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A Ismail, A Deeb, I Alhawary, W Elkassas, Y Hegazy

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Influence of lysozyme utilization with lactic acid bacteria in yoghurt on some foodborne pathogens

A.M. Ismail¹, A.M.M. Deeb², I.I. Alhawary^{2,3}, W.M. Elkassas¹, Y.M. Hegazy^{4*}

¹Department of Food Hygiene, Animal Health Research Institute, Kafrelshiekh Lab, Agriculture Research Center (ARC), Egypt

² Food control department, Faculty of Veterinary Medicine, Kafrelsheikh Univeristy, Egypt

³ Fish Processing and Biotechnology, Dean of faculty of Aquatic and Fisheries Science, Kafrelsheikh Univeristy, Egypt

⁴ Department of Animal medicine (Infectious Diseases), Faculty of Veterinary Medicine, University of Kafrelsheikh, 5-Elgeish street, Kafrelsheikh, Egypt

ABSTRACT: Despite the existence of many different food preservation methods, foodborne pathogens are still the most common related problems to dairy products. Developing alternative natural methods to control such problems is necessary. This research was planned to assess the bacteriological quality of yoghurt samples commercially available in Kafr El-Sheikh city, Egypt. Also to study the impact of natural antibacterial agents on the inhibition of selected foodborne pathogens artificially inoculated in yoghurt samples. The obtained results revealed poor bacteriological quality of the examined samples, as total staphylococci and coliforms were detected in 80% and 90% yoghurt samples, with mean values of $2.4 \times 10^3 \pm 0.13 \times 10^3$ CFU/g and $3.6 \times 10^5 \pm 2.4 \times 10^5$ MPN/g, respectively. *Staphylococcus aureus* was noticed in 4% of examined samples, polymerase chain reaction (PCR) confirmed the presence of virulence gene (clumping factor A- clfA) in one isolate. *Escherichiacoli* was detected in 14% of yoghurt samples, then serologically identified as O146:H5 (2), O111:H2 (2), O125:H11 (1), O26:H10 (1), and O158:H7 (1). Artificially inoculated yoghurt with *S. aureus* (7.39 log₁₀ CFU/g) and *E. coli* (8.3 log₁₀ CFU/g), were used to investigate the ability of hen egg-white lysozyme either with LAB classic or with ABT-5 (*L. acidophilus* LA5+ *S. thermophiles* + *Bifidobacterium Bb12*) starter cultures to inhibit the growth of these pathogens. During refrigerated storage, all yoghurt batches showed a reduction in pH value. It was observed that both pathogens couldn't be detected in the yoghurt batch containing LAB classic starter + heated lysozyme. Furthermore, *S. aureus* was completely inhibited in that containing ABT-5 cultures alone, while *E. coli* decreased by 6.23log. Conversely, there was not complete inhibition in other batches containing (LAB classic starter, LAB classic starter+ native lysozyme, ABT-5 culture+ native lysozyme and ABT-5 culture+ heated lysozyme) as the reduction rate during all storage periods for *S. aureus* count was 1.66, 2.63, 4.47, and 4.19 log₁₀ CFU/g, While for *E. coli* count was 3.3, 3.81, 5.53, and 4.89 log₁₀ CFU/g, respectively. This study highlighted the importance of adding natural antibacterial agents like lysozyme with LAB culture to yoghurt as a bio-control strategy to overcome foodborne pathogens that cause a public health hazard.

Keywords: Hen Egg White Lysozyme; ABT-5; *S. aureus*; clfA; *E. coli*

Corresponding Author:
Yamen M. Hegazy, Fac. Vet. Med. Kafrelsheikh University Egypt
E-mail address: yamen12@yahoo.com

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INTRODUCTION

Yoghurt is considered the most generally consumed fermented dairy product all over the world due to its high nutritive value and therapeutic properties (He et al., 2005). Although fermented dairy products are commonly considered to be safe, due to their acidic nature, inadequate hygienic conditions during processing, manipulation, and distribution decrease the quality of these products and increase the microbiological load which constitutes a public health hazard (Karagozlu et al., 2007). Foodborne pathogens like *S. aureus* and *E. coli* are the most commonly predominant human pathogenic bacteria associated with such low-quality fermented milk (Cancino-Padilla et al., 2017).

As a result of the health problems resulting from using artificial preservatives in dairy products, there is an urgent need for using naturally produced antimicrobial preservatives as alternatives for chemical ones without any adverse effects on public health to extend the product's shelf life and control food pathogens in refrigerated foods. Natural antimicrobial agents could be obtained from natural sources like animals, plants, and microbial sources (Pisoschi et al., 2018).

Hen egg-white lysozyme (LZ) is the most important natural antibacterial enzyme of animal origin that consists of 129 amino acids with a molecular weight of 14.3 kDa and has 4 disulfide bonds responsible for its high thermal stability (Carrillo et al., 2014). It is classified as Generally Recognized As Safe (GRAS) by the U.S. Food and Drug Administration (FDA). It could be very appropriate for milk products bio-preservation because of its absorption by casein micelles without loss in its enzymatic activity after absorption (L. de Roos et al., 1998), plus its stability in a wide range of temperature and pH (Abdou et al., 2013).

LZ breakdown the bacterial cell wall of susceptible Gram-positive bacteria by catalyzing the hydrolysis of β (1-4) glycosidic linkages between N-acetylmuramic acid and N-acetylglucosamine found in peptidoglycan (the major component of the bacterial cell wall) (Arabski et al., 2015). However, its limited antibacterial effect against Gram-negative bacteria due to the presence of an additional barrier consist of protein, phospholipids, and lipopolysaccharide (LPS) surrounding the peptidoglycan layer (Mann, 2017) can be improved by modifying its hydrophobicity and cationic charge. Thermal modification caused by increasing temperatures results in a significant reduction in the enzymatic activity, but greatly improved

its bactericidal action against Gram-negative bacteria (Ibrahim et al., 1996).

Likewise, Lactic Acid Bacteria (LAB) are effective natural antimicrobial agents of microbial origin that are identified as protective cultures. They have antagonistic action against some foodborne pathogenic microorganisms without adverse effect on the sensory quality of the product (Katikou et al., 2005). ABT-5 starter cultures are probiotic mixed cultures of LAB that consisting of both thermophilic (mainly *Streptococcus thermophilus*) and mesophilic strains (such as *Bifidobacterium* and *Lactobacillus acidophilus*) have been commonly used in commercial fermented dairy products today (Buriti et al., 2007).

Probiotics improve human health state, in addition to their antagonistic effect against foodborne pathogens (Yesillik et al., 2011). Its antimicrobial properties are due to the production of inhibitory substances such as bacteriocins, organic acids (lactic acid and acetic acid), H_2O_2 , and hydroxylated fatty acids (Russo et al., 2017).

The current study was planned to evaluate yoghurt commercially available in Kafr El-Sheikh city bacteriologically as well as investigate the antibacterial activity of native and modified LZ, for the first time up to our knowledge, either with LAB classic starters or in combination with a probiotic culture against *S. aureus* and *E. coli* in manufactured yoghurt.

MATERIALS AND METHODS

Samples collection and preparation

Fifty yoghurt samples were randomly collected from different supermarkets in Kafr El-Sheikh city, Egypt. All samples were transported quickly to the laboratory in an insulated ice-box for bacteriological examination. Serial dilutions (10-fold) were prepared from these samples using 0.1 % sterile peptone water (Oxoid) and suitable dilutions were used for staphylococcal and coliforms counts (ISO 6887-2: 2003).

Bacteriological examination

Total staphylococci count and isolation of *S. aureus* (ISO 6888-1: 2003)

The staphylococcal count was carried on Baird Parker agar medium plates (Oxoid) that incubated at 37°C for 48 hrs. Circular, smooth or rough appearance, convex, gray-black colonies were enumerated and the total staphylococci count /g was calculated.

Then, typical colonies of *S. aureus* (dark convex colonies with a thin white margin that bounded by a clear area spreading into the opaque medium) were picked and cultured on a nutrient semisolid medium for biochemical identification using the following tests: catalase, hemolysis, mannitol, oxidase and coagulase test according to (Quinn et al., 2002).

PCR was used for the detection of virulence adhesion gene (clumping factor A- *clfA*) in *S. aureus* at Animal Health Research Institute, Dokki, Egypt. The extraction of bacterial DNA from *S. aureus* isolates was carried out according to QIAamp DNA mini kit instructions using specific primers (Table 1). PCR Master Mix (total volume 25 µl) according to Emerald Amp GT PCR mastermix (Takara) Code No. RR310A kit was used. This mix included 12.5 µl Emerald Amp GT PCR mastermix (2x premix), 4.5 µl PCR grade water, 6 µl Template DNA, 1 µl from each forward and reverse primer (20 pmol). The cycling conditions were: primary denaturation at 94°C for 5 min, followed by secondary denaturation (35 cycles) at 94°C for 30 sec, annealing at 55°C for 40 sec, extension at 72°C for 45 sec, then final extension at 72°C for 10 min. About 20 µl of each PCR product was used in gel electrophoresis according to (Sambrook et al., 1989) with modification. A 100 bp DNA ladder was used as a marker for PCR product. This gel contained 1.5% agarose gel and 0.5 µg/ml ethidium bromide (for staining). The gel was photographed by a gel documentation system and the data was analyzed through computer software.

Total coliform count and isolation of *E. coli*

Coliforms count was carried out using the Most Probable Number (MPN/g) (ISO 4831: 2006). A loopful from positive MacConkey's broth tubes (yellow color with gas production) was streaked onto Eosin Methylene Blue agar plates (EMB) (Oxoid) then incubated at 37 °C for 24 hrs (APHA, 2004). The purified colonies were cultured on a nutrient semisolid medium for biochemical identification using the following tests: motility, indole production, methyl red, Voges - Proskauer, citrate utilization, hydrogen-sulphide production, urease, nitrate reduction, oxidation-fermentation test and fermentation of sugars ac-

ording to (MacFaddin, 2000).

The suspected *E. coli* strains were serologically confirmed at Animal Health Research Institute, Dokki, Egypt, by using rapid diagnostic *E. coli* antisera sets (Denka Seiken Co., Japan) according to (Kok et al., 1996).

Effect of natural antibacterial agents on survival of *S. aureus* and *E. coli* in manufactured yoghurt

Preparations of antibacterial agents

Starter cultures used in yoghurt manufacture

The LAB classic starter (YC-X11) containing *S. thermophilus* and *L. delbrueckii ssp. Bulgaricus* and the probiotic culture ABT-5 consisting of *L. acidophilus LA5*, *S. thermophilus*, and *Bifidobacterium Bb12* (Christian Hansen, Denmark) were used.

Lysozyme solution

Hen egg-white lysozyme (Tokyo Chemical Industry CO., LTD. 6-15-9 Toshima, KITA-KV Tokyo, Japan) stock solution of 1mg/ml in potassium phosphate buffer (10 mM, pH 6) was prepared according to (Masschalck et al., 2000), then divided into two parts, one part was kept refrigerated at 4°C and the other part was modified thermally by heating at 80°C for 20min, then insoluble aggregates were removed by centrifugation (3000×g, 15 min) (Ibrahim et al., 1996). The final concentration of native lysozyme (LZ) and heated lysozyme (HLZ) was 50 µg/ml. The enzymatic activity of LZ was measured against *Micrococcus lysodeketicus* by a turbidimetric method using a spectrophotometer (Touch et al., 2004).

Bacterial test strains

S. aureus and *E. coli* O₁₁₁:H₂ that were previously isolated in this study and subjected for confirmation were selected according to their virulence and serotype, respectively, then cultivated in sterile nutrient broth at 37°C for 24 hrs. The bacterial population was determined by the pour plate method after preparation of 10- fold serial dilutions to reach the level of 2.5×10⁷ (7.39 log₁₀ CFU/ml for *S. aureus*) and 2×10⁸ (8.3 log₁₀ CFU/ml for *E. coli*) CFU/ml.

Table 1. Oligonucleotide primers sequences (Metabion, Germany)

Gene	Sequence	Amplified product	Reference
<i>S. aureus</i>	GCAAAATCCAGCACAAACAGGAAACGA	638 bp	Mason et al.(2001)
<i>clfA</i>	CTTGATCTCCAGCCATAATTGGTGG		

Table 2. Bacterial load of the examined yoghurt samples

Tested microorganisms	No. of examined samples	Positive samples		Count /g		
		No.	%	Min.	Max.	Mean± SE
Staphylococcal count	50	40	80	1.0×10^1	5.6×10^4	$2.4 \times 10^3 \pm 0.13 \times 10^3$
<i>S. aureus</i>		2	4			
Coliforms count		45	90	7.4	1.1×10^7	$3.6 \times 10^5 \pm 2.4 \times 10^5$
<i>E. coli</i>		7	14			

Treatments

Yoghurt was manufactured according to (Robinson et al., 2002); raw cow's milk was heated in a water bath at 90 °C with agitation for 10 min then cooled to 42 °C. Then, it was divided into six batches (8 plastic cups for each batch) as follow:

batch 1: contained 2 % (v/v) LAB classic starter cultures (YC-X11-Yoflex®)

batch 2: contained 2 % (v/v) LAB classic starter + LZ solution

batch 3: contained 2 % (v/v) LAB classic starter + HLZ solution

batch 4: contained 2 % (v/v) ABT-5 culture

batch 5: contained 2 % (v/v) ABT-5 culture + LZ solution

batch 6: contained 2 % (v/v) ABT-5 culture + HLZ solution

Every batch was doubled, one inoculated with *S. aureus* (48 plastic cups) and the other inoculated with *E. coli* (48 plastic cups) at the rate of 7.39 and 8.3 log₁₀ CFU/g, respectively. All batches were incubated at 45 °C until yoghurt curd (50g yoghurt in each cup) was formed. After curdling, the batches were kept at 4 °C for 14 days. Samples were examined immediately after fermentation (curd time) and at 2nd, 4th, 6th, 8th, 10th, 12th, and 14th days of storage period to determine pH values using a pH meter (Jenway, UK) and evaluate the inhibitory effect of these natural antibacterial agents against both pathogens. The colony-forming units count (CFU/g) was calculated by culturing onto the specific media for each pathogen.

Statistical Analysis

Statistical analysis was performed using SPSS Version 16 software (SPSS, IL, USA). Data were analyzed using the General Linear model, Repeated Measures ANOVA to compare the significant effect of different treatment groups on pH of the yoghurt sam-

ples and also on the total *S. aureus* and *E. coli* count after 14 days of storage. Duncan's Multiple Range Test (DMRT) was used to determine significant differences between mean values. The alpha level for determination of significance was set at 0.05.

RESULTS

The results exhibited that staphylococci were detected in (40) 80% of the examined samples (Table 2) with counts ranged from 1.0×10^1 to 5.6×10^4 CFU/g. *S. aureus* was isolated from (2) 4 % of the examined samples. The result obtained from PCR (photo 1) recovered that the *clfA* virulence gene was detected in one studied strain of 638 pb.

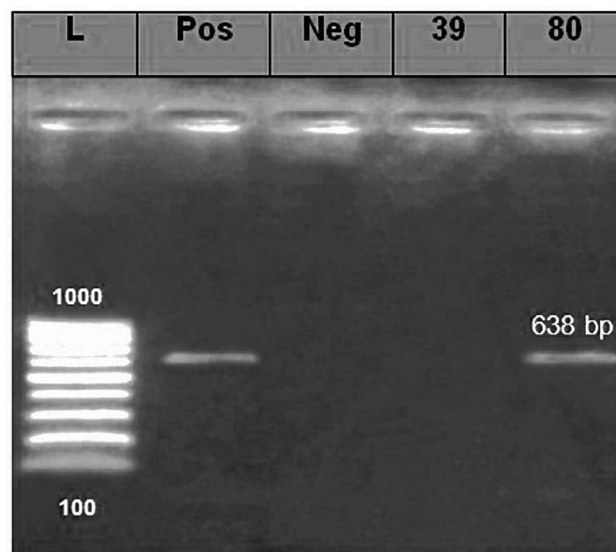
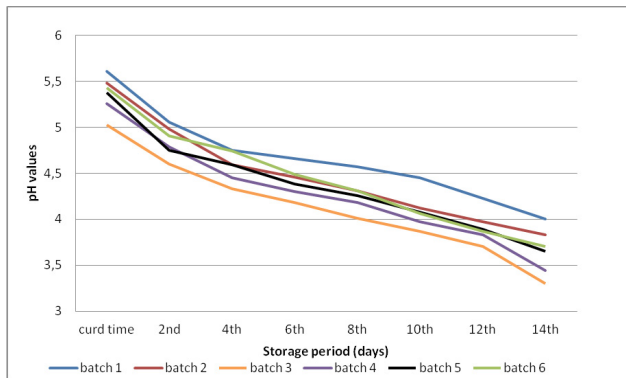


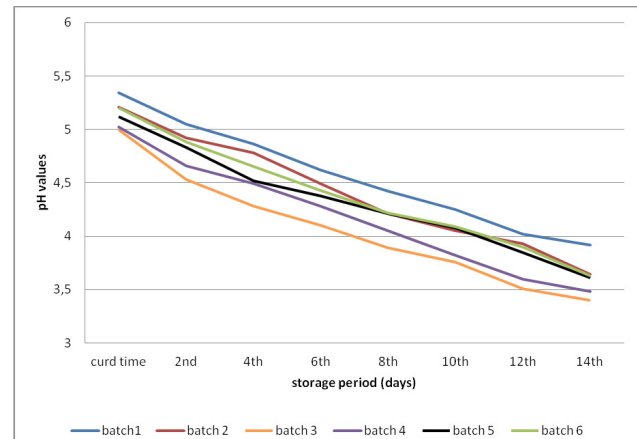
Photo 1. Agarose gel electrophoresis of cPCR amplified of *S. aureus* *clfA* gene. L: 100bp DNA ladder, Pos: control positive for *clfA* gene, Neg: control negative

Coliforms were detected in 45 (90%) of examined yoghurt samples with counts varied between 7.4 to 1.1×10^7 MPN/g. Whereas, *E. coli* was isolated from 7 (14%) of the examined samples (Table 2). The isolated *E. coli* strains were serologically identified to serogroups as follows O₁₄₆:H₅ (2), O₁₁₁:H₂ (2), O₁₂₅:H₁₁ (1), O₂₆:H₁₀ (1), and O₁₅₈:H₇ (1) (Table 3).



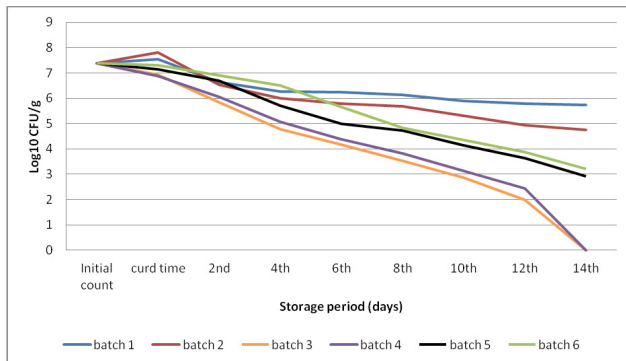
batch 1: LAB classic starter cultures batch 2: LAB classic starter + LZ
 batch 3: LAB classic starter + HLZ batch 4: ABT-5 culture
 batch 5: ABT-5 culture+ LZ batch 6: ABT-5 culture + HLZ

Figure 1. Changes in pH values in yoghurt batches inoculated with *S. aureus* during refrigerated storage (4°C)



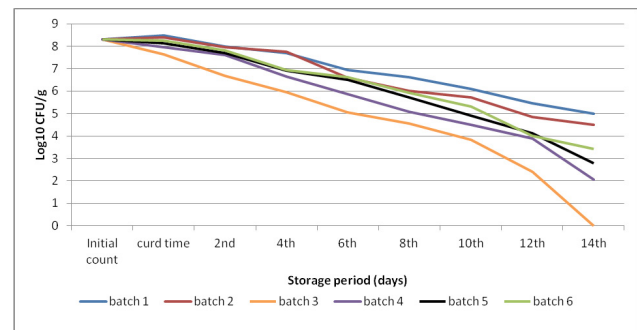
batch 1: LAB classic starter cultures batch 2: LAB classic starter + LZ
 batch 3: LAB classic starter + HLZ batch 4: ABT-5 culture
 batch 5: ABT-5 culture+ LZ batch 6: ABT-5 culture + HLZ

Figure 2. Changes in pH values in yoghurt batches inoculated with *E. coli* during refrigerated storage (4°C)



batch 1: LAB classic starter cultures batch 2: LAB classic starter + LZ
 batch 3: LAB classic starter + HLZ batch 4: ABT-5 culture
 batch 5: ABT-5 culture+ LZ batch 6: ABT-5 culture + HLZ

Figure 3. Behaviour of *S. aureus* in yoghurt batches during refrigerated storage (4°C)



batch 1: LAB classic starter cultures batch 2: LAB classic starter + LZ
 batch 3: LAB classic starter + HLZ batch 4: ABT-5 culture
 batch 5: ABT-5 culture+ LZ batch 6: ABT-5 culture + HLZ

Figure 4. Behavior of *E. coli* in yoghurt batches during refrigerated storage (4°C)

Table 3. Serological identification of the isolated *E. coli* from the examined yoghurt samples

<i>E. coli</i> serogroups	No.
O ₁₄₆ :H ₅	2
O ₁₁₁ :H ₂	2
O ₁₂₅ :H ₁₁	1
O ₂₆ :H ₁₀	1
O ₁₅₈ :H ₇	1

Figures (1-2) display the pH changes in yoghurt batches inoculated by *S. aureus* and *E. coli* during refrigerated storage. The pH values gradually decreased ($p < 0.05$) as the storage period extends from initial values at the termination of fermentation (curd

time) 5.61, 5.48, 5.03, 5.26, 5.38, and 5.43 to 4.00, 3.83, 3.30, 3.44, 3.65, and 3.70 at the day 14 of storage period (4 °C), respectively, for yoghurt batches inoculated by *S. aureus*. And from 5.34, 5.21, 5, 5.02, 5.12, and 5.2 initially to 3.92, 3.64, 3.40, 3.48, 3.61, and 3.63 at the end of the storage period for batches inoculated by *E. coli*, respectively.

Figure (3) showed viable counts of *S. aureus* in all batches decreased significantly ($p < 0.05$) as the storage period extended. The counts decreased from the inoculation count (7.39 log₁₀ CFU/g) to 5.73, 4.76, 2.92, and 3.2 log₁₀CFU/g in batches (1, 2, 5 & 6), respectively. Whereas, *S. aureus* growth in yoghurt batch 3 that containing HLZ with LAB classic starter and

batch 4 with ABT-5 culture alone couldn't be detected on the 14th day of refrigerated storage.

The survival pattern of *E. coli* in yoghurt batches (Figure 4) showed that the viable counts of *E. coli* in all batches decreased significantly ($p < 0.05$) during the refrigerated storage period. The counts decreased from the initial count (8.3 log₁₀CFU/g) to 5, 4.49, 2.07, 2.77, and 3.41 log₁₀CFU/g in batches (1, 2, 4, 5 & 6), respectively. However, *E. coli* growth was not detected on day 14th of refrigerated storage only in yoghurt batch 3 (LAB classic starter + HLZ).

As regards the result tabulated in (Table 4), the hydrolytic activity of LZ and HLZ was measured against *Micrococcus lysodeikticus* by spectrophotometric method. Enzymatic activity of denatured LZ decreased to 52.31% with about 47.69% loss in hydrolytic activity.

Table 4. Hydrolytic activity of lysozyme

lysozyme activity (Units / mg)		
LZ	HLZ	Reduction %
2.225	1.164	47.69

DISCUSSION

In the current study, staphylococci were detected in 80% of examined yoghurt samples found in the markets and ready to be used by consumers with a mean count of $2.4 \times 10^3 \pm 0.13 \times 10^3$ CFU/g. Yabaya and Idris (2012) found a lower staphylococcal count in collected yoghurt samples from different retail outlets within Kaduna Metropolis; 2×10^2 to 7×10^2 CFU/g. Conversely, higher staphylococcal counts were obtained by Omar et al. (2018); 8.3×10^5 to 1.4×10^7 CFU/g in collected yoghurt samples from different regions in Sharkia governorate, Egypt. The presence of staphylococci in yoghurt usually indicates improper hygienic conditions during manufacturing, handling, and storage, as contamination may be originated from using mastitic milk and food handlers with unclean hands or respiratory infection (Akram et al., 2013).

S. aureus was detected in 4 % of examined yoghurt samples. These samples were incompatible with the permissible limits reported by Egyptian standards (EOSQC, 2005), which stated that yoghurt must be free from *S. aureus*. Kandil et al. (2018), Meshref et al. (2019), Abd El-Halem et al. (2019), and Younis et al. (2020) isolated *S. aureus* from 36%, 88%, 5.7%, and 52 % of yoghurt samples collected from different governorates in Egypt, respectively. This varia-

tion in *S. aureus* incidence rates between studies may refer to the variation in the manufacturing practices and personal hygiene followed during yoghurt production. The presence of *S. aureus* in yoghurt even in low incidence must be considered a public health hazard, as the organism may lose its viability while its enterotoxins remain to elicit staphylococcal food poisoning symptoms (Erkmen, 1995). In the current study, the presence of virulence factor (clfA) in one of the two *S. aureus* isolates that considered one of the essential adhesion factors that responsible for the clumping of blood plasma protein fibrinogen allowing the organism to circulates in the blood and forms a biofilm (Vaudaux et al., 1995), also it has been identified as a virulence factor in an endocarditis case (Heilmann, 2011).

Coliforms are a group of Gram-negative bacteria of public health concern that frequently contaminate dairy products through several sources causing food illnesses (Giammanco et al., 2011). According to EOSQC (2005), coliforms count in yoghurt should be less than 10 CFU/g, in this study 86% of yoghurt samples were failed to comply with this permissible limit. Previous Egyptian studies detected coliforms at different rates as 50%, 76.67% of yoghurt samples examined by El-Leboudy et al. (2017), Fathi et al. (2019), respectively. High levels of coliforms in yoghurt may not only indicate fecal contamination but also give an indication of using low-quality raw milk, improper heat treatment, unhygienic manufacturing practices or contamination after processing, poor personnel hygiene, and using of unclean equipment and/or contaminated water (El-Kholiy et al., 2014).

E. coli is used as a marker organism for fecal contamination and reflects a possible presence of other enteric pathogens and/or toxigenic organisms which constitute a public health threat (Singh and Prakash, 2008). *E. coli* was isolated from 14% of examined yoghurt samples. These samples were failed to comply with EOSQC (2005) as yoghurt must be free from *E. coli*. The contamination rate of *E. coli* in our examined samples was less than those reported by Fathi et al. (2019), Younis et al. (2020), and Hassan et al. (2021) who detect it in 36.96%, 80%, and 25%, respectively, in their examined yoghurt samples collected from different localities in Egypt, while Sayed (2012) couldn't isolate *E. coli*. The different prevalence rates between studies may indicate poor hygienic conditions during yoghurt manufacturing and handling, as its presence in yoghurt referred to post pasteurization contami-

nation either before or during the packaging process (Omola et al., 2014).

The relatively high incidence of *E. coli* in yoghurt samples reported in the present and previous studies seems to confirm the survival of *E. coli* in both low pH and temperature and, subsequently, the implication of acidic foods as yoghurt in food poisoning diseases caused by *E. coli* (Sharp et al., 1995). The isolated *E. coli* strains were serotyped to O₁₄₆:H₅ (2), O₁₁₁:H₂ (2), O₁₂₅:H₁₁ (1), O₂₆:H₁₀ (1), and O₁₅₈:H₇ (1), most of these serogroups are commonly involved in food poisoning sporadic and outbreak cases all over the world (Valilis et al., 2018).

The effect of natural antibacterial agents on the survival of *S. aureus* and *E. coli* in manufactured yoghurt was studied. It was observed that pH values of yoghurt batches containing LZ and HLZ with LAB classic starter culture were slightly lower than that containing LAB classic starter culture alone, this reduction may be attributed to increasing the bacterial activity of starter culture that fermented lactose leading to more lactic acid production (Hofi and El-Shibiny, 1978). Also, this confirmed that starter culture (*S. thermophilus* and *L. Bulgaricus*) don't affected by the antibacterial activity of LZ and agreed with Vindero-la et al. (2002), Turchia et al. (2017), but the mechanism of this resistance needs more investigations. Moreover, probiotic yoghurt batches showed a slight decrease in pH compared to that with LAB classic starter culture alone, this drop in pH is due to probiotics not only produce lactic acid but also acetic acids resulting in low pH of yoghurt (Mobarez et al., 2008).

S. aureus was not detected in both yoghurt batches containing HLZ with LAB classic starter and ABT-5 culture alone at the end of cold storage. While the lowest reduction was belonging to batch 1 with LAB classic starter culture alone.

The bactericidal effect of yoghurt batch containing LAB classic starter with HLZ was significantly higher ($p < 0.05$) than those with LZ, this may be attributed to heat denaturation modifying native LZ (monomeric form) to oligomeric forms resulting in broadens its antibacterial activity to include some Gram-positive bacteria that resist to native LZ (Cegielska-Radziejewska et al., 2009). As, *S. aureus* carries an O-acetylation on the C-6 hydroxyl group of the N-acetylmuramyl residues of its peptidoglycan layer by the activity of integral membrane protein, OatA gene resulting in its resistance to native LZ (Bera et al., 2007).

Also, it worth to be mention that the survival of *S. aureus* were significantly lower ($p < 0.05$) in yoghurt batches with probiotic culture compared to yoghurt batch with classic starter culture alone that may be due to probiotics produce metabolites with antagonistic properties including organic acids, carbon dioxide, bacteriocins, H₂O₂, and low molecular weight antimicrobial compounds (Tejero-Sariñena et al., 2012).

Al-Delaimy and Hamamdeh (2013) and Abd El-Gawad et al. (2014) recorded that *S. aureus* was inhibited after the 10th day of cold storage in the probiotic yoghurt, while the bacteria were able to survive in traditional yoghurt sample after 15 days of storage period. While, Soliman and Ahmed (2019) stated a slight decrease in the number of *S. aureus* after 7 days of storage. Conversely, Saleem et al. (2021) reported a significant inhibitory effect of LZ on *S. aureus* when added to mozzarella cheese.

Our findings also showed that the yoghurt batch containing HLZ with LAB classic starter culture gave the best bactericidal effect toward *E. coli* it couldn't be detected at end of the storage period.

The markedly inhibitory effect of HLZ ($p < 0.05$) compared to LZ with LAB classic starter may be attributed to thermal modification leading to the partial unfolding of native LZ extending its antibacterial activity to include Gram-negative bacteria by penetrating the lipid layer of the bacterial cell membrane forming ion pores resulting in disturbance in the electrochemical potential (Leśnierowski and Cegielska-Radziejewska, 2012).

Additionally, it was noticed that the reduction in *E. coli* count were significantly higher ($p < 0.05$) in probiotic yoghurt batches than that with LAB classic starter culture alone. This inhibitory effect could be due to probiotics produce antibacterial substances.

A previous study by Vilcacundo et al. (2018) recorded that thermal modification of LZ at 80°C for 20 min was able to decrease the log of *E. coli*. On the other hand, Abd El-Gawad et al. (2014) revealed that 3.5 × 10⁵ CFU/g of *E. coli* were eliminated after 2 and 5 days of refrigerated storage in probiotic and traditional yoghurt samples, respectively.

The hydrolytic activity of HLZ decreased to 52.31%. This result disagreed with those of Thammasirirak et al. (2010), Vilcacundo et al. (2018) who reported that heated lysozyme at 80°C for 20 min was with 74.5 and 95% lytic activity, respectively. Previ-

ous studies by Ibrahim et al.(2001), Gorbenko et al. (2007) have confirmed the fact that the antibacterial activity of thermal modified LZ is not dependent on its enzymatic activity. As, LZ has two main additional non-enzymatic bacteriolytic mechanisms of action, which are not dependent on its enzymatic function (the indirect bacteriolytic-dependent mechanisms and electrostatic communication mechanism with bacterial oligonucleotides) (Abdou et al., 2013; Arabski et al., 2015).

Combined use of HLZ with probiotics showed better antimicrobial effect against both pathogens than using LAB classic starter alone. Conversely, using probiotic culture alone had better antibacterial effect against pathogens than its using in combination with HLZ. This indicates that probiotic culture alone gave the most potent inhibitory activity on pathogens than using mixture of probiotic culture with HLZ and using LAB classic starter alone. It could be expected that LZ had an antagonistic effect either on bifidobacteria that found in ABT-culture and/or its bacteriocin. Kim et al. (2015) reported that the inhibitory activity of bacteriocins produced by probiotics could be antagonized by enzymatic degradation. Also, the susceptibility of bifidobacteria to LZ was strain-specific (Rada et al., 2010). For this reason, the resistance to LZ at 25-50 mg/l should be commended as a standard for the selection of probiotic strains suitable for use in

the milk industry, as, LZ can be used as milk products preservatives at this concentration (Cardarelli et al., 2007).

CONCLUSION

Results of this study clearly indicate that the bacterial quality of locally produced yoghurt in Kafr El-Sheikh city, Egypt was inferior as compared to the Egyptian Standards. Therefore strict sanitary measures must be applied during manufacturing, storage, and distribution to produce safe and high-grade yoghurt. Moreover, our experimental results showed that the combination of HLZ with LAB classic starter exhibited a great bactericidal effect on both *S. aureus* and *E. coli* than its combination with probiotic culture. Also, using probiotic culture alone gave the most potent inhibitory activity on pathogens than using mixture of probiotic culture with HLZ and using LAB classic starter alone. So, we suggest adding these natural antimicrobials as food bio-controlling preservatives.

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CONFLICT OF INTEREST

None declared by the authors.

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