

Journal of the Hellenic Veterinary Medical Society

Vol 73, No 4 (2022)



Effects of Beta Vinasse Supplementation on Performance, Meat Quality and Ilio- Caecal Microflora in Quail Rations

DY Yeşilbağ, I Cetin, SS Cengiz, E Cetin, M Sarpasar

doi: [10.12681/jhvms.28016](https://doi.org/10.12681/jhvms.28016)

Copyright © 2023, Derya Yesilbag Yeşilbağ



This work is licensed under a [Creative Commons Attribution-NonCommercial 4.0](https://creativecommons.org/licenses/by-nc/4.0/).

To cite this article:

Yeşilbağ, D., Cetin, I., Cengiz, S., Cetin, E., & Sarpasar, M. (2023). Effects of Beta Vinasse Supplementation on Performance, Meat Quality and Ilio- Caecal Microflora in Quail Rations. *Journal of the Hellenic Veterinary Medical Society*, 73(4), 4847–4852. <https://doi.org/10.12681/jhvms.28016>

Effects of Beta Vinasse Supplementation on Performance, Meat Quality and Ilio-Caecal Microflora in Quail Rations

I. Cetin¹, D. Yesilbag^{*2}, S.S. Cengiz³, E. Cetin⁴, M. Sarpasar⁵

¹Department of Animal Nutrition and Nutritional Diseases, Faculty of Veterinary Medicine, Tekirdag Namik Kemal University, Tekirdag, Turkey

^{2,3,5}Department of Animal Nutrition and Nutritional Diseases, Faculty of Veterinary Medicine, Bursa Uludag University, Bursa, Turkey

⁴Department of Food Hygiene and Technology, Faculty of Veterinary Medicine, Tekirdag Namik Kemal University, Tekirdag, Turkey

ABSTRACT: The aim of this study was to evaluate the effects of Beta (β)-vinasse supplementation on the live weight (LW), live weight gain (LWG), feed intake (FI), feed conversion ratio (FCR), carcass yield, meat quality, and ilio-caecal bacteriological flora of quails. A total of 240 5-d-old Japanese (*Coturnix coturnix Japonica*) quails, including both males and females, were divided into 3 groups 80 quails and treated as follow: a control group (1) with 0 g β -vinasse/kg ration; (2) 15 g/kg β -vinasse and (3) 30 g/kg β -vinasse. The study lasted for 6 weeks. At the end of the experiment, supplementation with β -vinasse did not have a significant effect on FI and FCR. Dietary supplementation with 30 g/kg β -vinasse significantly ($P < 0.05$) increased LW (21 d) and LWG (5 to 21 d). The dietary treatment of quails with different levels of β -vinasse did not affect hot carcass weight, cold carcass weights, hot and cold carcass yields, and breast and thigh pH. The lightness (L^*) and yellowness (b^*) of breast values were significantly ($P < 0.05$) increased by 30 g/kg supplementation. Different levels of β -vinasse significantly increased *Lactobacillus* spp. in faeces. As a result, it was concluded that β -vinasse (by-product obtained from molasses) can be used in quail diets as an alternative feed source that will meet the nutritional needs of the animal and have positive effects on the digestive system, especially on the intestinal health (an increase in *Lactobacillus* spp. counts).

Keywords: betaine; β -vinasse; *Lactobacillus* spp.; performance; quails

Corresponding Author:

Derya Yesilbag, Department of Animal Nutrition and Nutritional Diseases, Faculty of Veterinary Medicine, Bursa Uludag University, 16059 Bursa, Turkey
E-mail address: dyesilbag@uludag.edu.tr

Date of initial submission: 13-09-2021
Date of acceptance: 22-09-2022

INTRODUCTION

In livestock nutrition, by-products can be used to reduce the cost of feed, which is one of the factors affecting profitability. Following this purpose, the desired reduction in feed cost can be achieved by researching by-products and increasing the biological value of existing this (Yang et al., 2021). Beta vinasse, which is the fermentation source of molasses, can be considered a good source of nutrients and minerals (Parnaudeau et al., 2008). The chemical composition of β -vinasse added to the diet in this study was 35.39% crude protein and 31.74% betaine. Due to betaine in its β -vinasse structure, β -vinasse has gained importance as a valuable feed material in poultry.

Since poultry cannot synthesize methyl groups, methyl group donors such as glycine, a trimethyl derivative of glycine, should be added to poultry diets (Ratriyanto et al., 2017). Moreover betaine functions a major role in the metabolism of protein and energy (Metzler-Zebeli et al., 2009; Ratriyanto et al., 2009). Furthermore, it has the potential to replace some of the dietary methionine by providing a methyl group during the conversion of homocysteine to methionine (Metzler-Zebeli et al., 2009, Cetin et al., 2021). Consequently, it can increase the availability of methionine for protein synthesis, thus a better performance can be obtained (Rao et al., 2011). Betaine supplementation the diet changes the microbial population of the digestive system of poultry (Kettunen et al., 1999). Additionally, betaine supplementation may protect intestinal cells and intestinal microbes from osmotic variation along with the digestive system (Ratriyanto et al., 2009).

The study objective was to evaluate the effects of β -vinasse on performance parameters, carcass characteristics, meat quality, and ileo-caecal microflora in quails.

MATERIALS AND METHODS

Animals, rations and experimental

Totally 240 five-day-old Japanese (*Coturnix coturnix Japonica*) quails, including both males and females, were obtained from the Uludag University Animal Health and Production, Research and Application Centre For quail breeding (Bursa, Turkey). The care and use of animals were following laws and regulations of Turkey and approved by the Ethical Committee of Bursa Uludag University (Decision no: 22.09.2020, 2020-10/09).

The quails were randomly divided into one control group and two treatment groups containing 80 quails each. Each group was randomly distributed into five subgroups, comprised of 16 quails each. The quails in all of the groups were reared under the same growing conditions. All the quails were housed at 32 °C for the first three days and decreased by 1 °C every three days to 27 °C at the end of the second week. The quails were kept under constant artificial lighting for 23 h/d. All quails had *ad libitum* access to water and the form of mash feed along the experimental period. The study lasted for 42 days.

The basal rations were mixed under commercial conditions as one batch, divided into respective parts, and then added to β -vinasse using a horizontal mixer. The quails received a basal ration of 23 % crude protein; 3000 kcal/kg metabolizable energy (ME) that was formulated to meet the National Research Council (NRC, 1994) requirements for nutrients, including vitamins and minerals. The ingredients and chemical compositions of the basal rations are presented in Table 1. The chemical composition of β -vinasse is showed in Table 2. No antibiotics or growth promoters were supplemented to any of the experimental groups. The chemical compositions of basal rations were determined according to the Association of Official Analytical Chemists methods (AOAC, 2006). Group feeding was applied in all experimental groups. The quails were fed either a basal ration (control group) or the basal ration with β -vinasse supplementation at doses 15 (1.5%, group 1) or 30 (3.0%, group 2) g/kg feed. The ME level of basal ratios were estimated using the following equation from Carpenter and Clegg (Leeson and Summers, 2001): $ME, kcal/kg = 53 + 38 [(CP, \%) + (2.25 \times \text{ether extract, } \%) + (1.1 \times \text{starch, } \%) + (1.05 \times \text{sugar, } \%)]$.

Performance parameters

At the beginning of the study, the quails were weighed individually after that the chicks were weighed weekly to calculate the individual live weight (LW) and live weight gain (LWG). Mortality was recorded when it happened. Feed intake (FI) was recorded weekly and stated as g per quail per week. The feed conversion ratio (FCR) was calculated as g feed per g live weight gain. At the end of the study, the sex ratio was determined in each group.

Slaughter, carcass characteristics, and meat quality

A total of 75 quails, 5 males from each replicate

Table 1. Ingredients (%) and chemical composition of the quail rations

	Control	Group 1	Group 2
Ingredients			
Maize, grain	53.64	53.64	52.76
Soybean meal	28.62	27.12	25.62
Full fat soybean	10.30	10.30	11.08
Corn gluten	1.33	1.33	1.33
Vegetable oil	1.80	1.80	1.85
B-vinasse	-	1.5	3.0
Dicalcium phosphate	1.95	1.95	1.96
Limestone	0.90	0.90	0.90
Salt	0.25	0.25	0.25
Vit-min premix ¹	0.25	0.25	0.25
DL-Methionine	0.34	0.34	0.34
L-Threonine	0.10	0.10	0.10
L-Lysine	0.21	0.21	0.21
Sodium bicarbonate	0.10	0.10	0.10
Choline Chloride	0.11	0.11	0.12
Anticoccidial	0.10	0.10	0.10
Values analysed, %			
Metabolisable energy (kcal/kg)	3030,24	3028,26	3032,30
Crude protein	23,38	23,36	23,32
Ether extract	6.72	6.70	6.75
Ash	10.15	10.35	10.06
Saccharose	4.56	4.55	4.65
Starch	38.08	38.64	38.63
Dry matter	90.82	90.49	90.09
Calcium	1.03	1.12	1.11
Total Phosphorus	0.70	0.72	0.73

¹Per 2.0 kg premix contains; Vit A 12 500 000 IU, Vit D₃ 4 000 000 IU, Vit E 125 000 mg, Vit K₃ 3 000 mg, Vit B₁ 2 700 mg, Vit B₂ 7 000 mg, Vit B₆ 4 000 mg, Vit B₁₂ 20 mg, Vit C 66 000 mg, Niacine 60 000 mg, Calcium d-pantothenate 15 000 mg, Folic acid 1 500 mg, Biotin 150 mg, Mn 75 000 mg, Fe 15 000 mg, Zn 60 000 mg, Cu 10 000 mg, Co 200 mg, I 1 200 mg, Organic Se 150 mg, Se 150 mg, Crina Poultry Plus 300 000 mg, Fitase 1 000 000 FTU, Xylanase 270 000 U, β -Glucanase 80 000 U, Fungal-1.3-B-Glucanase 70 000 U

pen, were randomly selected and weighed from each pen for carcass characteristics and meat quality evaluation at the end of the study. Quails were randomly slaughtered, stunned by cervical dislocation, and decapitated 6 h after feed withdrawal by cutting between the occipital and atlas bones using scissors. Hot carcass weights were decided with the exclusion of edible internal organs, heads, and feet containing the neck and abdominal fat. Carcasses were kept in plastic bags and stored at 4 °C overnight to determined cold carcass weight and then carcass yields were calculated.

To determine meat pH and meat color 5 quails from each replicate were used. The pH values of the breast and thigh meat samples were determined by homogenizing with distilled water. The homogenates were filtered and the pH of each sample was mea-

sured with a previously calibrated portable pH meter (Metler-Toledo, Switzerland) at room temperature. Meat color was get from an average of 3 readings across the surface of the pectoralis minor and pectoralis major previously exposed to air ambient for 30 min. Immediately before data collection, a Minolta Chromameter CR-300 (Minolta Inc., Tokyo, Japan) was calibrated with a white, and black tile and then the lightness (L^*), redness (a^*), and yellowness (b^*) of breast meat was measured. The mean value from each breast meat was calculated.

Microbiological methods

Sixty ilio-cecal content samples were collected for microbial population analyses. The ileo-caecal region is the area where the ileum ends, obtained by the fixation of the cecum. After the gastrointestinal tract was

removed, the ileo-cecal area was aseptically removed by ligating it with fine twine, and the ileo-cecal area connected through the intestinal tract was transferred to the laboratory by cold chain without separating it. After opening the ilio-caecum for each quail, all contents were placed in sterile stomach bags, and massaged by hand from the outside.

For the isolation and enumeration, 1 g of caecal content homogenized with 9 ml of 0.1% peptone water in a Seward Stomacher 80 Lab System for 2 min. Serial dilutions were made in sterile peptone water and plated in duplicate onto relevant selective media. *Lactobacillus* were grown on de Man Rogosa and Sharpe (MRS, Oxoid CM361) agar and were enumerated after 3 d of incubation at 35°C under 5% CO₂. Total coliform was grown on Violet Red Bile (VRB, Bio life, 4021852) agar using the “pour” plate technique and plates with colonies were used for enumeration after 24–48 h of incubation at 37°C.

Statistical analysis

Statistical analysis was performed using the SPSS

software package for Windows (SPSS, 2016). Variance analysis was used to determine the significance of the differences between the statistical calculations of the groups and the mean values of the group, Duncan test was used as a post-hoc test and the level of significance was used in all of the tests $P < 0.05$. Also, in this study, the sex ratio in quails was evaluated by the chi-square test and the level of significance used in all of the tests was $P < 0.05$. Results are expressed as mean \pm standard error of the mean.

RESULTS

The current study was carried out to determine the effects of different levels of β -vinsasse on the performance, carcass parameters, meat color, and ilio-caecal microflora (*Lactobacillus* spp., Coliform bacteria) of quails. The nutrient composition of β -vinsasse is presented in Table 2. The performance values obtained from the groups in this study, which was conducted to define the effects of β -vinsasse added to quail rations at different levels on performance parameters are given in Table 3. In this study, there were no differences in the sex ratio, FI and FCR. As shown in

Table 2. Nutrient composition of β -vinsasse

Nutrients	β -Vinsasse (%100 DM)	β - Vinsasse (% 63 DM)
Crude protein (%)	35,39	22,30
Crude ash (%)	17,46	11,00
Metabolisable energy (MJ/kg)	6,20	3,91
Crude cellulose (%)	1,26	0,80
Lysine (%)	0,21	0,137
Meth & Cys (%)	0,050	0,032
Methionine (%)	0,050	0,032
Threonine (%)	0,26	0,169
Valine (%)	0,32	0,206
İsoleucine (%)	0,21	0,136
Arginine (%)	0,09	0,061
Tryptophan (%)	0,06	0,039
Calcium (%)	0,04	0,028
Total phosphorus (%)	0,08	0,054
Sodium(%)	2,23	1,41
Potassium (%)	2,87	2,05
Betaine (%)	31,74	20,00
D.Lysine (%)	0,13	0,086
D.Meth & cys (%)	0,31	0,20
D.Methionine (%)	0,33	0,21
D.Threonine (%)	0,16	0,105
D.Valine (%)	0,20	0,130
D.İsoleucine (%)	0,13	0,084
D.Arginine(%)	0,06	0,039
D.Tryptophan (%)	0,03	0,024

Table 3. Effects of β -vinasse supplementation on live weight, live weight gain, feed intake and feed conversion ratio in quails

	Control	Group 1	Group 2	P value
Sex ratio (M/F)	38/42	37/43	36/44	1.00
Live weight (g) (n=80)				
5 d	10.96±0.17	11.30±0.20	11.56±0.18	0.084
21 d	106.17±1.24 ^b	108.67±1.12 ^b	112.42±1.06 ^a	0.001
42 d	190.86±3.19	192.80±2.66	195.14±3.23	0.612
Live weight gain (g) (n=80)				
5 to 21 d	95.18±1.27 ^b	97.36±1.10 ^b	100.86±1.07 ^a	0.002
22 to 42 d	84.63±3.46	84.13±2.92	82.71±3.35	0.911
5 to 42 d	179.86±3.21	181.49±2.65	183.58±3.24	0.691
Feed intake (g) (n=5)				
5 to 21 d	262.75±7.43	269.12±6.14	263.44±4.31	0.726
22 to 42 d	534.83±20.77	541.65±17.85	508.56±9.59	0.368
5 to 42 d	797.59±22.60	820.77±21.45	772.00±11.17	0.376
Feed conversion ratio (g/g) (n=5)				
5 to 21 d	2.76±0.10	2.77±0.04	2.61±0.04	0.275
22 to 42 d	6.98±0.77	6.43±0.16	6.15±0.15	0.469
5 to 42 d	4.45±0.19	4.47±0.09	4.21±0.06	0.325

^{a,b} Mean values with different superscripts in the same row differ significantly ($p < 0.05$)

M: Male F: Female

Values are expressed means ±standart deviation Group 1: 15 g/kg β -vinasse, Group 2: 30 g/kg β -vinasse

Table 4. Effects of β -vinasse supplementation on carcass characteristics and meat quality in quails (n=25)

	Control	Group 1	Group 2	P value
Carcass Characteristics				
Final live weight (g)	169.29±3.01	174.14±3.07	173.21±3.40	0.520
Hot carcass weight (g)	133.01±2.56	136.66±2.42	134.50±2.87	0.617
Cold carcass weight (g)	129.06±2.50	132.46±2.40	129.13±2.87	0.576
Hot carcass yield (%)	78.63±0.88	78.56±0.76	77.63±0.58	0.582
Cold carcass yield (%)	76.30±0.87	76.13±0.74	74.50±0.59	0.177
Meat Quality				
pH thigh	6.44±0.03	6.52±0.04	6.46±0.03	0.193
pH breast	5.98±0.02	6.01±0.03	6.02±0.02	0.486
L	36.64±0.48 ^b	38.61±0.84 ^b	47.18±1.41 ^a	0.000
A	10.41±0.33 ^a	10.40±0.34 ^a	7.92±0.46 ^b	0.000
B	1.24±0.22 ^b	2.53±0.46 ^b	6.22±0.60 ^a	0.000

^{a,b} Mean values with different superscripts in the same row differ significantly ($p < 0.05$)

Values are expressed means ±standart deviation

Group 1: 15 g/kg β -vinasse, Group 2: 30 g/kg β -vinasse

Table 3, significant differences ($P < 0.05$) LW (21 d) and LWG (5 to 21 d) were observed. Especially, β -vinasse at 30 g/kg caused a significant increase in LW (21 d) and LWG (5 to 21 d). The dietary treatment of quails with different levels of β -vinasse did not affect hot carcass weight, cold carcass weights, hot and cold carcass yields, and breast and thigh pH (Table 4). The lightness (L^*) and yellowness (b^*) of breast values were significantly ($P < 0.05$) increased in group 2. The effects of dietary supplementation with different levels of β -vinasse on ilio-caecal bacteriological flora are

provided in Table 5. The result of the study showed that the different levels of β -vinasse significantly increased ($P < 0.05$) *Lactobacillus* spp. ilio-caecal area. There are no statistical differences in coliform bacteria among the different groups.

DISCUSSION

The nutrient compositions of β -vinasse are presented in Table 2. As shown in Table 2, the β -vinasse used in this research contained 63% dry matter, 22.3% crude protein, 11% crude ash, 20% betaine, and

Table 5. Effects of β -vinasse supplementation on ilio-caecal microflora in quails (n=20)

	Control	Group 1	Group 2	P value
Coliform (Log cfu/g)	2.46±0.36	3.25±0.30	2.95±0.37	0.279
Lactobacillus (Log cfu/g)	2.98±0.16 ^b	3.69±0.13 ^a	3.50±0.15 ^a	0.004

^{a,b} Mean values with different superscripts in the same row differ significantly ($p < 0.05$)

Values are expressed means \pm standard deviation

Group 1: 15 g/kg β -vinasse, Group 2: 30 g/kg β -vinasse

2.05% potassium. In another study, ProMass (modified vinasse product) contains 35.47% crude protein, 5.94% crude ash, 10% betaine, and 1.66% potassium (Yalcin et al., 2010). These levels may vary according to processing conditions and extraction methods.

In the study, it was determined that the addition of 30 g/kg β -vinasse to the diet caused a significant increase in live weight and live weight gain, especially in the initial period. These results are in agreement with those obtained by (Attia et al., 2005; Nofal et al., 2015) who found that the addition of betaine to poultry diet improved significantly body weight. This improvement in performance may be due to the effect of betaine on nutrient digestibility. Also, the total betaine need cannot be met by endogenous metabolism, thus, dietary betaine supplementation may be beneficial to maintain or to improve chicken, health, and productive performance (Eklund et al., 2005). In their study, Cengiz et al. (2019) found that the addition of 30 g/kg bromass (modified vinasse) to the broiler ration caused a significant increase in body weight and body weight gain in broilers at the beginning and development period, while it caused a significant decrease in feed consumption. Lewicki (2001) and Stemme et al. (2005), in their studies, reported that the addition of β -vinasse to the diet caused a decrease in feed cost and a higher level of performance. Studies have also shown that this effect is more significant under stress conditions. Attia et al. (2009) stated that the effect of severe heat stress could partially be overcome by adding 1 kg betaine/ton to chick diets which improved weight gain and feed conversion compared to negative treatment. In contrast to our results, Uzunoglu and Yalcin (2019) showed that betaine dietary supplementation did not affect the growth performance indices of broilers, which could be due to different experimental conditions and the supplemented dose of betaine between the two studies. Different experimental results suggested the need for further experiments to escape the gap in contradictory results due to different levels of betaine used, animal species and environmental conditions, and composition of the ex-

perimental diets. Betaine supplement may stimulate protection of intestinal epithelium against osmotic disturbance; improve digestion, absorption, and nutrient utilization in broiler chickens (Mahmoudnia and Madani, 2012).

Our results showed that β -vinasse supplementation did not affect quail carcass characteristics, but caused significant changes in the values of parameters effective in meat quality, such as meat L^* , a^* , and b^* values. Likewise, Al-Sagan et al. (2021), in their study, determined that the addition of betaine at different levels to broiler diets did not cause a significant effect on carcass parameters and meat quality parameters (L^* , a^* value, and b^* value). In contrast, El-Shinnawy (2015) revealed that broilers that consumed betaine-supplemented diets had increased carcass and carcass part yield percentages. Attia et al. (2009), who indicated that heat stress and dietary betaine supplementation did not affect meat pH and color. Nutautaitė et al. (2020) showed that betaine dietary supplementation did not influence breast-muscle colors L^* , b^* , and the water holding capacity (WHC) in broilers meat.

Although there are many studies on the performance of meat and egg-oriented poultry on betaine and its derivatives, studies on the gastrointestinal microbiota population are limited. Betaine also acts as an organic osmolyte by stabilizing intestinal epithelial cells and supporting intestinal microbial growth (Metzler-Zebeli et al., 2009; Weiss et al., 2013). In the digestive tract, there are trillions of microorganisms that cooperate with the host organism with many mechanisms such as nutrient absorption, energy balance, and immunity (Kostadinović and Lević, 2018). Recent studies have focused on this issue, as a healthy and stable microbial community (microbiota) is crucial to animal health and performance. In this study, it was determined that the β -vinasse supplement added to the quail diet caused a significant increase in the beneficial microorganism population. In particular, the *Lactobacillus* species found in the gastrointestinal tract have received tremendous attention due to their

health-promoting properties. Lactic acid bacteria, particularly *Lactobacillus* strains, are frequently used as probiotics which are defined by the FAO/WHO as live microorganisms that when administered in adequate amounts confer a health benefit on the host (Walter, 2008). *Lactobacillus* strains have a high ability to attach to the intestinal epithelium and can establish in the chicken intestine within a day after hatching, so they are considered to be normal bacterial flora of the gastrointestinal tract of chickens (Shokryazdan et al., 2017; Shah et al., 2019). Ahmed et al., (2018) determined that using betaine alone did not have significant effects total count of *E.coli* and *Lactobacillus* bacteria. Although there are data on the small intestine histomorphology of betaine supplementation in the studies (Alahgholi et al., 2014; Klasing et al. 2002) conducted, data on the small intestine microbiota have not been found. In this sense, the data of this study will help reveal the effects on the small intestine microorganism population change.

CONCLUSIONS

As a result, it was concluded that β -vinsasse (by-product obtained from molasses) can be used in quail diets as an alternative feed source that will meet the nutritional needs of the animal and have positive effects on the digestive system, especially on the intestinal health. It has been demonstrated that the addition of β -vinsasse at 15 and 30 g/kg levels to quail diets has a positive effect on *Lactobacillus* type microorganisms that have probiotic effects in terms of intestinal health.

ACKNOWLEDGEMENTS

The authors thank Integro for supplying the feed additive supplement.

CONFLICT OF INTEREST STATEMENT

The authors declare no conflict of interest.

REFERENCES

- Ahmed MMN, Ismail ZSH, Abdel-Wareth AAA (2018) Effect of dietary supplementation of prebiotic, betaine and their combination on growth performance, nutrient digestibility, carcass criteria and cecum microbial population of ducks under hot environmental conditions. *Egypt Poult Sci* 38: 289-304.
- Alahgholi M, Tabeidian SA, Toghyani M, Fosoul SSAS (2014) Effect of betaine as an osmolyte on broiler chickens exposed to different levels of water salinity. *Archiv Tierzucht* 4: 1-12.
- Al-Sagan AA, Al-Yemni AH, Abudabos AM, Al-Abdullatif AA, Hussein EO (2021) Effect of different dietary betaine fortifications on performance, carcass traits, meat quality, blood biochemistry and hematology of broilers exposed to various temperature patterns. *Animals* 11:1555.
- AOAC (2000) Official Methods for Analysis. 18th ed. Arlington USA: Association of Official Analytical Chemists.
- Attia YA, Hassan RA, Qota EM (2009) Recovery from adverse effects of heat stress on slow growing chicks in the tropics I: Effect of ascorbic acid and different levels of betaine. *Trop Anim Health Prod* 41: 807-818.
- Attia YA, Hassan RA, Shehatta MH, Abd El-Hady SB (2005) Growth, carcass quality and serum constituents of slow growing chicks as affected by betaine addition to diets containing different levels of methionine. *Int J Poult Sci* 4: 856-865.
- Cengiz SS, Yesilbag D, Cetin I, Gunes NA, Eren M, Topaloglu G (2019) Use of bromass in broiler rations as a different protein source. *Revue Med Vet* 170: 95-103.
- Cetin I, Yesilbag D, Cengiz SS (2021) Effects of Beta Vinsasse Using on Performance, Egg Production and Egg Quality in Layer Quail Diets. *J Anatolian Env Anim Sciences* 6(1): 106-111.
- Eklund M, Bauer E, Wamatu J, Mosenthin R (2005) Potential nutritional and physiological functions of betaine in livestock. *Nutr Res Rev* 18: 31-48.
- El-Shinnawy AM (2015) Effect of betaine supplementation to methionine adequate diet on growth performance, carcass characteristics, some blood parameters and economic efficiency of broilers. *J Anim Poult Prod* 6: 27-41.
- Kettunen H, Peuranen S, Jatila H, Nurminen P, Saarinen M, Apajalahti J (1999) Effect of betaine on the microbiology of the chicken gastrointestinal tract. *Proceedings of the 12th European Symposium on Poultry Nutrition*; 1999 Aug 15-19; Veldhoven (NL): World's Poultry Science Association p 186.
- Klasing KC, Adler KL, Remus JC, Calvert CC (2002) Dietary betaine increases intraepithelial lymphocytes in the duodenum of coccidia-infected chicks and increases functional properties of phagocytes. *J Nutr* 132: 2274-2282.
- Kostadinović L, Lević J (2018). Effects of phytoadditives in poultry and pigs diseases. *J Agron Technol Eng Manag* 1: 1-7
- Leeson S, Summers JD (2001) *Nutrition of the Chicken*. 4th ed. Guelph, ON: University Books.
- Lewicki W (2001) An introduction to vinsasse from beet and cane molasses fermentation. *Int Sugar J* 103:126-128.
- Mahmoudnia N, Madani Y (2012) Effect of betaine on performance and carcass composition of broiler chicken in warm weather *Int J Agr Sci* 2: 675-683.
- Metzler-Zebeli BU, Eklund M, Mosenthin R (2009) Impact of osmoregulatory and methyl donor functions of betaine on intestinal health and performance in poultry. *Worlds Poult Sci J* 65: 419-441.
- Nofal ME, Magda A, Galal SMM, Mousa M, Doaa MM, Yassein M, Bealsh AMA (2015) Effect of dietary betaine supplementation on productive, physiological and immunological performance and carcass characteristic of growing developed chicks under the condition of heat stress. *Egypt Poult Sci* 35:237-259.
- NRC (1994) *Nutrient Requirements of Poultry*. 9th ed. Washington, DC: National Academy Press.
- Nutautaitė M, Alijošius S, Bliznikas, S, Šašytė V, Vilienė V, Pockevičius A, Racevičiūtė-Stupelienė A (2020) Effect of betaine, a methyl group donor, on broiler chicken growth performance, breast muscle quality characteristics, oxidative status and amino acid content. *Ital J Anim Sci* 19: 621-629.
- Parmaudeau V, Condom N, Oliver R, Cazevieville P, Recous (2008) Vinsasse organic matter quality and mineralization potential as influenced by raw material, fermentation and concentration processes. *Bioresour Technol* 99: 1553-1562.
- Rao SVR, Raju MWLN, Panda AK, Sahaira P, Sunder GS (2011) Effect of supplementing betaine on performance, carcass traits and immune re-

- sponses in broiler chicken fed diets containing different concentration of methionine. *Asian-Australas J Anim Sci* 24: 662-669.
- Ratriyanto A, Indreswari R, Nuhriawangsa AMP (2017) Effects of dietary protein level and betaine supplementation on nutrient digestibility and performance of Japanese Quails. *Braz JPoult Sci* 19: 445-454.
- Ratriyanto A, Mosenthin R, Bauer E, Eklund M (2009) Metabolic, osmoregulatory and nutritional functions of betaine in monogastric animals. *Asian-Australas J Anim Sci* 22: 1461-1476.
- Shah M, Zaneb H, Masood S, Ullah Khan R, Ashraf S, Sikandar A, Rehman HFU, Rehman HU (2019) Effect of Dietary Supplementation of Zinc and Multi-Microbe Probiotic on Growth Traits and Alteration of Intestinal Architecture in Broiler. *Probiotics and Antimicrob Proteins* (2019) 11:931-937.
- Shokryazdan P, Faseleh Jahromi M, Liang JB, Ramasamy K, Siew CC, Ho YW (2017) Effects of a *Lactobacillus salivarius* mixture on performance, intestinal health and serum lipids of broiler chickens. *PLoS ONE* 12(5): e0175959.
- SPSS 2016. SPSS Statistics for Windows, Version 24. Armonk, NY. IBM.
- Stemme K, Gerdes B, Harms A, Kamphues J (2005) Beet-vinasse (condensed molasses solubles) as an ingredient in diets for cattle and pigs- Nutritive value and limitations. *Anim Physiol Anim Nutr* 89:179-183.
- Uzunoglu K, Yalcin S (2019) Effects of dietary supplementation of betaine and sepiolite on performance and intestinal health in broilers. *Ank Üniv Vet Fak Derg* 66: 221-229.
- Walter J (2008) Ecological role of lactobacilli in the gastrointestinal tract: implications for fundamental and biomedical research. *Appl Environ Microbiol* 74:4985-4996.
- Weiss E, Eklund M, Semaskaite A, Urbaityte R, Metzler-Zebeli B, Sauer N, Ratriyanto A, Gruzauskas R, Mosenthin R (2013) Combinations of feed additives affect ileal fibre digestibility and bacterial numbers in ileal digesta of piglets. *Czech J Anim Sci* 58: 351-359.
- Yalcin SK, Eltan O, Karlı MA (2010) The nutritive value of modified dried vinasse (ProMass) and its effects on growth performance, carcass characteristics and some blood biochemical parameters in steer. *Rev Med Vet* 161: 245-252.
- Yang K, Qing Y, Yu Q, Tang X, Chen G, Fang R, Liu H (2021) By-Product Feeds: Current understanding and future perspectives. *Agriculture* 11: 207.