

Journal of the Hellenic Veterinary Medical Society

Vol 72, No 2 (2021)



Pharmacokinetics and plasma concentration of thiopental in normal and stressed chickens with hydrogen peroxide

Y.J. MOUSA, S.M. AMIN, KH.A. SHAABAN

doi: [10.12681/jhvms.27540](https://doi.org/10.12681/jhvms.27540)

Copyright © 2021, Y.J. MOUSA, S.M. AMIN, KH.A. SHAABAN



This work is licensed under a [Creative Commons Attribution-NonCommercial 4.0](https://creativecommons.org/licenses/by-nc/4.0/).

To cite this article:

MOUSA, Y., AMIN, S., & SHAABAN, K. (2021). Pharmacokinetics and plasma concentration of thiopental in normal and stressed chickens with hydrogen peroxide. *Journal of the Hellenic Veterinary Medical Society*, 72(2), 2961–2968. <https://doi.org/10.12681/jhvms.27540>

Pharmacokinetics and plasma concentration of thiopental in normal and stressed chickens with hydrogen peroxide

Y. J. Mousa^{*}, S. M. Amin, Kh. A. Shaaban

Department of Physiology, Biochemistry and Pharmacology, College of Veterinary Medicine, University of Mosul, Mosul, Iraq

ABSTRACT: No previous study deals with hydrogen peroxide (H₂O₂)-induced oxidative stress (OS) and its influence on thiopental anesthesia, its plasma concentration and pharmacokinetic profile in 7-14 day old chickens. OS induction was made by the daily supply of H₂O₂ from day one to the completion of the experiments on day 14th of chickens' age. Median Effective Doses (ED₅₀s) of hypnosis and analgesia were revealed an increase in thiopental efficacy by 7 and 18%, respectively in the stressed chickens in comparison to the normal one. Thiopental plasma concentration was analyzed at time 0.5, 1, 2, 4 and 24 hours after its injection at 35 mg/kg, IP to be 91.42, 54.35, 38.27, 22.30 and 7.51 µg/ml in the normal chickens and increased significantly by 74, 84, 48, 85 and 82% to be 159.01, 100.06, 56.71, 41.30 and 13.63 µg/ml in the stressed chickens, respectively. Thiopental pharmacokinetics parameters, which included Area Under the plasma Concentration (AUC_{0-∞}), Area Under the Moment Curve (AUMC_{0-∞}), Mean Residence Time (MRT) and elimination half-life (t_{1/2β}) were increased in the stressed chickens by 82, 94, 6 and 8%, as well as, Clearance (Cl), elimination rate constant (K_{el}), and Steady-State Volume of Distribution (V_{ss}) were decreased in the same group by 33, 14 and 41%, respectively. The results of this trial concluded that OS status intensifies thiopental anesthetic action mainly by increasing its plasma concentration and altering the pharmacokinetics profile, suggesting veterinarians to bear in mind when preparing the dose of thiopental to be given to stressed animals.

Keywords: Chickens, plasma concentration, pharmacokinetics, stress, thiopental

Corresponding Author:

Yaareb J. Mousa, Department of Physiology, Biochemistry and Pharmacology,
College of Veterinary Medicine, University of Mosul, Mosul, Iraq
E-mail address: yarub204@uomosul.edu.iq

Date of initial submission: 13-07-2020
Date of revised submission: 13-10-2020
Date of acceptance: 17-10-2020

INTRODUCTION

Thiopental belongs to the barbiturates anesthetics that used for the induction of short duration of the general anesthesia by its mode of action resulting from the potentiation of the inhibitory neurotransmitter effect, Gamma-Aminobutyric Acid (GABA), on its GABA_A receptor subtype, leading to an increase in the outflow of chloride ions inside the post-synaptic neurons that depresses the Central Nervous System (CNS) (Garcia et al., 2010; Clark et al., 2012; Naseri et al., 2017). Thiopental possesses excellent and reliable narcotic action with less efficient analgesia and muscle relaxation than other anesthetic agents like ketamine (Clark et al., 2012; Flecknell, 2015; Katzung and Trevor, 2015). H₂O₂ was used experimentally for the induction of the OS because it is well-known. A powerful oxidizing agent besides, its concentration (0.5%) and the length of treatment required to produce OS are well studied in the previous literature through its formation of free radicals that break down the vital cell components, stimulates lipid peroxidation and breaks down of proteins (including protein receptors) (Patockova et al., 2003; Sayre et al., 2008). The CNS considered more vulnerable to OS status, because of the high free radical formation, consuming a lot of oxygen, high amount of unsaturated fatty acids with less antioxidant activity (Pastore et al., 2003). The experimentally induced OS with H₂O₂ (0.5%) was found to modify the pharmacological response of some drugs that have a neuro-active mechanism like ketamine anesthesia (Mousa, 2014), diazepam and xylazine sedation (Mousa and Mohammad, 2012a;b) by pharmacodynamics interaction on their receptors and relevant pharmacokinetic interaction at a level of absorption, distribution, metabolism and elimination of the drugs which have an impact elevating drug efficacy and subsequent toxicity. The pharmacokinetics criteria of the drug are an important factor ensuring the drug delivery to the site of action on its target receptor to achieve efficacy and/or improve the preferred therapeutic effect with minimizing its adverse effect (Wen et al., 2015).

The purpose of this study was to use thiopental in normal chickens for the induction of the general anesthesia and to determine the possible efficacy of OS-induced by H₂O₂ at thiopental anesthesia, its plasma concentration and the pharmacokinetics profile.

MATERIALS AND METHODS

Animals

7-14-days old of broiler chickens (76 overall

chickens used in all the experiments) for both genders were used in all the study experiments with a regular bodyweight of 80–125 g. They were kept in cages at an ideal condition of 32–35°C and the floor of the cages comprising woody shreds. Chickens have admitted to the water and food ration freely. Thiopental [Thiopentone (Thiopental sodium), 2.5%, Egyptian International Pharmaceutical Industries Co, Egypt] diluted using normal saline solution (0.9% NaCl) to be injected by a dosage volume 5 ml/kg. The route of administration of thiopental was intraperitoneally (IP) as mentioned by a previous study in the chicks (Church, 1957).

Ethical considerations

This research and the use of experimental chickens have been valid, qualified and authenticated by the professional scientific board of the Veterinary Medicine College / Mosul University, according to international standards.

Induction of OS in the chickens

Chickens gotten from a native hatchery at one day old, divided randomly to control normal group that provided with tap water (H₂O group) whereas the other remaining chickens had treated with 0.5% H₂O₂ (50%, Scharlab, Spain) in drinking water (stressed or H₂O₂ group). The earlier studies authenticated that a continuous daily fresh supply of 0.5% H₂O₂ in tap water can prompt OS whenever assumed to chickens from 1st day and continued for 14 days. Then, H₂O₂ can prompt OS on the 7th, 10th and 14th days of chickens' lifetime through glutathione reduction and malondialdehyde raising in their concentrations at the chickens' brain and plasma (OS biomarkers) (Mousa and Mohammad, 2012a; Mousa, 2014; Kabirian et al., 2018; Aksoy and Alper, 2019). So that, 7-14-days of chickens' lifetime, were used in the current study.

Assessment of thiopental ED₅₀s in normal and stressed chickens

A. Thiopental hypnotic ED₅₀ in chickens

The hypnotic ED₅₀ value of thiopental was estimated in the normal and stress chickens, implement to the up-down technique mentioned earlier (Dixon, 1980). The first dosage for thiopental at 35 mg/kg, IP depends on a preliminary study. Chickens observed individually for 4 hours waiting for the hypnotic occurrence of thiopental showed by losing the righting reflex. The dosages of thiopental then would be di-

minished or augmented 10 mg reliant on the presence or lack of the hypnotic effect, respectively (Mousa and Al-Zubaidy, 2019; Mousa et al., 2019).

B. Thiopental analgesic ED₅₀ in chickens

For the normal and stressed chickens, the thiopental's analgesic ED₅₀ is estimated according to the previously described routine (Dixon, 1980). The first thiopental dosage be 35 mg/kg, IP depends on an introductory study. The chickens were measured before and after 15 minutes of thiopental injection by using the electro-stimulator device (Harvard apparatus, USA), which indicated by pain perception (distress call) in the chickens (Mousa, 2019; 2020; Mousa and Al-Zubaidy, 2019), the doses of thiopental then should be reduced or raised 10 mg as look or lack of the analgesia occur (the decrease or increase in doses was chosen whereby not higher than 30% of the first dose used of thiopental for accurate results).

Thiopental plasma concentration of normal and stress chickens

A single dosage of thiopental at 35 mg/kg, IP (which resembles the ED₁₀₀ of thiopental estimated in the previous experiment) was injected to normal and stressed chickens. Blood samples combined from the jugular vein of 5 chickens per measure time at 0.5, 1, 2, 4 and 24 hours for both the normal and stressed chicks. Then, adding the heparin (B. Braun Medical Inc, USA) (1:10) to the blood samples and undergoing centrifugation (Chalice, UK) at 3000 round per

minute along with 15 minutes to get plasma samples (Chauhan et al., 2020) which then freezes at -18 °C pending spectrophotometric analysis for 72 hours.

Spectrophotometric analysis

Preparation and calibration of the thiopental standards

Thiopental standards were made of 2.5, 5, 10, 20, 40, 80 and 160 µg/ml (Gustafsson et al., 1996) by dilution with 0.5 N of Sodium hydroxide (NaOH) and filtration then, the absorbance by Ultra Violet (UV) spectrophotometry (Lovibond, Germany) at 330 nanometers (nm) was taken by the procedure described by Goldbaun (1948) to reveal the standard curve of the simple linear regression to calculate the thiopental concentration in the unknown plasma samples. By using the equation of the simple linear regression of thiopental standards with a coefficient of determination ($R^2 = 0.8958$) (Fig. 1), thiopental concentration in plasma samples can be calibrated and calculated in normal and stressed chickens (Barwick, 2003) (Figure 1) were:

$y = a + bx$ ($y = 0.0974 + 0.0017x$) (Equation of the simple linear regression of thiopental standards)

y = absorbance of thiopental in the plasma samples at 330 nm

a = intercept

b = slope

x = thiopental concentration in the plasma samples (unknown)

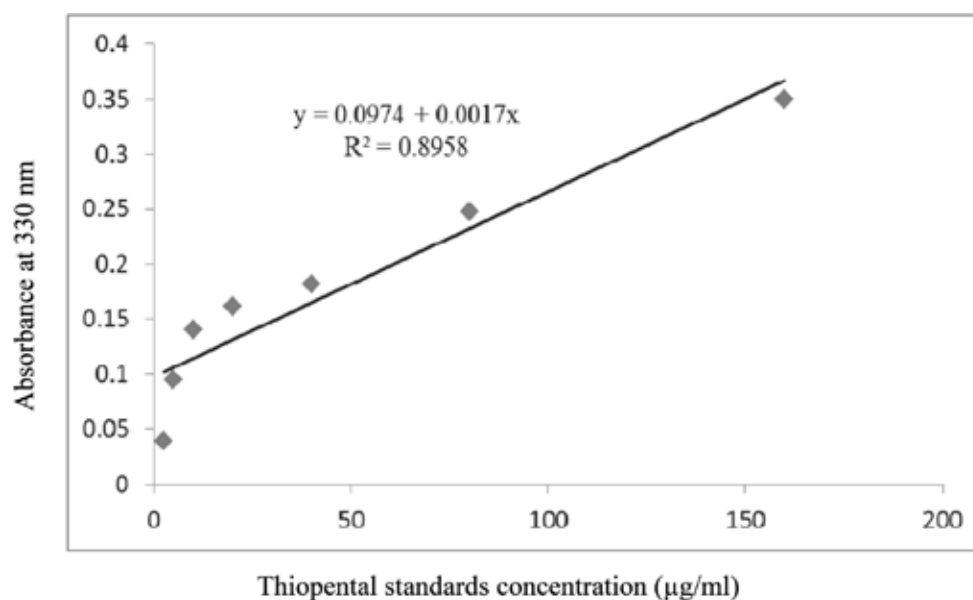


Figure 1. Equation of the simple linear regression for thiopental standards (2.5, 5, 10, 20, 40, 80 and 160 µg/ml), and their absorbance at 330 nm

Extraction and determination of thiopental in the plasma samples

A simple, validated and accurate method was used for determining thiopental concentration in the plasma (Goldbaum, 1948; Broughton, 1956) for both normal and stressed chickens in different measured times at 0.5, 1, 2, 4 and 24 hours after thiopental treatment. The procedure summarized by adding 10 ml of redistilled chloroform to 1 ml of the plasma and shake by tube shaker (Dragonlab, China) for 3 minutes. By using the filter paper, the aliquot will be obtained by using the funnel. Thereafter, 1 ml of NaOH (0.5 N) was added to the aliquot and the lower layer, then discarded and the remaining were centrifuged at 3000 rpm for 15 minutes to get the clear solution, which is the last consequent extraction result of the plasma sample which lastly analyzed by the UV spectrophotometry at 330 nm wavelength by using the quartz cuvette to record the absorbance for each unknown sample.

Determination of the thiopental pharmacokinetics parameters in the normal and stressed chickens

The non-compartmental method was used to find out the thiopental pharmacokinetics parameters in normal and stressed chickens by using a PKSolver program (Zhang et al., 2010) which included $AUC_{0-\infty}$ ($\mu\text{g}\cdot\text{h}/\text{ml}$), $AUMC_{0-\infty}$ ($\mu\text{g}\cdot\text{h}^2/\text{ml}$), MRT ($AUMC/AUC$)(h), Cl (dose/ AUC)(L/h/kg), Tmax (h), Cmax (μg), $t_{1/2\beta}$ (h), K_{el} ($0.693/t_{1/2\beta}$)(h^{-1}) and V_{ss} [dose. $AUMC/(AUC)^2$](L/kg). The increase or decrease in

the percentages of the thiopental pharmacokinetics parameters was calculated in the normal chickens and correlate to determine the changes in the pharmacokinetics parameters in the stressed chickens.

Statistics

The unpaired student T-test was implemented to relate the means of two groups of parametric statistical examinations by using the SPSS program (Katz, 2011; Petrie and Watson, 2013). The data would be considered significant when $p < 0.05$.

RESULTS

Thiopental ED₅₀ values the in normal and stressed chickens

A. The hypnotic ED₅₀ value of thiopental

The ED₅₀ values of thiopental that produce narcosis (lack of the righting reflex) in 50% of the chickens were 17.63 and 16.44 mg/kg, IP in the normal and stressed chickens, respectively, which have decreased by 7% that resembles the OS prompt by H₂O₂ to ameliorate the thiopental hypnotic ED₅₀ value as determined by the up-down technique (Table 1-A) which is stated here for the first time. The signs elicited by thiopental hypnosis in the chicks were ataxia, recumbency, locked eyelids, defecation, loss of the righting reflex, paddling movements of the legs and quit sleeping with normal breathing. These signs, as it observed, it became exaggerated at the stressed chickens.

Table 1-A. Thiopental's hypnotic ED₅₀ of normal and stress chickens

Parameter	Groups	
	Normal (H ₂ O)	Stress (H ₂ O ₂)
Hypnotic ED ₅₀ value = XF + K × D	17.63 mg/kg, IP	16.44 mg/kg, IP
The extent of the doses	15-35 mg/kg	5-35 mg/kg
First dose used	35 mg/kg	35 mg/kg
Last dose used (XF)	25 mg/kg	15 mg/kg
Table Value (K) (Dixon, 1980)	-0.737	0.144
± in the dose (D)	10 mg	10 mg
Chickens used	6 (XXOXOX)*	7 (XXXOOXO)*
OS influence on thiopental hypnotic ED ₅₀ = H ₂ O- H ₂ O ₂ / H ₂ O × 100 = 7%		

*X: effect (i.e. hypnosis), O: no effect (i.e. no hypnosis)

Tab water was given to the normal chickens while 0.5% H₂O₂ was given to the stress group from 1st to the termination of the experiments at day 14th

B. Thiopental's analgesic ED₅₀ value

Thiopental antinociceptive influence in the stressed chickens with H₂O₂ was increased by 18% in association with normal chickens that have been fined by calculation of the analgesic ED₅₀ value of thiopental essential to cause in 50% of the chickens

to be 13.61 and 11.20 mg/kg, IP in the normal and stressed chickens, respectively that resemble the ameliorative influence of OS on the thiopental analgesic ED₅₀ which is reported here for the first time in chicken and Table 1-B exemplify the various results gained from this trial.

Table 1-B. Thiopental's analgesic ED₅₀ of normal and stress chickens

Parameter	Groups	
	Normal (H ₂ O)	Stress (H ₂ O ₂)
Analgesic ED ₅₀ = XF + K × D	13.61 mg/kg, IP	11.20 mg/kg, IP
The extent of the doses used	5-35 mg/kg	5-35 mg/kg
First dose used	35 mg/kg	35 mg/kg
Last dose used (XF)	5 mg/kg	15 mg/kg
Table Value (K) (Dixon, 1980)	0.861	-0.380
± in the dose (D)	10 mg	10 mg
Chickens used	6 (XXOXXO)*	7 (XXXOXXX)*
OS influence on thiopental analgesic ED ₅₀ = Effect of OS= H ₂ O- H ₂ O ₂ / H ₂ O×100= 18%		

*X: result (analgesia), O: no result (no analgesia)

Tab water was given to the normal chickens while 0.5% H₂O₂ was given to the stress group from 1st to the termination of the experiments at day 14th

Nociceptive effect recorded pre and post 15 min as thiopental treatment by using an electro-stimulator device

Thiopental plasma concentration in the normal and stressed chickens at different times

Table 2 and Figure 2 show a significant elevation in the thiopental plasma concentration of the stressed chickens compared to the normal one. The plasma concentration of thiopental measured at time 0.5, 1,

2, 4 and 24 hours after its injection at 35 mg/kg, IP were 91.42, 54.35, 38.27, 22.30 and 7.51 μg/ml in the normal chickens while the plasma concentration was increased by 74, 84, 48, 85 and 82% to be 159.01, 100.06, 56.71, 41.30 and 13.63 μg/ml in the stressed chickens, respectively.

Table 2. Plasma concentration of thiopental in the normal and stressed chickens at different times

Groups	Times (Hour)				
	0.5	1	2	4	24
Normal (H ₂ O) (μg/ml)	91.42 ± 8.60	54.35 ± 4.95	38.27 ± 1.83	22.30 ± 2.12	7.51 ± 0.66
Stressed (H ₂ O ₂) (μg/ml)	159.01 ± 10.19*	100.06 ± 5.54*	56.71 ± 2.85*	41.30 ± 3.90*	13.63 ± 0.75*
OS influence on plasma concentration of thiopental = Effect of OS= H ₂ O ₂ - H ₂ O/ H ₂ O×100= (%)	+74	+84	+48	+85	+82

Data represent Mean ± Standard Error for 5 chickens/measured time

Tab water was given to the normal chickens while 0.5% H₂O₂ was given to the stress group from 1st to the termination of the experiments on day 14th

Thiopental therapy 35 mg/kg, IP for both the normal and stress chickens

*: significantly different from normal group (p < 0.05)

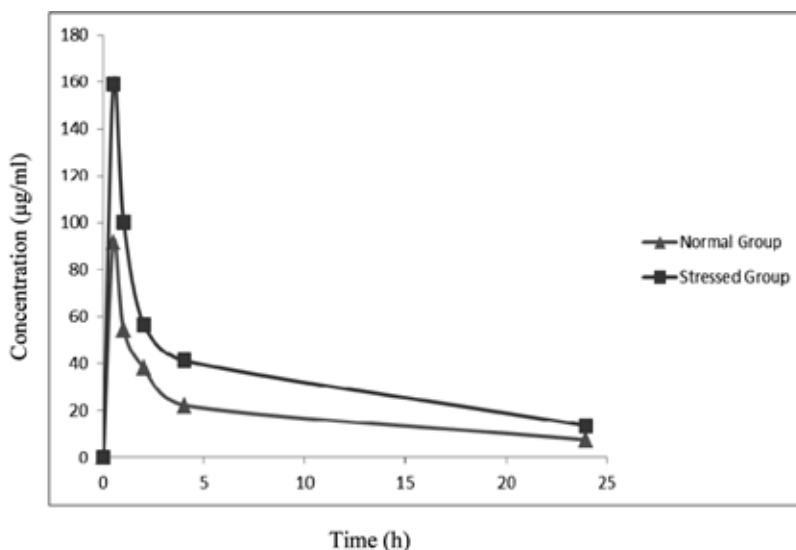


Figure 2. Thiopental's plasma concentration in the normal and stressed chickens at different times

Thiopental pharmacokinetics profiles in the normal and stressed chickens

Thiopental pharmacokinetics profiles in the normal chickens, which include $AUC_{0-\infty}$ (548.93 $\mu\text{g}\cdot\text{h}/\text{ml}$), $AUMC_{0-\infty}$ (8046.58 $\mu\text{g}\cdot\text{h}^2/\text{ml}$), MRT (14.66 h), $t_{1/2\beta}$ (10.49 h), Cl (0.06 L/h/kg), K_{el} (0.07 h^{-1}), V_{ss} (0.97 L/h/kg), T_{max} (0.5 h) and C_{max} (91.42 μg), while these values changed in the stressed chickens to be (999.55 $\mu\text{g}\cdot\text{h}/\text{ml}$), (15573.44 $\mu\text{g}\cdot\text{h}^2/\text{ml}$), (15.58 h), (11.36 h),

(0.04 L/h/kg), (0.06 h^{-1}), (0.57 L/h/kg), (0.5 h) and (159.01 μg), respectively.

Table 3 shows the pharmacokinetic parameters of thiopental ($AUC_{0-\infty}$, $AUMC_{0-\infty}$, MRT and $t_{1/2\beta}$) were increased in the stressed chickens by 82, 94, 6 and 8%, respectively. On another hand, the Cl, K_{el} , and V_{ss} were decreased in the same group by 33, 14 and 41%, respectively in comparison to the normal group.

Table 3. Pharmacokinetics profiles of thiopental in normal and stressed chickens

Pharmacokinetics parameters	Groups		Effect of OS= H_2O_2 - $\text{H}_2\text{O}/\text{H}_2\text{O}\times 100=$ (%)
	Normal (H_2O) group	Stressed (H_2O_2) group	
$AUC_{0-\infty}$ ($\mu\text{g}\cdot\text{h}/\text{ml}$)	548.93	999.55	+82
$AUMC_{0-\infty}$ ($\mu\text{g}\cdot\text{h}^2/\text{ml}$)	8046.58	15573.44	+94
MRT (h)= $AUMC/AUC$	14.66	15.58	+6
$t_{1/2\beta}$ (h)	10.49	11.36	+8
Cl (L/h/kg)= dose/ AUC	0.06	0.04	-33
K_{el} (h^{-1})= $0.693/t_{1/2\beta}$	0.07	0.06	-14
V_{ss} (L/kg)= dose. $AUMC/(AUC)^2$	0.97	0.57	-41
T_{max} (h)	0.50	0.50	0
C_{max} (μg)	91.42	159.01	-74

Tab water was given to the normal chickens while 0.5% H_2O_2 was given to the stress group from 1st to the termination of the experiments at day 14th

Thiopental treatment with dosage of 35 mg/kg, IP for both the normal and stressed chickens

DISCUSSION

The purpose of this study was to use thiopental in chickens for the induction of the general anesthesia and to find out the influence of OS status (induced with H_2O_2) on the thiopental anesthesia, its plasma concentration and pharmacokinetic profile. Thiopental is considered a general anesthetic that stimulates the rapid induction of anesthesia with its increasing the effect of GABA neurotransmitter on the GABA_A receptor which causes depression of the CNS (Clark et al., 2012; Flecknell, 2015; Katzung and Trevor, 2015). Thiopental frequently used as an anesthetic for short surgical operation because it has a good hypnotic effect with occasionally less analgesic and muscle relaxant efficiency (Clark et al., 2012; Katzung and Trevor, 2015) and is considered a safe drug of choice for using in anesthesia because it possesses many benefits including a familiar mechanism of action, less myocardial and cerebral ischemia, decreasing histamine release with a uniquely stable hemodynamic status during anesthesia (Atasoy et al., 1993; Sumitraa et al., 2004; Ninu et al., 2015; Biswas et al., 2017) and was found to have efficient anesthesia than other barbiturates (Ferreira et al., 2013; Shaaban et al., 2018; Brohi et al., 2019). The result of this trial clears up the anesthetic profile of thiopental in the normal and

stressed chickens through determining the analgesic and hypnotic ED_{50} values which were decreased in the stressed chickens with H_2O_2 that revealed an increase in thiopental efficacy and possibly increased its toxicity which was in agreement with earlier studies on diazepam sedation (Mousa and Mohammad, 2012a), xylazine analgesia (Mousa and Mohammad, 2012b) and ketamine anesthesia (Mousa, 2014). Thiopental interact and binds to the GABA_A /chloride ionophore receptor complex, thereby enhancing the inhibitory actions of GABA_A in the CNS and this will leads to synaptic inhibition, diminished neuronal excitability, induce anesthesia and lower the nociceptive threshold thus producing the analgesic effect (NCBI, 2020). Thiopental has a mode of action on the CNS (this attributed to reason for choosing it) because of the CNS is more vulnerable to OS status due to its high free radical formation, consuming a lot of oxygen, high amount of unsaturated fatty acids with less antioxidant activity (Pastore et al., 2003). The results show that thiopental plasma concentration was significantly increased with subsequent altering in its pharmacokinetic profile in the stressed chickens compared with the normal chickens. This modification in the thiopental pharmacological response in the stressed chickens attributed to H_2O_2 -induced OS and thus causes phar-

macokinetics interaction (Mousa, 2014). H_2O_2 -induce OS through its formation of free radicals that break down the vital cell components, especially the protein receptors (Patockova et al., 2003; Sayre et al., 2008), therefore, causes pharmacokinetic interaction by altering the biological constitutes important for absorption, distribution, metabolism and excretion of thiopental that determine the pharmacokinetic profile of thiopental. H_2O_2 -induce OS can alter the pharmacokinetic profile of the drugs through its direct and indirect break down of the cell components (especially proteins which included plasma proteins and protein carriers) and thus the organs (liver, kidney, CNS and so on) which have a simultaneous modification at a level of the absorption, distribution, metabolism and excretion of thiopental and so, leading to pharmacokinetics interaction (Chauhan et al., 2020) which is stated here in this study through measuring the pharmacokinetics profile of thiopental indicated by increasing the half-life and decreasing the clearance, elimination rate constant and distribution volume of thiopental of the stress chickens compared to the nor-

mal one. The pharmacokinetics interaction has an impact to elevate drug efficacy and subsequent toxicity and this criterion of the drug are an important factor ensuring the drug delivery to the site of action on its target receptor to achieve efficacy and/or improve the preferred therapeutic effect (Wen et al., 2015).

CONCLUSIONS

The outcome of this study reveals the increase of the thiopental pharmacological response affected by H_2O_2 -induced OS by increasing its concentration in the plasma and altering the pharmacokinetics profile. We recommended reducing the dose of thiopental to be given to the stressed animals.

ACKNOWLEDGEMENTS

Greetings and thanks to the Veterinary Medicine College, the University of Mosul for its conveniences and materials provided.

CONFLICT OF INTEREST

None declared by the authors.

REFERENCES

- Aksoy L and Alper Y (2019) The effects of royal jelly on oxidative stress and toxicity in tissues induced by malathion, an organophosphate insecticide. *J Hellen Vet Med Soc* 70(2):1517-1524.
- Atasoy N, Mercan N, Atalay C, Bayram E, Tas A (1993) Evaluation of a Mixture of Thiopental-Guafinesine-Metedomidine and Sevoflurane Anesthesia in Horses. *Asian J Anim Vet Adv* 4:191-199.
- Barwick V (2003) Preparation of Calibration Curves A Guide to Best Practice. LGC Ltd, UK, Oxford: pp. 1-27.
- Biswas DS, Hasan M, Mallick S, Juyena NS, Shoriotullah M, Alam MR (2017) Clinical and haematological changes upon administration of Xylazine-Ketamine and Xylazine-Thiopentone anaesthetic combinations in ewes. *Bang Vet* 34(1):9-19.
- Brohi RD, Kalhoro AB, Kachiwal AB, Kalhoro IB, Kalhoro DH, Ahmed S, Khan FA, Talpur HS, Rehman Z, Bhattaria, D (2019) Effect of Propofol and Thiopentone Sodium in Sheep Sedated with Xylazine Hydrochloride. *Pak J Zool* 51(1):1-7.
- Broughton PMG (1956) A rapid ultraviolet spectrophotometric method for the detection, estimation and identification of barbiturates in biological material. *Biochem J* 63:207-213.
- Chauhan VB, Modi CM, Patel UD, Patel HB (2020) The effect of piperine pre-conditioning on the pharmacokinetics of orally administered marbofloxacin in rats. *Vet Arhiv* 90:69-75.
- Church LE (1957) Combuthal as an Anesthetic for Baby Chicks. *Poultry Sci* 36 (4):788-791.
- Clark MA, Finkel R, Rey JA, Whalen K (2012) Lippincott's illustrated reviews: Pharmacology. Williams and Wilkins, USA, Philadelphia: pp. 127-140.
- Dixon WJ (1980) Efficient analysis of experimental observations. *Annu Rev Pharmacol Toxicol* 20:441-462.
- Ferreira TH, Brosnan RJ, Shilo-Benjamini Y, Moore SB, Hollingsworth SR (2013) Effects of ketamine, propofol, or thiopental administration on intraocular pressure and qualities of induction of and recovery from anesthesia in horses. *Am J Vet Res* 74(8):1070-1077.
- Flecknell P (2015) Laboratory animal anesthesia. Elsevier Inc, USA, Academic press: pp. 79-234.
- Garcia PS, Kolesky SE, Jenkins A (2010) General Anesthetic Actions on GABA_A Receptors. *Curr Neuropharmacol* 8(1):2-9.
- Goldbaum LR (1948) An ultraviolet spectrophotometric procedure for the determination of barbiturates. *J Pharmacol Exper Therap* 94(1):68-75.
- Gustafsson LL, Ebling WF, Osaki E, Stanski DR (1996) Quantitation of Depth of Thiopental Anesthesia in the Rat. *Anesthesiol* 84:415-427.
- Kabirian A, Batavani RA, Asri-Rezaei S, Soleimanzadeh A (2018) Comparative study of the protective effects of chicken embryo amniotic fluid, vitamin C and coenzyme Q10 on cyclophosphamide-induced oxidative stress in mice ovaries. *Vet Res Forum* 9:217-224.
- Katz MH (2011) Study design and statistical analysis: A practical guide for clinicians. Cambridge University Press, USA, New York: pp. 14-159.
- Katzung BG, Trevor AJ (2015) Basic and clinical pharmacology. McGraw-Hill Co Inc, USA, New York: pp. 423-438.
- Mousa YJ, Mahmood MB, Mohammad MS (2019) Administration of ketamine with the central and peripheral analgesics for induction of balanced anesthesia in the chicks. *Earth Environm Sci* 388:012021.
- Mousa YJ (2014) Anaesthetic properties of ketamine in chicks stressed with hydrogen peroxide. *Vet Med* 59(8):369-375.
- Mousa YJ (2019) Analgesic, antipyretic and anti-inflammatory efficacy of ketorolac in the chicks. *Ind J Anim Sci* 89(10):1086-1090.
- Mousa YJ (2020) Etomidate anesthesia in chicks: Effect of xylazine. *J Hell Vet Med Soc* 71(4):2463-2470.
- Mousa YJ, Al-Zubaidy MHI (2019) Anesthetic efficacy of ketamine, ketamine-tramadol and ketamine-ketorolac in the chicks. *Iran J Vet Res* 20(1):33-38.
- Mousa YJ, Mohammad FK (2012a) Effects of hydrogen peroxide on diazepam and xylazine sedation in chicks. *Interdiscip Toxicol* 5(4):179-183.
- Mousa YJ, Mohammad FK (2012b) The analgesic efficacy of xylazine and dipyrone in hydrogen peroxide-induced oxidative stress in chicks. *Iraq J Vet Sci* 26(2):69-76.
- NCBI (National Center for Biotechnology Information) (2020) PubChem

- Compound Summary for CID 3000715, Thiopental. Retrieved October 13, 2020 from <https://pubchem.ncbi.nlm.nih.gov/compound/Thiopental>.
- Naseri M, Parham A, Moghimi A (2017) The effect of sodium thiopental as a GABA mimetic drug in neonatal period on expression of GAD65 and GAD67 genes in hippocampus of newborn and adult male rats. *Iran J Basic Med Sci* 20(9):996–1001.
- Ninu AR, Tayal R, Kumar A, Behl SM, Chawla SK (2015) Comparison of thiopentone and ketamine as induction and maintenance agents in buffaloes undergoing diaphragmatic herniorrhaphy. *J App Anim Res* 43(4):462-467.
- Pastore A, Federici G, Bertini E, Piemonte F (2003) Analysis of glutathione: implication in redox and detoxification. *Clin Chim Acta* 333:19-39.
- Patockova J, Marhol P, Tumova E, Krsiak M, Rokyta R, Stipek S, Crkovska J, Andel M (2003) Oxidative stress in the brain tissue of laboratory mice with acute post insulin hypoglycemia. *Physiol Res* 52:131-135.
- Petrie A, Watson P (2013) *Statistics for veterinary and animal sciences*. Blackwell Science, UK, Oxford: pp. 90-140.
- Sayre LM, Perry G, Smith MA (2008) Oxidative stress and neurotoxicity. *Chem Res Toxicol* 21:172-188.
- Shaaban MF, Kassem MM, EL Kammar MH, Korittum SA (2018) Clinical Evaluation of Intravenous Propofol Alone or in Combination With Diazepam, Ketamine HCl and Thiopental Sodium to Induce General Anaesthesia in Dogs. *Alex J Vet Sci* 57(1):106-114.
- Sumitraa M, Manikandana P, Kuppu-Raob KV, Nayeemc M, Manohard BM, Puvanakrishnan R (2004) Cardiorespiratory effects of diazepam-ketamine, xylazine-ketamine and thiopentone anesthesia in male Wistar rats-A comparative analysis. *Life Sci* 75:1887-1896.
- Wen H, Jung H, Li X (2015) Drug Delivery Approaches in Addressing Clinical Pharmacology-Related Issues: Opportunities and Challenges. *Am Assoc Pharmaceut Sci J* 17(6):1327-1340.
- Zhang Y, Huo M, Zhou J, Xie S (2010) PKSolver: An add-in program for pharmacokinetic and pharmacodynamic data analysis in Microsoft Excel. *Comp Meth Prog Biomed* 99(3):306-314.