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Implications of supplemental microbial phytase and essential oils (*Thymus vulgaris L.* and *Mentha pulegium L.*) on carcass traits, meat organoleptic and antioxidant status in broiler chickens

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ABSTRACT: This experiment was carried out to study the influence of microbial phytase and herbal essential oils (*Thymus vulgaris L* and *Mentha pulegium L.*) on carcass traits, meat organoleptic and antioxidant status in broilers. Three hundred and eighty-four male broilers (Ross -308) were used in factorial arrangements with eight treatments includes (0 and 200 mg/kg thyme essential oil), (0 and 200 mg/kg *Mentha* essential oil), (0 and 500 IU/kg microbial phytase) and 4 replicates (12 chicks per replicate) according to a completely randomized design and in three experimental periods include: starter 1 to 10 days, grower 11 to 24 days and finisher 25 to 42 days. Regarding the effects of experimental treatments on carcass traits, the use of thyme and mentha essential oils increased the percentage of abdominal fat ($P<0.05$). The use of thyme essential oil significantly increased the levels of blood superoxide dismutase (SOD) and glutathione peroxidase (GPX) ($P<0.05$). Regarding the effects of experimental treatments on meat organoleptic, the use of thyme essential oil increased the overall acceptance of chicken meat ($P<0.05$). In conclusion, the use of essential oils, although it leads to an increase in antioxidant parameters, also leads to an improvement in meat quality.

Keywords: Broiler, essential oils, antioxidant status and organoleptic.

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INTRODUCTION

Due to the legal prohibition on the use of antibiotics as an additive has increased the tendency to use natural compounds derived from aromatic plants in food and animal nutrition (Abd El-Ghany and Ismail, 2014; Besharati et al., 2020; Cimrin et al., 2020). Essential oils can show potentially beneficial properties such as: antiviral effects (Brochot et al., 2017), antimicrobial (Adaszyska-Skwirzyńska and Szczerbińska, 2018; Sabo and Knezevic, 2019; Sevim et al., 2020; Abd El-Ghany, 2020), antioxidant (Chowdhury et al., 2018; Pirgozliev et al., 2019), digestibility enhancement (Zhai et al., 2018; Yang et al., 2019) and anti-parasitic (Sabuna et al., 2019; Sidiropoulou et al., 2020). Nowadays, herbal products as extracts, powder and juices are being much investigated and used by researchers in the field of food sciences and animal nutrition. This makes the world to further investigate such type of natural products for an increase in production efficiency in poultry business. Different types of phytogenic products such as mentha, garlic, anise, cinnamon, coriander, oregano, chili, pepper, rosemary, rosehip and thyme can be used as a growth promotor and gut health modulator in poultry (Criste et al., 2017). Thyme (*Thymus vulgaris*) and pennyroyal (*Mentha pulegium*) are plants of the mint family. Thyme (*Thymus vulgaris*) active constituents are thymol and carvacrol, which can affect animal metabolism and physiology. The active substances in thyme can stimulate secretion of pancreatic enzymes, thereby improving digestion efficiency and absorption of protein and other nutrients from the bird's gastrointestinal tract (Bouhitt et al., 2019). Pennyroyal (*Mentha pulegium*) is a perennial, aromatic, popular and herbaceous plant, which can reach up to half a meter in height. It has been used as an infusion, herbal tea or powder for the treatment of some diseases such as bronchitis, whooping cough, the common cold, sore throat and digestive disorders (Gülçin et al., 2020). Grigors et al. (2010) reported that the antioxidant capacity of thyme essential oil was equivalent to that of ascorbic acid. The antioxidant capacity of thyme essential oils is attributed to the thymol phenolic compounds, carvacrol and thymohydroquinone (Tabari et al., 2017; Valdivieso-Ugarte et al., 2019; Ahmadi and Jafarizadeh-Malmiri, 2020). The positive effects of phytase in poultry are documented (Farhadi et al., 2017; Roofchaei et al., 2019) and microbial phytase supplementation has been shown to increase crude protein and amino acids digestibility (Baghban-Kanani et al., 2018).

Feed, as one of the influential parts, can affect the quality of poultry meat. Factors that affect meat quality are intricate and occur entire the production chain (Vašková et al., 2015; Semjon et al., 2020). Supplying poultry meat of sufficient quantity and quality to meet communal requirement is a primary goal of the broiler industry (Yadav and Jha, 2019). From a consumer perspective, poultry is a very yummy and essential element in the human diet due to its nutritional, dietetic, and sensory attributes, and its rapid culinary preparation (Vašková et al., 2018). Therefore, the objective of this study was to investigate the effects of supplemental microbial phytase and essential oils (*Thymus vulgaris* L. and *Mentha pulegium* L.) on carcass traits, meat organoleptic and antioxidant status in broilers.

MATERIALS AND METHODS

Animals and feed

The research animal ethics committee of Islamic Azad University approved this experimental protocol (no. 2019/03-384). A total of 384 male broilers (Ross-308) were used in factorial arrangements with eight treatments includes (0 and 200 mg/kg thyme essential oil), (0 and 200 mg/kg *Mentha* essential oil), (0 and 500 IU/kg microbial phytase) and 4 replicates (12 chicks per replicate) according to a completely randomized design and in three experimental periods include starter 1 to 10 days, grower 11 to 24 days and finisher 25 to 42 days. The diets were formulated using the UFFDA software program to provide the advised levels of nutrients as specified for the Ross-308 broiler. Thyme and mentha essential oils were prepared from Barij Essential Oil Company (Kashan, Iran). Meri-Phyze 5000 is a microbial phytase manufactured by Meriden Animal Health in the United Kingdom. Each gram of Meri-Phyze 5000 contains at least 5000 units of phytase (FTU) enzyme.

Carcass quality

At the end of the experimental period, on the 42nd day, two birds per replicate were selected and were slaughtered to determine carcass traits. The percent of the carcass was determined by comparing spleen weight, abdominal fat, intestine, gizzard, liver, bursa of Fabricius, breast, thigh, back, and neck weights to corresponding carcass weights. To determine the organoleptic properties, including meat color, they were randomly measured in four positions on the surface of each half of the pectoralis muscle (right half), and the mean values of color per muscle were calculated according to Zhuang and Savage (2009) method by the

Table 1. Nutritional contents of diets fed to broilers during the starter, growing and finishing periods

	Starter period (1 to 10 days of age)	Growing period (11 to 24 days of age)	Finishing period (25 to 42 days of age)
Corn	47.75	54.93	56.75
Soybean meal	44.50	37.90	36.71
oil	3.26	3.39	3.05
powder of bone	2.26	2.18	1.87
Oyster powder	0.29	0.29	0.33
Salt	0.46	0.47	0.42
Vitamin supplement *	0.25	0.25	0.25
Mineral supplement *	0.25	0.25	0.25
DL-methionine	0.37	0.34	0.38
Metabolic energy (kcal / kg)	3000	3000	3000
Crude protein (%)	27.50	20.95	20.64
Calcium (%)	1.05	0.88	0.80
Usable phosphorus (%)	0.50	0.43	0.39
Sodium (%)	0.32	0.21	0.19
Lysine (%)	1.67	1.26	1.23
Methionine + cysteine (%)	1.18	0.99	0.85
Trepidophan (%)	0.35	0.28	0.28

*Provides per kg of diet:trans-retinol 3784 mg, cholecalciferol 125mg,-tocopherol acetate 100 mg, vitamin K33 mg, vitamin B12 mg, vitamin B25 mg, vitamin B64 mg, vitamin B12 0017 mg, niacin 40 mg, folic acid 18 mg,D-biotin 015 mg, calcium D-pantothenate 15 mg, Mn 100 mg,Fe 80 mg, Zn 80 mg, Cu 8 mg, I 2 mg, Co 05 mg, Se 015 mg.

colorimetric device. A *, b *, and L * indicate redness, yellowness, and brightness of the meat, respectively.

Antioxidant status

At the age of 42, the antioxidant status and lipid peroxidation level were determined, blood samples were taken from the wing vein of two chickens from each replicate in order to take their serum samples. After blood sampling, the chickens were slaughtered, and 2 g was taken from their livers and kept at -20 ° C with the serum samples until analysis. Total serum antioxidant capacity (TAC.S) and malondialdehyde level (MDA) were determined. After homogenizing in buffer solution (1.15% potassium chloride, pH 7.4), liver samples were centrifuged at 4 ° C for 15 minutes at 5000 rpm, and from the supernatant obtained to measure the activity of GPX and SOD, as well as MDA levels, were used. Glutathione peroxidase was determined using RANDOX kits (Germany) according to the manufacturer's instruction. Superoxide dismutase activity was assayed by the Winterbourn et al. (1975) method. The level of MDA, which is an indicator of lipid peroxidation, was measured in the serum and liver tissue of broilers.

Organoleptic properties of meat

The meat samples (from the right half of the pectoralis muscle) were then cooked at 170 ° C after be-

ing placed in an aluminum foil in an electric oven for 39-45 minutes, depending on the weight of the meat samples. Pieces of meat were prepared without adding spices and fat to prevent overlapping the chicken meat taste. The cooked samples were immediately cut into 12 pieces and presented to the panel members at random. The traits evaluated in the experiment were odor, taste, crispness, juiciness, and general acceptability. No information on meat or experimental treatments and working methods was provided to panel members. Respondent members were instructed to complete the evaluation form immediately after eating the meat samples according to their decision. A five-point scale was used, which is 1 to the lowest taste, juiciness, and crispness and 5 to the highest score compared to these indicators (Cross et al, 1986).

Statistical analyses

Statistical analysis of the data was performed using the GLM procedure of SAS software (9.2) (Pangan and Macit, 2019). The statistical model of the project was as follows:

$$Y_{ijkl} = \mu + A_i + B_j + C_k + AB_{ij} + AC_{ik} + BC_{jk} + \epsilon_{ijkl}$$

where: Y_{ijkl} = a dependent variable, μ = overall mean, A_i = the effect of Thyme essential oil, B_j = the effect of Mentha essential oil, C_k = the effect of microbial phytase, AB_{ij} = the interaction of thyme and

mint essential oils, AC_{ik} = the interaction of Thyme essential oil and microbial phytase, BC_{jk} = the interaction of Mentha essential oil and microbial phytase; ABC_{ijk} = the interaction of Thyme, Mentha essential oils and phytase and ϵ_{ijkl} = the residual deviation of the observation from the effects in the model. Tukey's test at the 5% level of probability was used to compare means.

RESULTS

Carcass quality

The effects of dietary supplementation essential oils (thyme and mentha) and microbial phytase on the carcass quality of chickens are shown in Table 2. The use of thyme and mentha essential oils increased the percentage of abdominal fat in chickens ($P < 0.05$). However, other carcass quality parameters in experimental broilers were not significantly different from control ($P > 0.05$).

Antioxidant status

The effects of dietary supplementation essential oils (Thyme and Mentha) and microbial phytase on the antioxidant status of chickens are shown in Table 3. The use of thyme essential oil significantly increased the levels of SOD and GPX in chickens ($P < 0.05$). However, other antioxidant parameters in experimental broilers were not significantly different from control ($P > 0.05$).

Organoleptic properties of meat

The effects of dietary supplementation essential oils (thyme and mentha) and microbial phytase on meat organoleptic properties of chickens are shown in Table 4. The use of thyme essential oil increased the overall acceptance of chicken meat ($P < 0.05$). However, other organoleptic properties of meat in experimental broilers were not significantly different from control ($P > 0.05$).

DISCUSSION

Carcass quality

These results were contradicted with the findings of Sadeghi et al. (2012) and Shirzadegan et al. (2014), who reported that the essential oils did not affect liver weight and abdominal fat. It seems that because these medicinal plants, in addition to having high antimicrobial and antioxidant properties, have antifungal properties, stimulate appetite, increase the digestibility of nutrients and feed consumption, and improve

the condition of the gastrointestinal tract. By adding these substances, the digestibility of nutrients (Aydin et al., 2010) and increases the energy released from the diet and excess energy is stored in the ventricular area in the form of fat in the animal. Baghban-Kanani et al. (2020) stated that difference in bird performance fed diets with phytase supplement may be due to a number of factors including phytase source, feed ingredients, and dietary characteristics. In this study, microbial phytase had no significant effect on carcass quality, resulting in differences in feed ingredients.

Antioxidant status

Considering that GPX and MDA enzymes are part of the body's first line of defense against free radicals (Ghobadi et al., 2017) and total antioxidant capacity is also an indicator of anti-radical, enzymatic, and non-enzymatic antioxidant activity (Banerjee et al., 2017; Akpro et al., 2019). Therefore, according to the results obtained from the liver's parameters, it can be said that the use of thyme essential oil has improved and strengthened the antioxidant status in the liver and serum of broilers. As the antioxidant status improves, the level of MDA (which is an indicator of lipid peroxidation in the body) decreased (Bai et al., 2019; Hamidi et al., 2021). Antioxidant activities of essential oils such as thyme and oregano have been reported in various studies (Han et al., 2017; Sakkas and Papadopoulou, 2017; Cao et al., 2018). Traditionally, it is considered that the excessive reactive oxygen species (ROS) can be removed or reduced by antioxidant enzymes such as GPX and SOD. The enhanced ROS generation can lead to oxidative damages and decrease antioxidant enzymes activities (Pan et al., 2021).

Organoleptic properties of meat

Taste is considered a major factor in the marketing of broilers. Flavor-related amino acids (valine, isoleucine, leucine, phenylalanine, arginine, proline, and methionine) are related to the tangy flavor in meat (Ali et al., 2019; Lengkidworraphiph et al., 2020). This shows that the used essential oils affect the amino acid profiles and improve the taste of the meat. Adding plant essential oils to poultry diets can have a positive effect on some physiological characteristics, carcass quality, as well as the quality of stored meat (Dávila-Ramírez et al., 2020; Ashour et al., 2020). Herbal additives are also used as a substitute for synthetic antioxidants due to their antioxidant properties, and while maintaining the quality

Table 2. Effects of adding essential oils (thyme and mentha) and microbial phytase on the carcass quality of broilers

effects	Back and neck	Breast	Thigh	Bursa of fabricius	Spleen	Liver	Gizzard	Intestine	Abdominal fat
Main effects									
Thyme									
0	26.7450	32.9662	26.8906	0.1743	0.1356	2.5575	2.3450	7.3175	1.7962 ^b
200	27.2575	32.4043	26.5606	0.1756	0.1556	2.7500	2.4575	7.3868	2.1300 ^a
SEM	0.2360	0.3516	0.4813	0.0115	0.0072	0.0642	0.0672	0.1815	0.0825
p-value	0.1377	0.2697	0.6322	0.9398	0.0630	0.0865	0.2483	0.7893	0.0087
mentha									
0	26.8943	33.1325	26.3975	0.1693	0.1493	2.5275	2.3962	7.2531	1.8031 ^b
200	27.1081	32.2381	27.0537	0.1806	0.1418	2.8100	2.4062	7.4512	2.1231 ^a
SEM	0.2360	0.3516	0.4813	0.0115	0.0072	0.0642	0.0672	0.1815	0.0825
p-value	0.5280	0.0847	0.3446	0.4989	0.4717	0.0048	0.9171	0.4479	0.0114
Microbial phytase									
0	27.1375	32.9412	26.2106	0.1768	0.1425	2.6525	2.4106	7.2106	1.9056
500	26.8650	32.4293	27.2406	0.1731	0.1487	2.6850	2.3918	7.4937	2.0206
SEM	0.2360	0.3516	0.4813	0.0115	0.0072	0.0642	0.0672	0.1815	0.0825
p-value	0.4223	0.3136	0.1433	0.8209	0.5480	0.7239	0.8453	0.2811	0.3347
Interactions									
Thyme × mentha									
0 0	26.7375	33.54	26.7925	0.1750	0.1462	2.4650	2.3362	6.9462	1.6162
0 200	26.7525	32.3925	26.9887	0.1737	0.1250	2.7100	2.3537	7.6887	1.9762
200 0	27.0512	32.7250	26.0025	0.1637	0.1525	2.59	2.4562	7.56	1.99
200 200	27.4637	32.0837	27.1187	0.1875	0.1587	2.91	2.4587	7.2137	2.27
SEM	0.3337	0.4973	0.6807	0.0163	0.0102	0.0909	0.0950	0.2567	0.1168
p-value	0.5571	0.6154	0.5057	0.4530	0.1926	0.6837	0.9378	0.0445	0.7350
Thyme × phytase									
0 0	27.0975	32.9925	26.7850	0.1725	0.1325	2.6037	2.3362	7.1912	1.6612
0 500	26.3925	32.94	26.9962	0.1762	0.1387	2.5712	2.3537	7.4437	1.9312
200 0	27.1775	32.89	25.6363	0.1812	0.1525	2.7012	2.4850	7.23	2.15
200 200	27.3375	31.9187	27.4850	0.17	0.1587	2.7987	2.43	7.5437	2.11
SEM	0.3337	0.4973	0.6807	0.0163	0.0102	0.0909	0.0950	0.2567	0.1168
p-value	0.2074	0.3648	0.2408	0.6513	1	0.4816	0.7064	0.9061	0.1970
mentha × phytase									
0 0	27.1025	33.06	26.2062	0.1687	0.1462	2.4587	2.4175	7.0962	1.8162
0 500	26.6862	33.2050	26.5887	0.17	0.1525	2.5962	2.3750	7.41	1.79
200 0	27.1725	32.8225	26.2150	0.1850	0.1387	2.8462	2.4037	7.3250	1.9950
200 500	27.0437	31.6537	27.8925	0.1762	0.1450	2.7737	2.4087	7.5775	2.2512
SEM	0.3337	0.4973	0.6807	0.0163	0.0102	0.0909	0.0950	0.2567	0.1168
p-value	0.6705	0.1990	0.3510	0.7629	1	0.2595	0.8049	0.9061	0.2383
effects									
Thyme	mentha	phytase							
0 0 0	27.0575	32.7725	26.8250	0.1750	0.1450	2.4425	2.3250	7.0225	1.6050
200 0 0	27.1475	33.3475	25.5875	0.1625	0.1475	2.4750	2.51	7.17	2.0275
0 200 0	27.1375	33.2125	26.7450	0.17	0.12	2.7650	2.3475	7.36	1.7175
0 0 500	26.4175	34.3075	26.76	0.1750	0.1475	2.4875	2.3475	6.87	1.6275
200 200 0	27.2075	32.4325	25.6850	0.20	0.1575	2.9275	2.46	7.29	2.2725
200 0 500	26.9550	32.1025	26.4175	0.1650	0.1575	2.7050	2.4025	7.95	1.9525
0 200 500	26.3675	31.5725	27.2325	0.1775	0.13	2.6550	2.36	8.0175	2.2350
200 200 500	27.72	31.7350	28.5525	0.1750	0.16	2.8925	2.4575	7.1375	2.2675
SEM			0.4720	0.7032	0.9626	0.0231	0.0145	0.1285	0.1344
p-value			0.5376	0.0735	0.5905	0.5983	0.7179	0.7649	0.7650

Means with different letters in each column have a significant difference (P <0.05).

Table 3. Effects of adding essential oils (thyme and mentha) and microbial phytase on the antioxidant status of broilers

effects	TAC. S	MDA. S ¹	GPX.L	SOD.L	MDA.L ²
Main effects					
Thyme					
0	1.6875	4.7812	2.6918 ^b	15.4193 ^b	1.13
200	1.7025	4.8687	2.9750 ^a	17.0781 ^a	1.2875
SEM	0.1203	0.3897	0.0990	0.4055	0.0617
p-value	0.9305	0.8752	0.0546	0.0080	0.0837
mentha					
0	1.6175	4.7687	2.92	16.3275	1.1875
200	1.7725	4.8812	2.7468	16.17	1.23
SEM	0.1203	0.3897	0.0990	0.4055	0.0617
p-value	0.3713	0.84	0.2287	0.7859	0.6307
Microbial phytase					
0	1.7425	4.6187	2.6987	16.1418	1.15
500	1.6475	5.0312	2.9681	16.3556	1.2675
SEM	0.1203	0.3897	0.0990	0.4055	0.0617
p-value	0.5817	0.4615	0.0665	0.7126	0.1908
Interactions					
Thyme × mentha					
0 0	1.6062	4.7750	2.7275	15.3112	1.05
0 200	1.7687	4.7875	2.6562	15.5275	1.21
200 0	1.6287	4.7625	3.1125	17.3437	1.3250
200 200	1.7762	4.9750	2.8375	16.8125	1.25
SEM	0.1701	0.5511	0.1401	0.5734	0.0872
p-value	0.9652	0.8576	0.4743	0.5208	0.1908
Thyme × phytase					
0 0	1.7675	4.6875	2.59	14.9375	1.05
0 500	1.6075	4.8750	2.7937	15.9012	1.21
200 0	1.7175	4.55	2.8075	17.3462	1.25
200 500	1.6875	5.1875	3.1425	16.81	1.32
SEM	0.1701	0.5511	0.1401	0.5734	0.0872
p-value	0.7058	0.6867	0.6438	0.2033	0.6307
mentha × phytase					
0 0	1.7425	4.6375	2.8712	16.56	1.1625
0 500	1.4925	4.90	2.9687	16.0950	1.2125
200 0	1.7425	4.60	2.5262	15.7237	1.1375
200 500	1.8025	5.1625	2.9675	16.6162	1.3225
SEM	0.1701	0.5511	0.1401	0.5734	0.0872
p-value	0.3713	0.7878	0.2319	0.2482	0.4446
effects					
Thyme	mentha	phytase			
0 0 0	1.7225	4.95	2.6250	1463.50	0.90 ^d
200 0 0	1.7625	4.3250	3.1175	18.4850	1.4250 ^a
0 200 0	1.8125	4.4250	2.5550	15.24	1.20 ^b
0 0 500	1.49	4.60	2.83	15.9875	1.20 ^b
200 200 0	1.6725	4.7750	2.4975	16.2075	1.0750 ^c
200 0 500	1.4950	5.20	3.1075	16.2025	1.2250 ^b
0 200 500	1.7250	5.15	2.7575	15.8150	1.22 ^b
200 200 500	1.88	5.1750	3.1775	17.4175	1.4250 ^a
SEM		0.2406	0.7794	0.1981	0.8110
p-value		0.632	0.4888	0.2287	0.0750

Means with different letters in each column have a significant difference (P <0.05).

¹MDA level on blood samples²MDA level on liver samples

Table 4. Effects of adding essential oils (thyme and mentha) and microbial phytase on organoleptic properties of meat in broilers

effects	Acceptance	Crisp	Flavor	Hydration	Smell	b	a	L
Main effects								
Thyme								
0	3.03375	3.0512	2.9425	3.1775	2.9812 ^b	10.7143 ^b	3.8837 ^a	51.4737 ^b
200	3.7937	3.0275	3.4818	3.5206	3.5131 ^a	12.5368 ^a	2.9650 ^b	53.3618 ^a
SEM	0.1691	0.2319	0.2441	0.1778	0.1587	0.2859	0.0897	0.2856
p-value	0.1615	0.9429	0.1314	0.1852	0.0262	0.0001	<0.0001	<0.0001
mentha								
0	3.0643	2.8781	3.0362	3.1850	3.1850	11.2443	3.7106 ^a	51.8468 ^b
200	3.3487	3.2006	3.3881	3.5131	3.3093	12.0068	3.1381 ^b	52.9887 ^a
SEM	0.1691	0.2319	0.2441	0.1778	0.1587	0.2859	0.0897	0.2856
p-value	0.2463	0.3353	0.3183	0.2044	0.5846	0.0715	<0.0001	0.0093
Microbial phytase								
0	3.1018	3.0681	3.0600	3.2475	3.0593	10.9687 ^b	3.7062 ^a	52.2475
500	3.3112	3.0106	3.3643	3.4506	3.4350	12.2825 ^a	3.1425 ^b	52.5851
SEM	0.1691	0.2319	0.2441	0.1778	0.1587	0.2859	0.0897	0.2856
p-value	0.3902	0.8623	0.3868	0.4273	0.1072	0.0034	0.0002	0.4075
Interactions								
Thyme × mentha								
0 0	2.7178	2.7287	2.5912	2.8725	2.70	10.2262	4.5437 ^a	50.5412
0 200	3.3487	3.3737	3.2937	3.4825	3.2625	11.2025	3.2337 ^b	52.4062
200 0	3.4100	3.0275	3.4812	3.4975	3.67	12.2625	2.8775 ^c	53.1525
200 200	3.3487	3.0275	3.4825	3.5437	3.3562	12.8112	3.0525 ^b	53.5712
SEM	0.2392	0.3279	0.3452	0.2515	0.2244	0.4043	0.1268	0.4040
p-value	0.1615	0.3353	0.3200	0.2735	0.0627	0.6019	<0.0001	0.0861
Thyme × phytase								
0 0	2.8962	3.0925	2.7162	3.1375	2.6525	9.0650	4.5050 ^a	50.8187
0 500	3.1712	3.01	3.1687	3.2175	3.31	12.3627	3.2625 ^b	52.1287
200 0	3.3075	3.0437	3.4037	3.3575	3.4662	12.8725	2.9075 ^c	53.6762
200 500	3.4512	3.0112	3.56	3.6837	3.56	12.2012	3.0225 ^{bc}	53.0475
SEM	0.2392	0.3279	0.3452	0.2515	0.2244	0.4043	0.1268	0.4040
p-value	0.7862	0.9399	0.6718	0.6289	0.2213	<0.0001	<0.0001	0.0246
mentha × phytase								
0 0	2.9487	3.0437	2.7012	3.0750	3.0125	10.7525	4.1712 ^a	51.7162
0 500	3.1800	2.7125	3.3712	3.2950	3.3575	11.7362	3.2500 ^b	51.9775
200 0	3.2550	3.0925	3.4187	3.4200	3.1062	11.1850	3.2412 ^b	52.7787
200 500	3.4425	3.3087	3.3575	3.6062	3.5125	12.8287	3.0350 ^b	53.1987
SEM	0.2392	0.3279	0.3452	0.2515	0.2244	0.4043	0.1268	0.4040
p-value	0.9279	0.4121	0.3002	0.9471	0.8926	0.4224	0.0095	0.8459
effects								
Thyme	mentha	phytase						
0 0 0	2.5325	2.9350	2.0925	2.7775	2.34	8.0375	5.8125 ^a	49.3525
200 0 0	3.3650	3.1525	3.31	3.3725	3.6850	13.4675	2.5300 ^c	54.0800
0 200 0	3.2600	3.25	3.34	3.4975	2.9650	10.0925	3.1975 ^b	52.2850
0 0 500	2.9050	2.5225	3.09	2.9675	3.06	12.4150	3.2750 ^b	51.7300
200 200 0	3.25	2.9350	3.4975	3.4325	3.2475	12.2775	3.2850 ^b	53.2725
200 0 500	3.4550	2.9025	3.6525	3.6225	3.6550	11.0575	3.2250 ^b	52.2250
0 200 500	3.4375	3.4975	3.2475	3.4675	3.56	12.3125	3.2500 ^b	52.5275
200 200 500	3.4475	3.12	3.4675	3.7450	3.4650	13.3450	2.8200 ^c	53.8700
SEM	0.3383	0.4638	0.4883	0.3557	0.3174	0.9864	0.1794	0.9215
p-value	0.7547	0.8653	0.6082	0.7145	0.6819	0.5718	<0.0001	0.5713

Means with different letters in each column have a significant difference (P <0.05).

of meat, they have no adverse effects on the product and the environment (Pateiro et al., 2018; Hashemi et al., 2020). Lipid oxidation also plays a role in determining the final pH and total pigment concentration and, consequently, the color of the meat in the muscle (Kılıç et al., 2018; Domínguez et al., 2019). Plant essential oils with their antioxidant properties reduce the lipid oxidation of stored meat (Aminzare et al., 2019). They can also affect *in vivo* antioxidant systems such as vitamin E, superoxide dismutase, and glutathione peroxidase (Diniz do Nascimento et al., 2020; Sharma et al., 2020). Therefore, the observed variation in the amount of redness and yellowness in meat can be attributed to the antioxidant effect of essential oils in meat. The rate of discoloration in fresh meat is related to the rate of pigment oxidation, oxygen consumption, and the effectiveness of the meth myoglobin regenerative system (Buzrul, 2017; Khan et al., 2018; Flores et al., 2020).

CONCLUSIONS

The use of thyme essential oil increased the overall acceptance of chicken meat. Our results showed that essential oil influences chicken meat antioxidant properties. Moreover, it affected the smell, b and L parts of the meat. Use of thyme essential oil in broiler ration is recommended.

CONFLICT OF INTEREST

The authors declare that they have no known competing financial interests or personal relationship that could influence the work reported in this paper.

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REFERENCES

Abd El-Ghany, W. A., 2020. Phytobiotics in poultry industry as growth promoters, antimicrobials and immunomodulators - A review. *J. World Poult. Res.* 10 (4), 571-579.

Abd El-Ghany, W.A., Ismail M., 2014. Tackling of experimental colisepticaemia in broiler chickens using phytobiotic essential oils and antibiotic alone or in combination. *Iranian J. Vet. Res.* 15 (2), 110-115.

Adaszynska-Skwirzynska, M., Szczerbińska, D., 2018. The antimicrobial activity of lavender essential oil (*Lavandula angustifolia*) and its influence on the production performance of broiler chickens. *J. Anim. Physiol. Anim. Nut.* 102 (4), 1020-1025.

Ahmadi, O., Jafarizadeh-Malmiri, H., 2020. Intensification and optimization of the process for thyme oil in water nanoemulsions preparation using subcritical water and xanthan gum. *Zeitschrift für Physikalische Chemie* 1 (ahead-of-print).

Akpro, L.A., Gbogouri, G.A., Konan, B.R., Issali, A.E., Konan, K.J.L., Brou, K.D., Nemlin, G.J., 2019. Phytochemical compounds, antioxidant activity and non-enzymatic browning of sugars extracted from the water of immature coconut (*Cocos nucifera* L.). *Sci. African.* 6, e00123. <https://doi.org/10.1016/j.sciaf.2019.e00123>

Ali, M., Lee, S.Y., Park, J.Y., Jung, S., Jo, C., Nam, K.C., 2019. Comparison of functional compounds and micronutrients of chicken breast meat by breeds. *Food Sci. Anim. Res.* 39 (4), 632.

Aminzare, M., Hashemi, M., Ansarian, E., Bimkar, M., Azar, H.H., Mehrabi, M.R., Afshari, A., 2019. Using natural antioxidants in meat and meat products as preservatives: A review. *Adv. Anim. Vet. Sci.* 7 (5), 417-426.

Ashour, E.A., Abd El-Hack, M.E., Swelum, A.A., Osman, A.O., Taha, A.E., Alhimaidi, A.R., Ismail, I.E., 2020. Does the dietary graded levels of herbal mixture powder impact growth, carcass traits, blood indices and meat quality of the broilers?. *Italian J. Anim. Sci.* 19 (1), 1228-1237.

Aydin, A., Pekel, A.Y., Issa, G., Demirel, G., Patterson, P.H., 2010. Effects of dietary copper, citric acid, and microbial phytase on digesta pH and ileal and carcass microbiota of broiler chickens fed a low available phosphorus diet. *J. Appl. Poult. Res.* 19 (4), 422-431.

Baghban-Kanani, P., Hosseintabar-Ghasemabad, B., Azimi-Youvalari, S., Seidavi, A., Ayaşan, T., Laudadio, V., Tufarelli, V., 2018. Effect of different levels of sunflower meal and multi-enzyme complex on performance, biochemical parameters and antioxidant status of laying hens. *S. Afr. J. Anim. Sci.* 48 (2), 390-399.

Baghban-Kanani, P., Hosseintabar-Ghasemabad, B., Azimi-Youvalari, S., Seidavi, A., Laudadio, V., Mazzei, D., Tufarelli, V., 2020. Effect of dietary sesame (*Sesame indicum* L) seed meal level supplemented with lysine and phytase on performance traits and antioxidant status of late-phase laying hens. *Asian-Australas J. Anim. Sci.* 33 (2), 277.

Bai, X., Dai, S., Li, J., Xiao, S., Wen, A., Hu, H., 2019. Glutamine improves the growth performance, serum biochemical profile and antioxidant status in broilers under medium-term chronic heat stress. *J. Appl. Poult. Rese.* 28 (4), 1248-1254.

Banerjee, R., Verma, A.K., Siddiqui, M.W., 2017. Natural Antioxidants: Applications in Foods of Animal Origin. CRC Press.

Besharati, M., Palangi, V., Niazifar, M., Nemat, Z., 2020. Comparison study of flaxseed, cinnamon and lemon seed essential oils additives on quality and fermentation characteristics of lucerne silage. *Acta Agric. Slovenica.* 2 (424), 115.

Bouhitt, F., Najar, M., Agha, D.M., Melki, R., Najimi, M., Sadki, K., Le-walle, P., Hamal, A., Lagneaux, L., Merimi, M., 2019. The biological response of mesenchymal stromal cells to thymol and carvacrol in comparison to their essential oil: An innovative new study. *Food Chem. Toxic.* 134, 110844.

Brochot, A., Guilbot, A., Haddioui, L., Roques, C., 2017. Antibacterial, antifungal, and antiviral effects of three essential oil blends. *Microbiologyopen.* 6 (4), e00459.

Buzrul, S., 2017. Advances in High Hydrostatic Pressure for Meat and Meat Processing. In Advanced Technologies for Meat Processing (pp. 227-258). CRC Press.

Cao, T.L., Yang, S.Y., Song, K.B., 2018. Development of burdock root inulin/chitosan blend films containing oregano and thyme essential oils. *Int. J. Molecular Sci.* 19 (1), 131.

Chowdhury, S., Mandal, G.P., Patra, A.K., Kumar, P., Samanta, I., Pradhan, S., Samanta, A.K., 2018. Different essential oils in diets of broiler chickens: 2. Gut microbes and morphology, immune response, and some blood profile and antioxidant enzymes. *Anim. Feed Sci. Technol.* 236, 39-47.

Cimrin, T., Tunca, R.I., Avsaroglu, M.D., Ayasan, T., Kücükseran, S., 2020. Effects of an antibiotic and two phylogenetic substances (cinnamaldehyde and 1,8-cineole) on yolk fatty acid profile and storage period-associated egg lipid peroxidation level. *Rev. BrasileiraZootech.* 49, e20190270.

Criste, R.D., Panaite, T.D., Tabuc, C., Sărăcile, M., Șoica, C., Olteanu, M., 2017. Effect of oregano and rosehip supplements on broiler (14-35 days) performance, carcass and internal organs development and gut health. *AgroLife Sci. J.* 6 (1), 75-83.

Cross, H.R., Durland, P.R., Seideman, S.C., 1986. Sensory qualities of meat. *Mus. Food.* 279-320.

Dávila-Ramírez, J.L., Munguía-Acosta, L.L., Morales-Coronado, J.G., García-Salinas, A.D., González-Ríos, H., Celaya-Michel, H., Barreira-Silva, M.A., 2020. Addition of a mixture of plant extracts to diets for growing-finishing pigs on growth performance, blood metabolites, carcass traits, organ weight as a percentage of live weight, quality and sensorial analysis of meat. *Animals.* 10 (7), 1229.

Diniz do Nascimento, L., Moraes, A.A.B.D., Costa, K.S.D., Pereira Galúcio, J.M., Taube, P.S., Costa, C.M.L., Faria, L.J.G.D., 2020. Bioactive natural compounds and antioxidant activity of essential oils from spice plants: New findings and potential applications. *Biomolecules.* 10 (7), 988.

Domínguez, R., Pateiro, M., Gagaua, M., Barba, F.J., Zhang, W., Lorenzo, J.M., 2019. A comprehensive review on lipid oxidation in meat and meat products. *Antioxidants.* 8 (10), 429.

Farhadi, D., Karimi, A., Sadeghi, G., Rostamzadeh, J., Bedford, M.R., 2017. Effects of a high dose of microbial phytase and myo-inositol supplementation on growth performance, tibia mineralization, nutrient digestibility, litter moisture content, and foot problems in broiler chickens fed phosphorus-deficient diets. *Poult. Sci.* 96 (10), 3664-3675.

Flores, D. R. M., da Fonseca, A. F. P., Schmitt, J., Tonetto, C. J., Junior, A. G. R., Hammerschmitt, R. K., Nörnberg, J. L., 2020. Lambs fed with increasing levels of grape pomace silage: Effects on meat quality. *Small Rum. Res.* 195, 106234.

Ghobadi, A., Shirazi, A., Najafi, M., Kahkesh, M.H., Rezapoor, S., 2017. Melatonin ameliorates radiation-induced oxidative stress at targeted and nontargeted lung tissue. *Medical Phy. J.* 42 (4), 241.

Grigore, A., Paraschiv, I. N. A., Colceru-Mihul, S., Bubueanu, C., Draghici, E., Ichim, M., 2010. Chemical composition and antioxidant activity of *Thymus vulgaris* L. volatile oil obtained by two different methods. *Romanian Biotech. Let.* 15 (4), 5436-5443.

Gülçin, İ., Gören, A.C., Taslimi, P., Alwasel, S.H., Kılıç, O., Bursal, E., 2020. Anticholinergic, antidiabetic and antioxidant activities of Anatolian pennyroyal (*Mentha pulegium*)-analysis of its polyphenol contents by LC-MS/MS. *Biocat. Agric. Biotechnol.* 23, 101441.

Hamidi, O., Chamani, M., Ghahri, H., Sadeghi, A. A., Malekinejad, H., Palangi, V., 2021. Effects of Supplemental Chromium Nanoparticles on IFN- γ expression of Heat Stress Broilers. *Biol. Trace Elem. Res.* 1-9.

Han, F., Ma, G.Q., Yang, M., Yan, L., Xiong, W., Shu, J.C., Xu, H.L., 2017. Chemical composition and antioxidant activities of essential oils from different parts of the oregano. *J. Zhejiang Uni. Sci. B.* 18 (1), 79-84.

Hashemi, M., Daneshamooz, S., Raeisi, M., Jannat, B., Taheri, S., Noori, S.M.A., 2020. An overview on antioxidants activity of polysaccharide edible films and coatings contains essential oils and herb extracts in meat and meat products. *Adv. Anim. Vet. Sci.* 8 (2), 198-207.

Khan, A. Z., Kumbhar, S., Liu, Y., Hamid, M., Pan, C., Nido, S. A., Huang, K., 2018. Dietary supplementation of selenium-enriched probiotics

enhances meat quality of broiler chickens (*Gallus gallusdomesticus*) raised under high ambient temperature. *Biol. Trace Elem. Res.* 182 (2), 328-338.

Kılıç, B., Şimşek, A., Claus, J. R., Karaca, E. S. R. A., Bilecen, D. A. M. L. A., 2018. Improving lipid oxidation inhibition in cooked beef hamburger patties during refrigerated storage with encapsulated polyphosphate incorporation. *LWT*. 92, 290-296.

Lengkidworraphiphat, P., Wongpoomchai, R., Bunmee, T., Chariyakornkul, A., Chaiwang, N., Jaturasitha, S., 2020. Taste-Active and Nutritional Components of Thai Native Chicken Meat: a Perspective of Consumer Satisfaction. *Food Sci. Anim. Res.* <https://doi.org/10.5851/koosta.2020.e94>

Palangi, V., Macit, M., 2019. In situ crude protein and dry matter ruminal degradability of heat-treated barley. *Rev. Méd. Vét.* 170, 123-128.

Pan, X., Zhang, L., Xing, T., Li, J., Gao, F., 2021. The impaired redox status and activated Nrf2/ARE pathway in wooden breast myopathy in broiler chickens. *Anim. Bios.* 34 (4), 652-661.

Pateiro, M., Barba, F. J., Domínguez, R., Sant'Ana, A. S., Khanegah, A. M., Gavahian, M., Lorenzo, J. M., 2018. Essential oils as natural additives to prevent oxidation reactions in meat and meat products: A review. *Food Res. Int.* 113, 156-166.

Pirgozliev, V., Mansbridge, S. C., Rose, S. P., Mackenzie, A. M., Beccaccia, A., Karadas, F., Bravo, D., 2019. Dietary essential oils improve feed efficiency and hepatic antioxidant content of broiler chickens. *Animal.* 13 (3), 502-508.

Roofchaei, A., Rezaeipour, V., Vatandour, S., Zaefarian, F., 2019. Influence of dietary carbohydrases, individually or in combination with phytase or an acidifier, on performance, gut morphology and microbial population in broiler chickens fed a wheat-based diet. *Anim. Nut.* 5 (1), 63-67.

Sabo, V. A., Knezevic, P., 2019. Antimicrobial activity of *Eucalyptus camaldulensis* Dehn. plant extracts and essential oils: A review. *Indust. Crops Prod.* 132, 413-429.

Sabuna, C., Harimurti, S., Nurcahyo, R. W., 2019. Utilization of distillation waste of Lemon Grass (*Cymbopogon nardus*) as litter for reducing parasite diseases and its influence on broiler performance. In IOP Conference Series: Earth Environ. Sci. 387 (1), 012066. IOP Publishing.

Sadeghi, G. H., Karimi, A., PadidarJahromi, S. H., Azizi, T., Daneshmand, A., 2012. Effects of cinnamon, thyme and turmeric infusions on the performance and immune response in of 1-to 21-day-old male broilers. *Brazilian J. Poul. Sci.* 14 (1), 15-20.

Sakkas, H., Papadopoulou, C., 2017. Antimicrobial activity of basil, oregano, and thyme essential oils. *J. Mic. Biotechnol.* 27 (3), 429-438.

Semjon, B., Marcinčáková, D., Koréneková, B., Bartkovský, M., Nagy, J., Turek, P., Marcinčák, S., 2020. Multiple factorial analysis of physicochemical and organoleptic properties of breast and thigh meat of broilers fed a diet supplemented with humic substances. *Poult. Sci.* 99 (3), 1750-1760.

Sevim, B., Gümüş, E., Harman, H., Ayasan, T., Başer, E., Altay, Y., Akbulut, K., 2020. Effects of dietary rosemary essential oil on growth performance, carcass traits and some hematological values of chukar partridge. *Turkish J. Agric. Food Sci. Technol.* 8 (2), 430-435.

Sharma, K., Guleria, S., Razdan, V. K., Babu, V., 2020. Synergistic antioxidant and antimicrobial activities of essential oils of some selected medicinal plants in combination and with synthetic compounds. *Ind. Crops Prod.* 154, 112569.

Shirzadegan, K., 2014. Reactions of modern broiler chickens to administration of cinnamon powder in the diet. *Iranian J. Appl. Anim. Sci.* 4 (2), 367-371.

Sidiropoulou, E., Skoufos, I., Marugan-Hernandez, V., Giannenas, I., Bonos, E., Aguiar-Martins, K., Tzora, A., 2020. In vitro anticoccidial study of oregano and garlic essential oils and effects on growth performance, faecal oocyst output and intestinal microbiota in vivo. *Frontiers Vet. Sci.* 7, 420.

Tabari, M. A., Youssefi, M. R., Maggi, F., Benelli, G., 2017. Toxic and repellent activity of selected monoterpenoids (thymol, carvacrol and linalool) against the castor bean tick, *Ixodes ricinus* (Acari: Ixodidae). *Vet. Parasitol.* 245, 86-91.

Valdivieso-Ugarte, M., Gomez-Llorente, C., Plaza-Díaz, J., Gil, Á., 2019. Antimicrobial, antioxidant, and immunomodulatory properties of essential oils: A systematic review. *Nutrients.* 11 (11), 2786.

Vašková, J., Patlevič, P., Žatko, D., Marcinčák, S., Vaško, L., Nagy, K. K. J., 2018. Effects of humic acids on poultry under stress conditions. *Slovenian Vet. Res.* 55 (4), 245-253.

Vašková, J., Patlevič, P., Žatko, D., Vaško, L., Marcinčák, S., 2015. Impact of humic acids on trace element content under different conditions. *FOLIA.* 59 (3), 159-164.

Winterbourne, C. C., Hawkins, R. E., Brian, M., Carrell, R. W., 1975. The estimation of red cell superoxide dismutase activity. *J. Lab. Clinical Med.* 85 (2), 337-341.

Yadav, S., Jha, R., 2019. Strategies to modulate the intestinal microbiota and their effects on nutrient utilization, performance, and health of poultry. *J. Anim. Sci. Biotechnol.* 10 (1), 1-11.

Yang, Y. F., Zhao, L. L., Shao, Y. X., Liao, X. D., Zhang, L. Y., Lin, L. U., Luo, X. G., 2019. Effects of dietary graded levels of cinnamon essential oil and its combination with bamboo leaf flavonoid on immune function, antioxidative ability and intestinal microbiota of broilers. *J. Int. Agric.* 18 (9), 2123-2132.

Zhai, H., Liu, H., Wang, S., Wu, J., Klunder, A. M., 2018. Potential of essential oils for poultry and pigs. *Anim. Nut.* 4 (2), 179-186.

Zhuang, H., Savage, E. M., 2009. Variation and Pearson correlation coefficients of Warner-Bratzler shear force measurements within broiler breast fillets. *Poult. Sci.* 88 (1), 214-220.