizadeh et al., 2012; Marasas et al., 2004). FMNs are categorized as a group 2B substance (human carcinogenic) by International Agency for Research on Cancer (IARC, 1993) and immunosuppressive by World Health Organization (WHO, 2002). The chronic FMNs effect on animals include impairment of basic immune function, nephrotoxicity, hepatotoxicity, respiratory disorders, and reduced milk production (Diaz et al., 2000). FMNs do not interact with DNA like AFs, however, due to similarity with sphingosine it might intervene with the biosynthesis of sphingolipids (Shier, 1992), which in turn influence the essential cellular activities as sphingolipids are essential for membranes, inter and intra cellular communication (Merrillet al., 1993).

OCHRATOXINS (OTS)

In South Africa, Ochratoxin A (OTA) was first reported in 1965 (Van der Merwe et al., 1965) and isolated from cornmeal contaminated by *Aspergillus ochraceus*. Later in 1969, ochratoxin (OT) was isolated and reported in the United States from corn (Shotwell et al., 1969). OTA was later identified as secondary metabolite associated with *Aspergillus* and *Penicillium* spp (Duarte et al., 2010). Historical reports revealed that OTA was found in Egyptian tombs and considered for the suspicious deaths of many architects (Pittet, 1998).

OTA contains 7-carboxy-5-chloro-8-hydroxy-3,4 dihydro- (3R) -methylisocoumarin in a carboxyl group linked to L-β-phenylalanine (Figure 4) (Fernandez-Cruz et al., 2010). OTA is a crystalline white powder that is stable in food processes but unstable in the presence of light. Acid hydrolysis of OTA changes it to phenylalanine and an optically active lactone acid named OTα (IARC, 1993).

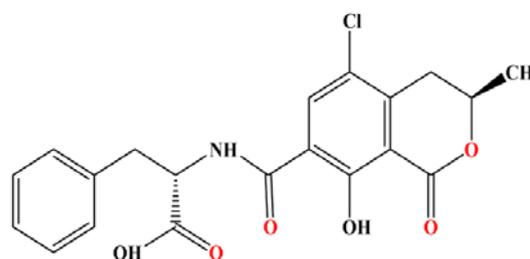


Figure 4. Chemical structure of Ochratoxin A

OTA is produced by various species of *Aspergillus* and *Penicillium* genera. Main producers are *A.*

carbonarius, and *A. ochraceus*, (Bachaet al., 2009). Some species of *Aspergillus* (*A. niger*, *A. carbonarius*, *A. ochraceus*, *P. verrucosum*) are responsible for OTA production (Bhat et al., 2010). OTA is a natural contaminant in various foods, like cocoa, corn flour, cereals, dried fruits, maize, soya beans, peanuts, nuts, fish, milk, eggs, poultry, kidney beans, tea, and some herbs (Batista et al., 2009). Hence, in tropical areas, it is however linked to moldy green coffee beans. It is also found in a coffee brew and roasted coffee beans (Sibanda et al., 2002). OTA contaminates spices, and dried fruit, whereas, grapes are commonly contaminated with OTA during storage (Bhat et al., 2010). Exposure of OTA to human generally occurs by the intake of poorly stored food products. The presence of OTA has been observed in the tissues and organs of animals and humans, including blood, breast milk, and meat (Kumar et al., 2008). The optimum temperature and water activity to produce OTA are 25-30 °C and 0.98, respectively (Milani, 2013).

Health Hazards of OTA

OTA has been associated with carcinogenic, nephrotoxic, teratogenic, and immunosuppressive effects in humans and animals (da Rocha et al., 2014). OTA was categorized as carcinogen to humans (group 2B) by the International Agency for Research on Cancer (IARC, 1993). OTA was linked with Balkan endemic nephropathy (BEN), a disease of kidney which was observed in certain areas of Balkan countries (Pfohl-Leschkowicz et al., 2002). The previous reports reported OTA as a potent teratogen, immunosuppressive, liver toxin, and carcinogen in animals (Pfohl-Leschkowicz and Manderville, 2007).

TRICHOHECENES (TCT)

In 1949, Trichothecin from *Trichothecium roseum* was isolated and defined for the first time by Freeman and Morrison. Trichothecin discovery was associated with other TCT for example, T-2 toxin (T-2), and deoxynivalenol (DON) (Omurtag, 2008). TCTs are chemically defined by a tetracyclic sesquiterpenoid 12, 13-epoxytrichothec-9-ene ring system (Zöllner and Mayer-Helm, 2006). TCTs were further categorized as macrocyclic, or non-macrocyclic, depending on macrocyclic presence of ester or an ester-ether bridge between C-4 and C-15 (Merhej et al., 2011).

TCT is a family of mycotoxins that includes more than 200 compounds divided into four subclasses (Group A-D), based on their functional characteristic. The most toxic TCTs are Group A (Bhat et al., 2010).

Generally, TCT are found as contaminants in cereals and their derivative (Foroud and Eudes, 2009). TCT found in food/feedstuffs are produced by *Fusarium graminearum* and *F.culmorum*. *F.pseudograminearum*, *F. graminearum*, and *F. culmorum* are accountable of producing deoxynivalenol (DON) toxins (Figure 5) which is a member of TCT (Ashiq, 2015; Glenn, 2007). TCTs are leading source of contamination in grains like oats, maize, barley and wheat (Zöllner and Mayer-Helm, 2006). TCT have also been observed in cereal products and milk (Spanjer et al., 2008). *F. culmorum*, *F. sporotrichioides* and *F. graminearum*, were reported to produce DON toxins, (Merhej et al., 2011).

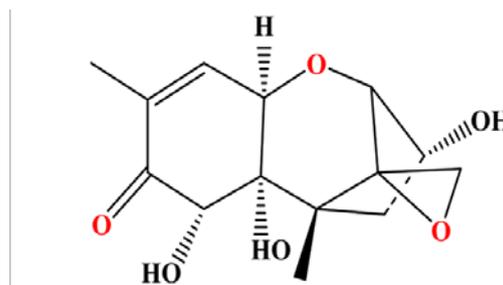


Figure 5. Chemical structure of Deoxynivalenol (DON)

Health Hazards of TCT

In humans and animals TCTs are associated with serious and chronic toxicosis. In eukaryotes contact of TCT results in slowed growth. Therefore, TCTs can affect wide range of invertebrates, mammals, fish, and plants (Wannemacher et al., 1997). TCTs can inhibit cell division, RNA and DNA synthesis, which might influence the structure and integrity of membranes, as well as function of mitochondria (Cundliffe and Davis, 1977), DON and T-2 toxins affect immunity through inhibition of cell and protein synthesis (Bhat et al. 2010). DON lowers the body's level of antibodies and immunoglobulins (Richard, 2007).

ZEARALENONE (ZEN)

ZENs are estrogenic lactone resorcylic acid compounds mainly produced by *Fusarium* species (Diekman and Green, 1992). The production of ZEN is mainly associated with *Fusarium graminearum* and *F. culmorum* (Logrieco et al., 2002). ZEN is a non-steroidal mycotoxin and referred as F-2 toxin (Zinedine et al., 2006). ZEN is biosynthesized by various species of *Fusarium* via a polyketide pathway (Huffman et al., 2010). Chemically ZEN is 3, 4, 5, 6, 9, 10-hexahydro-14, 16-dihydroxy-3-methyl-1H-2 benzoxacyclopentadecin 1, 7 (8H) - dione, is a macrocyclic β -resor-

cyclic acid lactone (Figure 6) (Cozzini and Dellaflora, 2012).

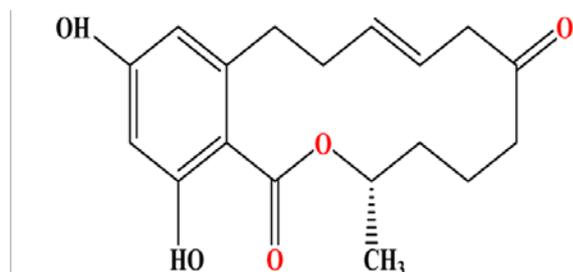


Figure 6. Chemical structure of Zearalenone

The crops that are mainly contaminated by ZENs are wheat, corn, oats, maize, barley, rice, millet, and sorghum (Zinedine et al., 2006). The presence of ZEN was reported in cereal by-products of corn and soya meal (Schollenberger et al., 2007), eggs (Sypecka et al., 2004) and milk (Seeling et al., 2005). ZENs are stable to heat, but under alkaline conditions the toxins can be degraded at a high temperature (>150 °C) (European Commission, 2000). Children are more affected by ZEN contaminated foods because of their high consuming rate of cereal based foods (Bhat et al., 2010).

Health Hazards of ZEN

ZENs may cause abortion, infertility, and problems with reproduction system (especially swine)

and are linked with cervical cancer (El-Nezamiet al., 2002). ZEN contaminated feed ingestion leads to interference with exocrine and endocrine systems. Like other environmental estrogens, ZEN is able to interfere with the function of sex steroids (Bhat et al., 2010). Metabolites of ZEN bind to receptors of estrogen and activate transcription of genes (Fink-Gremmels and Malekinejad, 2007). Because of ZEN estrogenic activity, contaminated feed with ZEN showed changes in the reproductive tract, fertility reduction, and rise in number of fetal resorptions (Morgavi and Riley, 2007). Among animals, pigs have been the most sensitive to ZEN and poultry is least affected (Bhat et al., 2010).

MYCOTOXIN REGULATIONS

Food and Agriculture Organization declared mycotoxins as the major contaminants, accountable for 25% of food crops across the world. The mycotoxin contamination presents a serious threat to food security and economy (Aiko and Mehta, 2015; Alshannaq and Yu, 2017). Maximum allowable limits of different types of mycotoxins in food and feed have been specified by various national and international monitoring organizations. The US Food and Drug Administration (FDA), World Health Organization (WHO), European Union (EU), and several other countries have set the maximum limit of the mycotoxins in food, which are summarized in Table 1.

Table 1. Maximum allowable limits of mycotoxins in food commodities

| Country/Region | Mycotoxins | Products | Maximum limit (ppb) | References |
|---|--|--|---------------------|---|
| European Union | Aflatoxins: B1 G1, B2, G2 | Maize, wheat, rice, spices, almonds, oil seeds, dried fruits, cheese | 0.1-8 | EC (European Commission 2010). |
| | | | 4-15 | |
| | Aflatoxin M1 | Milk, eggs, meat | 0.5 | CAC (Codex Alimentarius Commission 2015). |
| | Fumonisin (FB1 and FB2) | Maize-based breakfast cereals and maize-based snacks | 800 | (EC 2007). |
| | | | Raw maize grain | 4000 |
| Maize flour and maize meal | | | 2000 | (CAC 2015) |
| Maize intended for direct human consumption | | | 1000 | (EC 2007). |
| Ochratoxin A | Cereals, dry fruits, wine, spices, oat, raisins, coffee, cocoa, soybeans, meat | 0.5-15 | (EC 2006). | |
| | | Wheat, barley and rye | 5 | (CAC 2015). |

| | | | | |
|--------------|------------------------------------|---|-----------|--|
| | Deoxynivalenol (DON) | wheat, oats and maize | 1250 | (EC 2007). |
| | | Flour, meal, semolina and flakes derived from wheat, maize or barley | 1000 | (CAC 2015). |
| | | Cereal grains (wheat, maize and barley) destined for further processing. | 2000 | (CAC 2015). |
| | Zearalenone | Unprocessed cereals other than maize | 100 | (EC 2007). |
| | | Maize intended for direct human consumption, maize-based snacks and maize-based breakfast cereals | 100 | (EC 2007). |
| | | cereal flour, bran and germ for direct human consumption | 75 | (EC 2007). |
| USA | Aflatoxins B1, B2, G1, G2 | Total aflatoxins in food for human consumption corn, peanut products, cottonseed meal, maize, wheat, rice, peanut, sorghum, pistachio, almond, ground nuts, tree nuts, figs, cottonseed, spices | 20 | USDA (United States Department of Agriculture 2015). |
| | Aflatoxin M1 | Milk, milk products | 0.5 | FDA (Food and Drug Administration 2011) |
| | Total Fumonisin (FB1, FB2 and FB3) | Cereals | 2000-4000 | (Alshannaqand Yu 2017). |
| | | Corn products and cleaned maize used for popcorn | 2000-3000 | |
| | Ochratoxin A | Cereals, wheat, barley, and rye and derived products. | 5 | (CAC 2015). |
| China | Aflatoxin B1 | Corn, corn flour (grits, flake) and corn products, peanut and its products, peanut oil, corn oil | 20 | (Clever 2018) |
| | | Paddy rice, brown rice, rice. Vegetable oil and fat | 10 | |
| | | Wheat, barley, other grains, | 5.0 | |
| | | Wheat flour, cereal, other husked grains and bean products, other cooked nuts and seeds. Soy sauce, vinegar, fermented paste | | |
| | Aflatoxin M1 | Foods intended for special dietary uses. | 0.5 | |
| | | Milk and milk products | 0.5 | |
| | Ochratoxin A | Grains and grain products, beans and bean products, Baked coffee beans, ground coffee (roast coffee) | 5.0 | (Clever 2018). |
| | | Instant coffee | 10.0 | |
| | Deoxynivalenol | Corn, corn flour (grits, flake). | 1000 | (Clever 2018). |
| | | Barley, wheat, cereal, wheat flour | | |
| | Zearalenone | Wheat, wheat flour, corn, corn flour (grits, flake) | 60 | (Clever 2018). |

| | | | | |
|--|---|---|--------------|------------------------------------|
| South Korea | Aflatoxin B1 | Grains, cereal products, dried fruits, Meju, and streamed rice | 10 | (Yoshizawa 2011; Chun 2011). |
| | Aflatoxin B1 | Baby foods | 0.1 | (Chun 2011). |
| | Aflatoxin B2, G1, and G2 | Grains, cereal products, dried fruits, Meju, streamed rice, and baby foods | 15 | |
| | Aflatoxin M1 | Raw milks and milks prior to manufacturing processing | 0.5 | (Yoshizawa 2011; Chun 2011). |
| | Fumonisin B1, B2 | Grain products, Cereals processed corn products for popcorn, confectionaries (contain>50% corns). | 1000 | (Chun 2011). |
| | | Corn processed food, corn powder | 2000 | |
| | | Corn | 4000 | |
| | Deoxynivalenol | Grain and their processed foods | 1000 | (Chun 2011). |
| | | Corn and their processed foods | 20000 | |
| | | Cereals | 500 | |
| Ochratoxin A | Meju | 20 | (Chun 2011). | |
| | Instant coffee and raisins | 10 | | |
| | Red pepper powder | 7 | | |
| | Grains and their processed food (grinding, cutting, etc). Coffee beans, and roasted coffee. | 5 | | |
| | Baby foods for infants and young children | 0.5 | | |
| Zearalenone | Grains and processed grain foods | 200 | (Chun 2011). | |
| | Confectionaries | 50 | | |
| | Baby foods | 20 | | |
| Indonesia | Total aflatoxins | Corn feed | 50 | (Suparmoet al. 2011) |
| | Total aflatoxins | All foods | 35 | |
| | Aflatoxin B1 | All foods | 20 | |
| | Aflatoxin B1 | Peanut, corn and their products | 15-20 | |
| | Aflatoxin M1 | Dried milk and related products | 5 | |
| | Aflatoxin M1 | Milk, drink milk products, fermented milk and rennin hydrolyzed milk products, concentrated milk, cream, cheese, pudding, yogurt, whey and their products | 0.5 | |
| | Fumonisin B1, B2 | Corn (raw material) | 2000 | (Suparmoet al. 2011) |
| | | Corn foods products, e.g., popcorn, corn chips | 1000 | |
| | Ochratoxin A | Spices | 20 | (Suparmoet al. 2011) |
| | | Instant Coffee | 10 | |
| Cereals (rice, corn, sorghum, wheat) and their products and coffee | | 5 | | |
| Zearalenone | Maize | Not detectable | FAO (2003) | |
| Japan | Total | All foods | 10 | (Kawamura 2011; Srianujata 2011) |
| | Aflatoxin B1 | Rice | 10 | (Srianujata 2011; Yoshizawa 2011). |
| | Aflatoxin B1 | Other grains | 5 | (Srianujata 2011). |
| | Deoxynivalenol | Wheat | 1, 100 | (FAO 2003). |
| | Zearalenone | Compound feeds | 1000 | (FAO 2003). |

| | | | | |
|---------------|---------------------------|--|-----------|-------------------------|
| Brazil | Aflatoxins B1, G1, B2, G2 | Oil seeds, nuts, dried fruits, cereals, spices | 20 | (Anfossiet al. 2016). |
| | Aflatoxin M1 | Milk and infant formula | 0.5-5 | |
| | Fumonisin | Maize | 2000-5000 | (Anfossiet al. 2016). |
| | Ochratoxin A | Cereals, dried fruits, coffee, cocoa, wine, beer, grape juice, spices, liquorice, blood products | 2-30 | (Anfossiet al. 2016). |
| | Deoxynivalenol | Cereals, bakery products | 750-3000 | (Anfossiet al. 2016). |
| | Zearalenone | Cereals, bakery products, maize oil | 200-1000 | (Anfossiet al. 2016). |
| India | Aflatoxins | Oil seeds, nuts, dried fruits, cereals, spices | 30 | (Anfossiet al. 2016) |
| | Aflatoxin M1 | Milk and infant formula | 0.5 | |
| | Ochratoxin A | Cereals, dried fruits, coffee, cocoa, wine, beer, grape juice, spices, liquorice, blood products | 20 | (Anfossiet al. 2016). |
| | Deoxynivalenol | Cereals, bakery products | 1000 | (Anfossiet al. 2016). |
| Russia | Aflatoxin B1 | Maize | 5 | (Abdallah et al. 2015). |
| | Aflatoxin M1 | Milk | 0.5 | |
| | Ochratoxin A | Cereals, dried fruits, coffee, cocoa, wine, beer, grape juice, spices, liquorice, blood products | 5 | Anfossiet al. 2016). |
| | Deoxynivalenol | Cereals, bakery products | 700-1000 | Anfossiet al. 2016). |
| | Zearalenone | Cereals, bakery products, maize oil | 1000 | Zinedineet al. 2007). |

CONTROL STRATEGIES FOR MYCOTOXIN CONTAMINATION

It is difficult to control contamination of mycotoxin in the field, during harvesting, storage and transportation of food and feed commodities (Umesha et al., 2017). The factors like soil moisture, invasion mostly with insects and mineral deficiencies contribute to mycotoxin contamination (Murphy et al., 2006). The way to protect and ensure consumers safety is to prevent fungal contamination of food/feed commodities which eventually leads to mycotoxin production (Figure 7). Plantation and pre-harvesting approaches can be improved by exercising preventing measures to reduce mycotoxin contamination. The control of mycotoxins in food produce is essential for public health and can be achieved by number of strategies such as, prevention of fungal contamination in food and feed, decontamination of mycotoxins and continuous surveillance. The fungal growth in food and feedstuff can be prevented by implementation of hygiene practices, drying and storage under appropriate conditions (Tola and Kebede, 2016). The high moisture content of food produce favors the growth of fungi and mycotoxin production. The drying of food produce can reduce the moisture content and hence prevents the growth of fungi and mycotoxins (Cheinet al., 2019a). The mold growth and accumulation of mycotoxins in

food can be prevented by using natural preservatives such as acetic acid, lactic acid, benzoic acid and various essential oils (Chein et al., 2019b; Sriwattanachai et al., 2018).

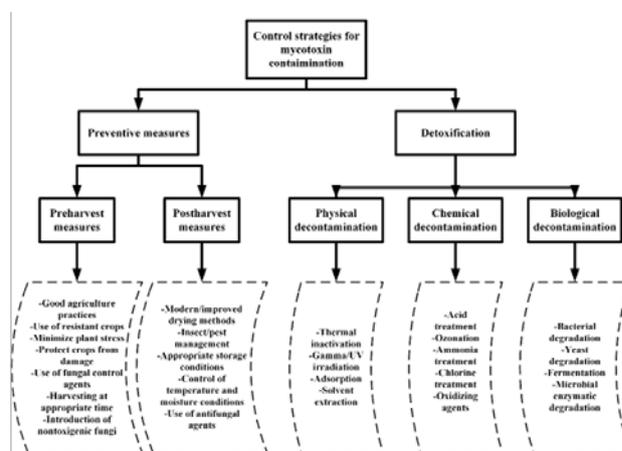


Figure 7. Aflatoxin prevention and control strategies in food and feed commodities. (Adopted from Ashiq, 2015)

Prevention of mycotoxin contamination may not always be possible, so decontamination process is important (Aiko and Mehta, 2015; Rustom, 1997). The mycotoxins can be decontaminated by physical and chemical treatments. Irradiation, cooking, boiling and extrusion are categorized as physical treatments

for decontamination of mycotoxins. During food processing, food undergoes heat treatment that might result in thermal inactivation of mycotoxins. However, most of the mycotoxins are heat stable and may not be easily inactivated by heat processing (Bullerman and Bianchini, 2007). The thermal deactivation of mycotoxins is influenced by certain essential factors such as temperature, water content and duration of exposure to heat. Mycotoxins can be decontaminated by chemical treatment however, the resultant degradation products might influence the food quality and safety (Aiko and Mehta, 2015).

BIODEGRADATION OF MYCOTOXINS

In comparison to other degradation approaches, biodegradation provides a better chance of deactivation of mycotoxin. Certain microbes and enzymes may reduce secondary metabolites toxicity such as AFs, FMNs, OTAs, TCTs, and ZENs can be converted into less toxic metabolites through changes in the structure of these mycotoxins (Pintonet et al., 2010). Biodegradation has been widely used in many countries to detoxify mycotoxins. Cleavage of ring, acetylation,

hydrolysis, glycosylation, deamination, and decarboxylation are the key procedures for biotransformation reactions (Guan et al., 2008). Biodegradation typically begins with the mycotoxin's identification through high performance liquid chromatography or enzyme linked immunosorbent test following by incubation media with specific microbes (Ding et al., 2015). In addition, many bacterial species can detoxify mycotoxins by biotransformation mechanism (Dalié et al., 2010; Tokai et al., 2005). The degradation of mycotoxins by different microorganisms are summarized in table 2. Lactic acid bacteria (LAB) were reported to inhibit the accumulation of mycotoxins and it was found that the inhibition of mycotoxin accumulation was not dependent on low pH rather it was associated with production of low molecular weight LAB metabolites (Dalié et al., 2010).

Earlier studies focused on mycotoxin toxicity and biodegradation mechanisms. The mechanism of microbial degradation of mycotoxins involves the use of microbial catabolic pathways, which results in fewer toxic effects or harmless end products. (Yang et al., 2014).

Table 2. Biodegradation of mycotoxins by microbes

| Mycotoxin | Microbes | Biodegradable products | Mechanism | References |
|----------------|--|--|--|--|
| Aflatoxin | <i>Nocardia corynebacteroides</i> , <i>Corynebacterium rubrum</i> , <i>Pseudomonas putida</i> , <i>Rhodococcus spp.</i> , and <i>Saccharomyces Cerevisiae</i> | Aflatoxicol; Aflatoxin M1; Aflatoxin B2a | Act on lactone ring | (Adeboet al. 2015; Du et al. 2017). |
| Fumonisin B1 | <i>Exophialaspiniifera</i> , <i>Sphingopyxis</i> spp, <i>Sphingomonas spp.</i> | Fumonisin hydrolyzed B1 (HFB1) | Removes tricarballylate groups with carboxylesterases in C-14 and C-15 | (Du et al. 2017; Vanhoutteet al. 2016) |
| Ochratoxin A | <i>Bacillus licheniformis</i> , <i>Bacillus spp.</i> , <i>Brevibacteriumiodinum</i> , <i>Acinetobacter calcoaceticus</i> , <i>Brevibacterium epidermidis</i> , <i>Lactobacillus acidophilus</i> , <i>Aspergillus versicolor</i> , <i>Aspergillus niger</i> , <i>Aspergillus ochraceus</i> , <i>Rhodoturulaspp.</i> , <i>Saccharomyces spp.</i> | Phenylalanine; Ochratoxin A | Hydrolyze the amide bond | (Du et al. 2017; Vanhoutteet al. 2016) |
| Trichothecenes | <i>Blastobotrys capitulate</i> , <i>Trichomonascus</i> , <i>Aspergillus</i> , <i>Curtobacterium</i> spp, <i>Anaerovibriolipolytica</i> , <i>Selenomonas</i> and <i>Saccharomyces</i> | 3-acetyl T-2 toxin; T-2 toxin 3-glucoside; Neosolaniol | Acetylation deacetylation, deep oxidation, oxygenation, Epimerization, and glucosylation | (Du et al. 2017; Vanhoutteet al. 2016) |
| Zearalenone | <i>Mucor bainieri</i> , <i>Rhizopus spp.</i> , <i>Cunninghamellabainieri</i> , <i>Alternaria alternate</i> , <i>Thamidium elegans</i> , <i>Aspergillus ochraceous</i> , <i>Rhodococcus spp.</i> , <i>Streptomyces rimosus</i> , <i>Trichosporonmycotoxinivorans</i> , <i>Pseudomonas spp.</i> , <i>Aspergillus niger</i> and <i>Acinetobacter spp</i> | (α -ZEL) and β -zearalenol (β -ZEL) ; ZOM-1; 2, 4- dimethoxyl zearalenone; zearalenone- 4- β -D-glucoside | Cleavage of the lactone ring and change of the hydroxyl groupC-4 | (Du et al. 2017; Vanhoutteet al. 2016) |

CONCLUSION

The fungal contamination and mycotoxin accumulation account for major food spoilage in the world and can greatly influence the world economy. The presence of mycotoxins in food or feedstuff not only accounts for food spoilage but poses a serious health hazard to both humans and animals. Therefore, the control of mycotoxins in food and feed is essential to ensure the food safety and food security. Mycotoxins accumulation in food can be controlled by implementation of good agriculture practices. The consumption of mycotoxin contaminated food can lead to serious health hazards. Mycotoxins can be eliminated from food or feedstuff by physical and chemical decontamination. The complete elimination of mycotoxins from food and feed commodities is a difficult task and requires a combined effort from policy makers,

government agencies, farmers, processors and distributors. However, the developing countries should opt for the modern strategies for minimizing the mycotoxin contamination to control their level in food commodities within the maximum allowable limits. The maximum allowable limits for mycotoxins, their control strategies should be communicated with farmers and processors.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

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