

# Journal of the Hellenic Veterinary Medical Society

Vol 74, No 2 (2023)



## Kaempferol and Isorhamnetin alleviate Lipopolysaccharide-Induced Anxiety and Depression-Like Behavioral in Balb/C Mice

M Ekici, H Güngör, DG Mert

doi: [10.12681/jhvms.30232](https://doi.org/10.12681/jhvms.30232)

Copyright © 2023, M Ekici, H Güngör, DG Mert



This work is licensed under a [Creative Commons Attribution-NonCommercial 4.0](https://creativecommons.org/licenses/by-nc/4.0/).

### To cite this article:

Ekici, M., Güngör, H., & Mert, D. (2023). Kaempferol and Isorhamnetin alleviate Lipopolysaccharide-Induced Anxiety and Depression-Like Behavioral in Balb/C Mice. *Journal of the Hellenic Veterinary Medical Society*, 74(2), 5749-5760.  
<https://doi.org/10.12681/jhvms.30232> (Original work published July 5, 2023)

## Kaempferol and Isorhamnetin Alleviate Lipopolysaccharide-Induced Anxiety and Depression-Like Behavioral in Balb/C Mice

M. Ekici<sup>1\*</sup>, H. Gungor<sup>2</sup>, D.G. Mert<sup>3</sup>

<sup>1</sup> Sivas Cumhuriyet University, Faculty of Veterinary Medicine, Department of Veterinary Physiology, Sivas

<sup>2</sup> Sivas Cumhuriyet University, Faculty of Veterinary Medicine, Department of Veterinary Pharmacology and Toxicology, Sivas

<sup>3</sup> Ankara City Hospital, Department of Psychiatry, Ankara

**ABSTRACT:** The etiology of anxiety and depression is linked to inflammation and oxidative stress. Isorhamnetin and Kaempferol are strong antioxidants with anti-inflammatory neuroprotective properties. This study, it was aimed to investigate the effect of Kaempferol and Isorhamnetin in Lipopolysaccharide (LPS)-induced anxiety and depression model in mice. Thirty Balb/C mice were divided into six groups of five mice each weighing 25-35 g. Kaempferol (50 mg/kg and 100 mg/kg) and Isorhamnetin (30 mg/kg and 60 mg/kg) were given orally to the treatment group, and the vehicle was given to the control and LPS groups for fourteen days, followed by intraperitoneal (0.83 mg/kg) LPS injection (control group excluding) on the fifteenth day. At 3 hours after the administration of LPS, each group was applied with the Elevated Plus-Maze (EPM) test, the Light/Dark test, and the Open Field test (OFT). At 24 hours after LPS administration, the Forced Swimming Test (FST) was applied and at 28 hours, the Tail Suspension Test (TST). After the behavioral test, the prefrontal cortex and hippocampus tissues of rats were harvested by cervical dislocation under high-dose anesthesia. From these tissues, malondialdehyde (MDA) levels, total oxidant status (TOS) and total antioxidant status (TAS) levels, tumor necrosis factor alpha (TNF- $\alpha$ ), interleukin-1beta (IL-1 $\beta$ ), and interleukin-6 (IL-6) levels, and brain-derived neurotrophic factor (BDNF) levels were measured. Kaempferol and Isorhamnetin ameliorated LPS-induced anxiety evaluated by OFT, Light/Dark test, and EPM, and LPS-induced depression evaluated by FST and TST. Kaempferol and Isorhamnetin alleviated LPS-induced increased oxidative stress in the prefrontal cortex and hippocampus by decreasing MDA and TOS levels and increasing TAS levels. In addition, it was observed that Kaempferol and Isorhamnetin regulated the increase in prefrontal and hippocampal inflammation caused by LPS by decreasing TNF- $\alpha$ , IL-1 $\beta$ , and IL-6 levels. Most importantly, LPS reduced prefrontal and hippocampal BDNF levels, but the treatment groups reversed it. These results show the possible therapeutic potential of Kaempferol and Isorhamnetin, along with the importance of oxidative stress, inflammation, and BDNF in the etiopathogenesis of anxiety and depression.

**Keywords:** Anxiety, Depression, Oxidative Stress, Mice, Kaempferol

*Corresponding Author:*

M. Ekici Sivas Cumhuriyet University, Faculty of Veterinary Medicine, Department of Veterinary Physiology, Sivas, 58140  
E-mail address: mehmetekici@cumhuriyet.edu.tr

Date of initial submission: 20-04-2022  
Date of acceptance: 18-11-2022

## INTRODUCTION

Anxiety and depression are commonly seen as psychiatric disorders throughout the world (Steimer, 2002). These two psychiatric disorders are related to biochemical, cognitive, behavioral, and psychological changes which are defined as negative emotional experiences in individuals (Steimer, 2002). Both depression and anxiety have negative effects on the life of the individual and cause a significant functional loss, and depression and anxiety disorders are frequently observed together (Brenes, 2007; Hussein et al., 2018). The etiopathogenesis of both disorders has not yet been clarified, and antidepressant drugs from the selective serotonin reuptake inhibitors (SSRI) group are the first choice in treatment. Drug therapy has been reported to be effective in approximately one-third of patients, including the prevention of recurrence (Xu et al., 2012; Zhao et al., 2016; Jamali-Raeufy et al., 2019).

The efficacy of flavonoids, which affect cytokine, neurotrophin, and oxidative system, has been investigated in the treatment of depression and anxiety (Mesfin et al., 2014; Saki et al., 2014; Sulakhiya et al., 2016). Cytokines play a role in the modulation of various neurological functions (Mormède et al., 2004; O'Connor et al., 2009). The brain can rapidly create new synapses in the functions necessary for learning and adaptation or eliminate synapses. BDNF is one of the molecular factors required for healthy neuroplasticity. BDNF is the neurotrophin enabling growth and differentiation of neurons, synapse formation, and the survival of existing neurons (Cheng et al., 2018). Free radicals are highly reactive chemical species that emerge during normal metabolic processes (Chi et al., 2016). Compared to other organs, the central nervous system is more sensitive to the toxic effects of reactive oxygen species.

Flavonoids have a diphenyl propane structure and are secondary metabolites of a plant group (Ko et al., 2020). Kaempferol (3,5,7-trihydroxy-2-(4-hydroxy-phenyl)-4H-1-benzopyran-4-one) is a natural flavonoid widely found in various fruits and vegetables (tea, apples, strawberries, green beans, broccoli, spinach). Isorhamnetin (30-O-methyl-quercetin) is a flavonoid found in various plant-based food products such as apples and grapes, and herbal drugs such as sea buckthorn and *Ginkgo biloba* (Rammal et al., 2008; Zhang et al., 2012; Jangra et al., 2014). In vivo studies have shown that both of these flavonoids have several antioxidants, anti-inflammatory, and neuroprotective biological ac-

tivities (Brenes, 2007; Xu et al., 2012; Zhao et al., 2016; Hussein et al., 2018; Jamali-Raeufy et al., 2019).

There are various therapeutic regimes for anxiety and depression. However, the treatment process is negatively affected by the low efficacy of the drugs, delayed therapeutic effects, various side effects, and poor drug compliance (Mesfin et al., 2014; Sulakhiya et al., 2016). Therefore, it is important that the etiopathogenesis of these disorders is investigated and that more effective new compounds with a low side-effect profile are developed (Saki et al., 2014). The aim of this study was to investigate the protective effect against depression and anxiety of Kaempferol and Isorhamnetin, which are flavonoids with neuroprotective, anti-inflammatory, and antioxidant properties, in a lipopolysaccharide-induced model in mice through animal behavior tests and biochemical analyses.

## MATERIALS AND METHODS

### Chemical reactants and drugs

Kaempferol and Isorhamnetin were purchased from Aktin Chemicals, Inc. (Chengdu, China) and LPS from Sigma Chemical Company (St. Louis, MO, USA). Trichloroacetic acid (TCA), thiobarbituric acid (TBA) and Hydrochloric acid (HCl) were obtained from Merck Chemicals (Darmstadt, Germany). These chemicals were used instead of a kit for MDA analysis. The commercial ELISA kits used in the measurements of TOS, TAS, TNF- $\alpha$ , IL-1 $\beta$ , and IL-6 levels, and BDNF, were purchased from Shanghai Sunred Biological Technology Co., Ltd. (Shanghai, China). All the chemical reactants were of analytical purity.

### Animals and study groups

This study was conducted in the Experimental Animals Laboratory of Sivas Cumhuriyet University Medical Faculty in compliance with the guidelines for the Care and Use of Laboratory Animals. The study protocol was approved by the Animal Ethics Committee of Sivas Cumhuriyet University (decision no: 65202830-050.04.04-172). The study sample comprised thirty male mice (Balb/C), each weighing 25-35 g. The mice were kept under standard animal laboratory conditions of a 12-hour light/dark cycle, at  $24 \pm 2^\circ\text{C}$ , 35-60% humidity, and food and water *ad libitum*. Following an adaptation period of one week, the mice were randomly divided into 6 groups ( $n=5/\text{group}$ ): Control, LPS, LPS+Kaempferol50, LPS+Kaempferol100, LPS+Isorhamnetin30, and LPS+Isorhamnetin60 groups.

The control group received 10% Dimethyl sulfoxide (DMSO) by oral gavage for 14 days and followed by intraperitoneal (i.p.) saline on day fifteen. The LPS group received 10% DMSO by oral gavage (dose volume 5 ml/kg bw) for 14 days, then a single i.p. injection (dose volume 10  $\mu$ l/g bw) of 0.83 mg/kg LPS on the 15th day (Mormède et al., 2004; O'Connor et al., 2009). Kaempferol was administered to the LPS+Kaempferol50 and LPS+Kaempferol100 groups for 14 days by oral gavage (dose volume 5 ml/kg bw) at doses of 50 mg/kg and 100 mg/kg, respectively, followed by a single i.p. injection of LPS at a dose of 0.83 mg/kg on the 15th day (Cheng et al., 2018). The LPS+Isorhamnetin30 and LPS+Isorhamnetin60 groups received Isorhamnetin by oral gavage (dose volume 5 ml/kg bw) at a dose of 30 mg/kg and 60 mg/kg, respectively, for 14 days, then a single i.p. injection of LPS at a dose of 0.83 mg/kg on the 15th day (Chi et al., 2016) (Figure 1).

The LPS was dissolved in saline. The animals were taken to the laboratory room where the behavioral tests were to be performed 30 mins before the behavioral tests for adaptation to the environment. The laboratory room was illuminated with dim red lights and the behavioral tests were conducted in this environment.

### Behavioral Analysis

At 3 hours after the administration of LPS, each group was applied with the Elevated Plus-Maze (EPM) test, the Light/Dark test, and the Open Field test (OFT). At 24 hours after LPS administration, the Forced Swimming Test (FST) was applied and at 28 hours, the Tail Suspension Test (TST). All the behav-

ioral tests were conducted using behavioral platforms made specifically for each test and were recorded in a computer video recording tracking system and analyzed in Ethovision XT<sup>©</sup> software (Noldus Information Technologies, Wageningen, Holland). Moreover, to avoid bias, all behavioral tests were performed by experienced researchers unaware of the groups, using a blind technique. To minimize the potential outcomes of the behavioral evaluations on the biochemical parameters, the neurochemical and behavioral evaluations were made in distinct groups. The behavioral test and experiment design were adapted according to the previously described (Sulakhiya et al., 2016).

### Open field test

This test is used to evaluate anxiety-like behavior and locomotion. The open field apparatus was made of a grey polyvinyl chloride plastic panel 40 cm high by 40 cm wide and 40 cm deep. Peripheral and central areas were formed. The middle area was defined as a 10 x 10 cm square. At the start of the test, each mouse was placed in the middle area and a video recording of 5 minutes was made. The time spent in the middle area was evaluated on the video recordings (Parrella et al., 2022).

### Light/Dark test

This test is used to evaluate anxiety-like behavior. Mice have an innate tendency to avoid light areas and unfamiliar surroundings. An apparatus was created for this purpose consisting of two sections of light and darkness. The time that the mice spent in the light and dark sections were measured throughout the test period of 5 minutes. Each mouse was placed in a box measuring 40 x 40 x 40 cm in a dark box measuring 40 x 40 x 20 cm, with a 6 x 6 cm door connecting the light

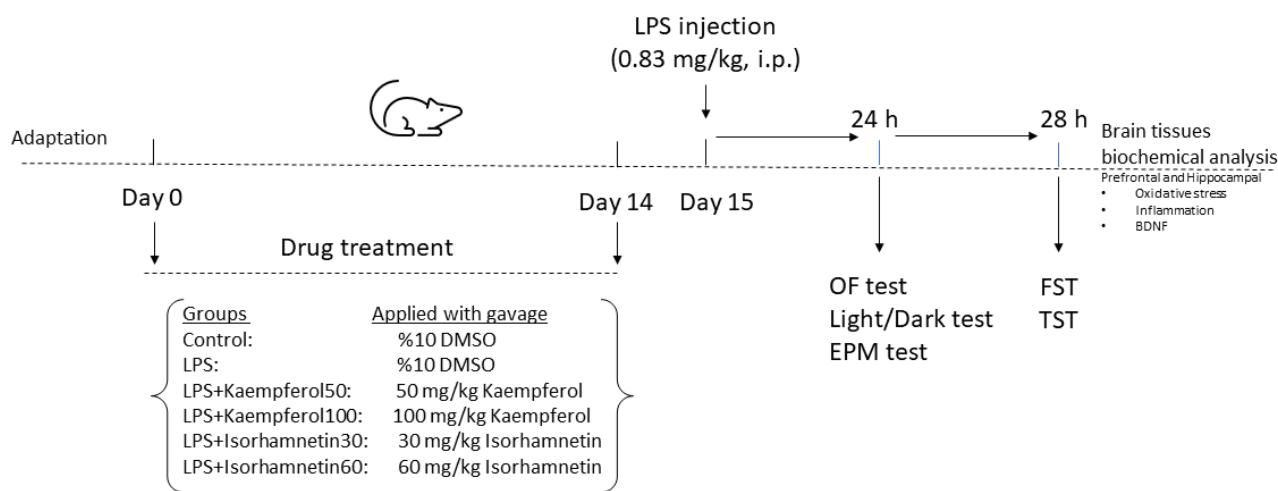


Figure 1: Experimental design

and dark sections. The times spent in the light and dark areas were recorded for evaluation (Peng et al., 2019).

#### *Elevated Plus-Maze test*

This test is used to evaluate anxiety-like behavior. There are a total of four arms, two open opposite each other and two closed on the opposite sides. These arms are separated from each other by a central region in the middle. In this test apparatus, the open and closed arms are 35 cm long and 5 cm wide, the middle area is 5 x 5cm and it is elevated from the ground at 50 cm. It is an elevated test apparatus. The elevation and open arms are perceived by the mice as unfamiliar areas. Each mouse was placed in the middle area and the time spent in the open and closed arms was evaluated throughout 5 minutes (Sulakhiya et al., 2016; Peng et al., 2019).

#### *Forced Swimming test*

This test is used to evaluate depression-like behavior. Water at a temperature of 25°C was placed at a depth of 20 cm in a glass cylinder 25 cm in depth and 10 cm in diameter. Each mouse was placed in the water and left to swim for 6 minutes. The period of complete lack of movement was measured in the last four minutes of the six minutes (Sulakhiya et al., 2016).

#### *Tail Suspension test*

This is a mouse behavioral test used in the screening of potential antidepressant drugs and evaluation of other manipulations expected to be affected by behavior related to depression. Each mouse is suspended by the tail which is taped so that it cannot escape or grasp nearby surfaces. Throughout the test period of 6 mins, the time spent mobile and immobile is measured (Sulakhiya et al., 2016).

#### **Analysis of Prefrontal and Hippocampal Oxidative Stress, Inflammation, and BDNF levels**

After completion of the behavioral tests, the animals were anaesthetized with xylazine (10mg/kg, i.p., Xylazine Bio 2%, Bioveta, Czech Republic) and ketamine (60 mg/kg, i.p., Alfamine, Egevet, Turkey), blood samples were taken then euthanasia was applied with the cervical dislocation method. The prefrontal and hippocampal regions were isolated from the brain. An amount of tissue was weighed, and then placed in phosphate-buffered saline (pH:7.4) buffer solution, to which three 2mm metal beads were added. The tissues were then double cycle homogenized in a bead beater homogenizer [HT 24 bead beating

homogenizer (OPS Diagnostics)]. A portion of the homogenates was separated for MDA analysis. The other portion was centrifuged for 10 mins at +4°C at 4000 rpm. The supernatants collected were stored at -80°C until analysis. The levels of TOS, TAS, BDNF, and proinflammatory cytokines (TNF- $\alpha$ , IL-1 $\beta$  and IL-6) were measured with commercial ELISA kits (Sunred Biological Technology, Shanghai, China). Briefly, serum samples, biotinylated antibodies, and Streptavidin-HRP were added sequentially to the plate coated with canine-specific antibodies and allowed to incubate for one hour. Then, the washing process was performed, the substrate solutions were added, and the reaction was started, respectively. The reaction was stopped by adding a stop solution and reading in an ELISA plate reader (Thermo Scientific™ Multiskan™ GO Microplate Spectrophotometer, USA) at 450 nm within 10 minutes. Analyses were performed in duplicate from all samples. The lipid peroxidation level (MDA) from the homogenate was determined spectrophotometrically (Ohkawa et al., 1979). The total protein amount from the supernatant was measured with an autoanalyzer.

#### **Statistical analysis**

Data obtained in the study were analyzed statistically using SPSS 23.0 software. Conformity of the behavioral test data to normal distribution was assessed with the Kolmogorov-Smirnov test. In all the groups, the data for each behavioral test and all the biochemical data were determined to show normal distribution. Statistically significant differences between groups were evaluated with One-way Variance analysis (ANOVA). The post-hoc Tukey HSD test was used in the comparisons of sub-groups in respect of the total time in the central area in the OFT, the time spent in the light and dark areas in the Light-Dark Test, the total time spent in the open and closed arms of the EPM test, the total periods of immobility in the FST and TST, and all the biochemical analysis data. Results were expressed as mean  $\pm$  standard deviation (SD) values. A value of  $p < 0.05$  was accepted as statistically significant.

## **RESULTS**

#### **Behavioral Results**

##### *Effect of Kaempferol and Isorhamnetin on OFT*

In the OFT, the time spent in the middle area was statistically significantly shorter in the LPS group than in the control group ( $p < 0.05$ ). The time spent in

the middle area was statistically significantly longer in the LPS+Kaempferol50 and LPS+Isorhamnetin30 groups than in the LPS group ( $p<0.05$ ) (Table 1).

*Effect of Kaempferol and Isorhamnetin on Light-Dark Test*

In the Light-Dark Test, the time spent in the light area was statistically significantly shorter in the LPS group than in the control group ( $p<0.05$ ). The time spent in the light area was statistically significantly longer in the LPS+Kaempferol50, LPS+Kaempferol100, and LPS+Isorhamnetin30 groups than in the LPS group ( $p<0.05$ ) (Table 2). The time spent in the dark area was statistically significantly longer in the LPS group than in the control group ( $p<0.05$ ). The time spent in the dark area was statistically significantly shorter in the LPS+Kaempferol50, LPS+Kaempferol100, and LPS+Isorhamnetin30 groups than in the LPS group ( $p<0.05$ ) (Table 2).

*Effect of Kaempferol and Isorhamnetin on EPM Test*

In the EPM test, the time spent in the open arms was statistically significantly shorter in the LPS group than in the control group ( $p<0.05$ ). The time spent in the open arms was statistically significantly longer in

the treatment groups than in the LPS group ( $p<0.05$ ) (Table 3). The time spent in the closed arms was statistically significantly longer in the LPS group than in the control group ( $p<0.05$ ). The time spent in the closed arms was statistically significantly shorter in the LPS+Kaempferol50, LPS+Isorhamnetin30, and LPS+Isorhamnetin60 groups than in the LPS group ( $p<0.05$ ) (Table 3).

*Effect of Kaempferol and Isorhamnetin on FST*

In the FST, the duration of total immobility was evaluated. The duration of total immobility was statistically significantly longer in the LPS group than in the control group ( $p<0.05$ ), and statistically significantly shorter in the LPS+Kaempferol50, and LPS+Isorhamnetin30 groups than in the LPS group ( $p<0.05$ ) (Table 4).

*Effect of Kaempferol and Isorhamnetin on TST*

In the TST, the duration of total immobility was evaluated. The duration of total immobility was statistically significantly longer in the LPS group than in the control group ( $p<0.05$ ), and statistically significantly shorter in all the treatment groups than in the LPS group ( $p<0.05$ ) (Table 5).

**Table 1.** Open Field test results

	The time spent in the middle area (sec)
<b>Control</b>	82.2 ± 7.3
<b>LPS</b>	46.0 ± 6.5*
<b>LPS+Kaempferol50</b>	70.4 ± 7.9 <sup>#</sup>
<b>LPS+Kaempferol100</b>	53.8 ± 4.7
<b>LPS+Isorhamnetin30</b>	73.0 ± 9.3 <sup>#</sup>
<b>LPS+Isorhamnetin60</b>	49.8 ± 8.7

Data are presented as the mean ± SD

ANOVA,  $p<0.05$

\* $p<0.05$  versus Control

<sup>#</sup> $p<0.05$  versus LPS

**Table 2.** Light/Dark test results

	The time spent in the light area (sec)	The time spent in the dark area (sec)
<b>Control</b>	130.4 ± 11.9	169.6 ± 11.9
<b>LPS</b>	52.4 ± 7.3*	247.6 ± 7.3*
<b>LPS+Kaempferol50</b>	86.4 ± 12.7 <sup>#</sup>	213.6 ± 12.7 <sup>#</sup>
<b>LPS+Kaempferol100</b>	73.4 ± 9.2 <sup>#</sup>	226.6 ± 9.2 <sup>#</sup>
<b>LPS+Isorhamnetin30</b>	76.4 ± 7.5 <sup>#</sup>	223.6 ± 7.5 <sup>#</sup>
<b>LPS+Isorhamnetin60</b>	64.8 ± 8.7	235.2 ± 8.7

Data are presented as the mean ± SD

ANOVA,  $p<0.05$

\* $p<0.05$  versus Control

<sup>#</sup> $p<0.05$  versus LPS

**Table 3.** Elevated Plus-Maze test results

	The time spent in the open arms (sec)	The time spent in the closed arms (sec)
<b>Control</b>	73.0 ± 8.8	231.4 ± 10
<b>LPS</b>	20.8 ± 4.3*	284.6 ± 12.1*
<b>LPS+Kaempferol50</b>	54.4 ± 10.6 <sup>#</sup>	269.4 ± 14.6
<b>LPS+Kaempferol100</b>	71.8 ± 7.8 <sup>#</sup>	242.0 ± 11.2 <sup>#</sup>
<b>LPS+Isorhamnetin30</b>	50.4 ± 8.8 <sup>#</sup>	257.2 ± 15.1 <sup>#</sup>
<b>LPS+Isorhamnetin60</b>	72.0 ± 8.5 <sup>#</sup>	231.6 ± 7.9 <sup>#</sup>

Data are presented as the mean ± SD

ANOVA, p&lt;0.05

\*p&lt;0.05 versus Control

#p&lt;0.05 versus LPS

**Table 4.** Forced Swimming test results

	Immobility (sec)
<b>Control</b>	58.8 ± 8.6
<b>LPS</b>	87.8 ± 6.2*
<b>LPS+Kaempferol50</b>	77.0 ± 8.3
<b>LPS+Kaempferol100</b>	61.8 ± 7.5 <sup>#</sup>
<b>LPS+Isorhamnetin30</b>	75.6 ± 9.8
<b>LPS+Isorhamnetin60</b>	61.4 ± 8.7 <sup>#</sup>

Data are presented as the mean ± SD

ANOVA, p&lt;0.05

\*p&lt;0.05 versus Control

#p&lt;0.05 versus LPS

**Table 5.** Tail Suspension test results

	Immobility (sec)
<b>Control</b>	130.6 ± 15.7
<b>LPS</b>	223.2 ± 22.2*
<b>LPS+Kaempferol50</b>	178.4 ± 8.6 <sup>#</sup>
<b>LPS+Kaempferol100</b>	153.0 ± 9.3 <sup>#</sup>
<b>LPS+Isorhamnetin30</b>	143.0 ± 7.8 <sup>#</sup>
<b>LPS+Isorhamnetin60</b>	123.8 ± 10.6 <sup>#</sup>

Data are presented as the mean ± SD

ANOVA, p&lt;0.05

\*p&lt;0.05 versus Control

#p&lt;0.05 versus LPS

## Biochemical Analysis Results

### Effect of Kaempferol and Isorhamnetin on Prefrontal and Hippocampal Oxidative Stress

In the evaluation of the MDA level in the prefrontal and hippocampal regions, the MDA level was statistically significantly higher in the LPS group than in the other groups (p<0.05) (Figure 2b). The prefrontal and hippocampal MDA levels in the treatment groups were statistically significantly lower than those of the

LPS group (p<0.05). The TOS values in the hippocampal area were statistically significantly lower in the control group than in the LPS group (p<0.05). Except for the LPS+Kaempferol50 group, the values of the LPS group were statistically significantly higher than in the other groups where flavonoid was used (p<0.05). The TOS values in the prefrontal area were statistically significantly higher in the LPS group than in the other groups (p<0.05) (Figure 2a).

The TAS values in the hippocampal area were statistically significantly lower in the LPS group than in the other groups ( $p<0.05$ ). In the evaluation of the TAS values in the prefrontal area, no statistically significant difference was determined between the control group and the LPS group ( $p>0.05$ ). The values in the LPS group were statistically significantly lower than in the LPS+Kaempferol100, and LPS+Isorhamnetin60 groups ( $p<0.05$ ) (Figure 2c).

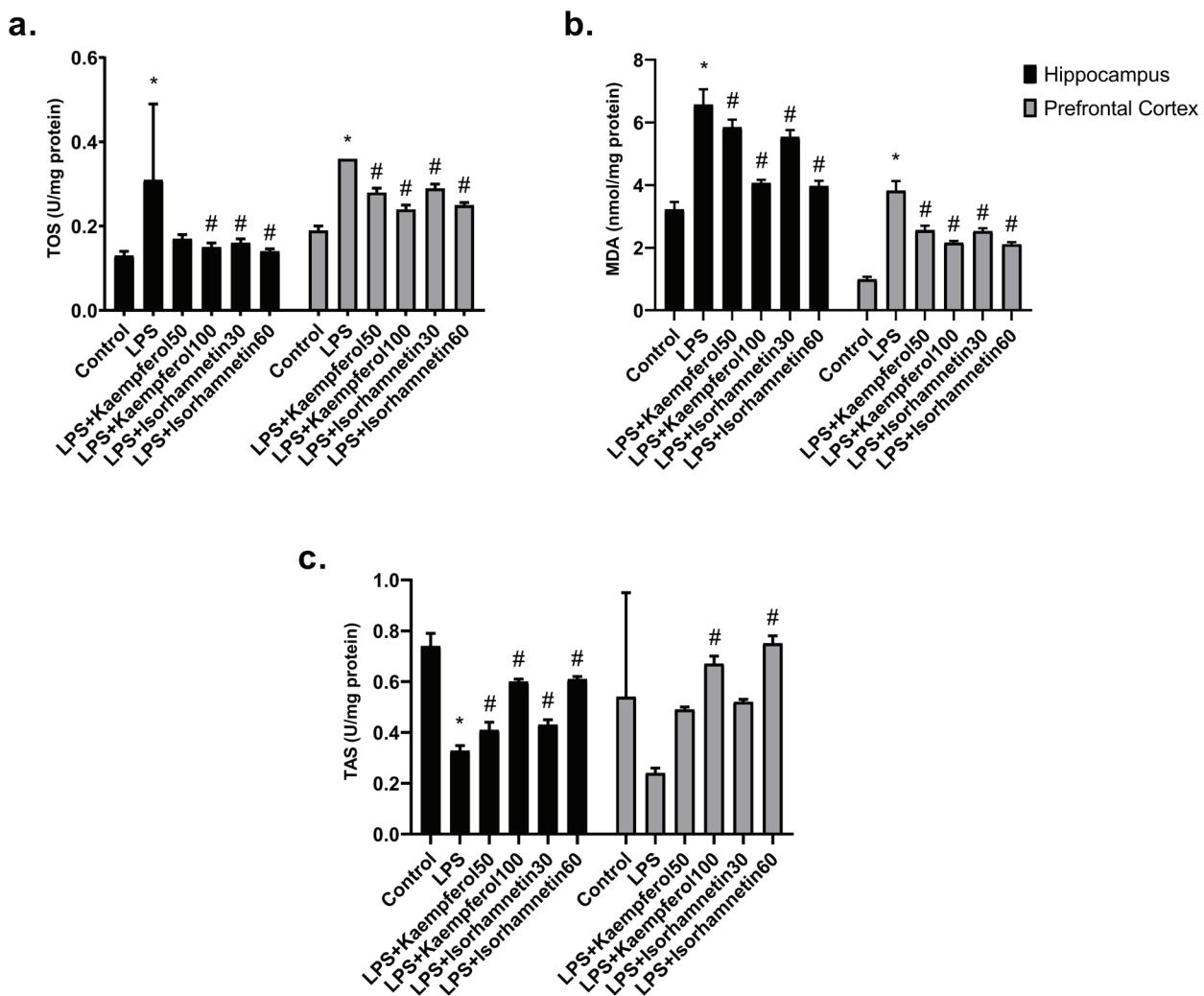
#### *Effect of Kaempferol and Isorhamnetin on Prefrontal and Hippocampal Inflammation*

TNF- $\alpha$ , IL-6, and IL-1 $\beta$  values in the prefrontal and hippocampal areas were determined to be statistically significantly higher in the LPS group than in the control group ( $p<0.05$ ). However, Kaempferol

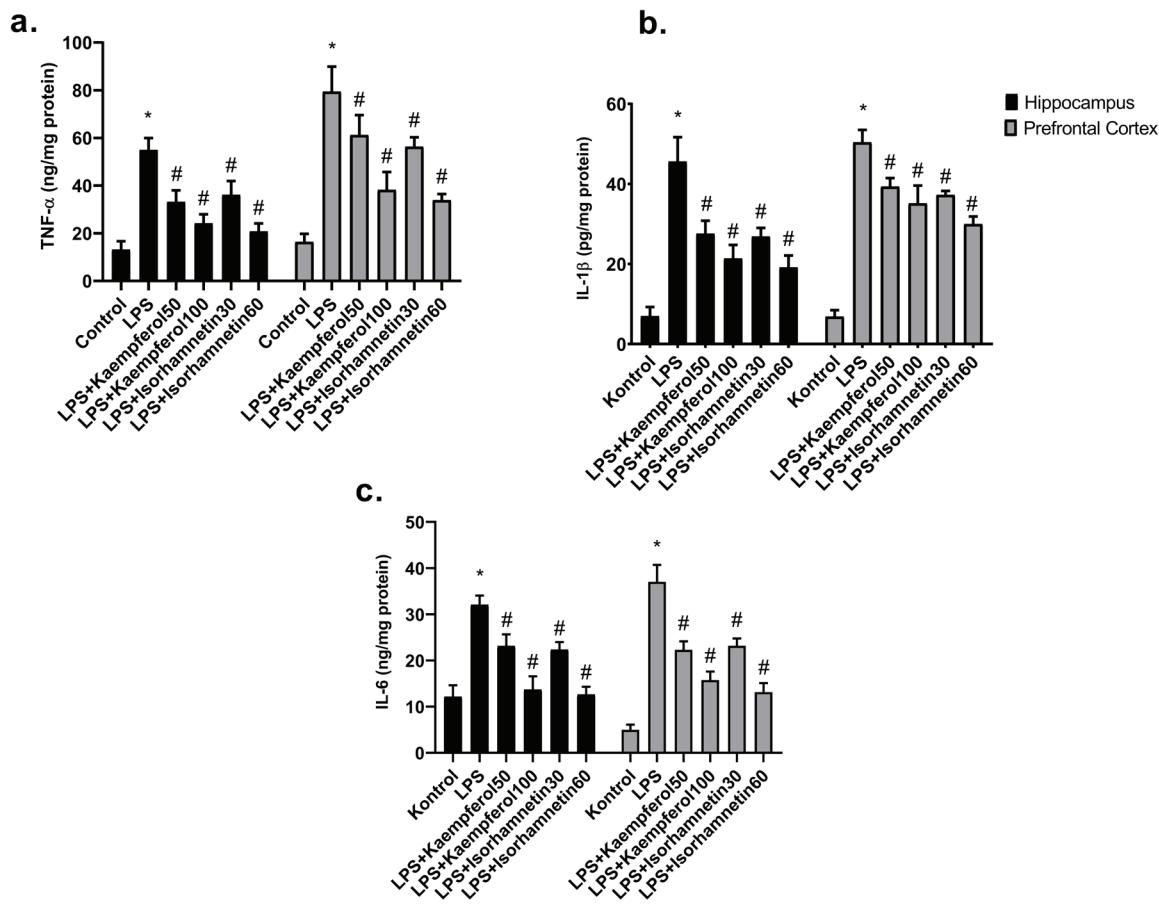
and Isorhamnetin statistically decreased these proinflammatory cytokines in both brain tissue ( $p<0.05$ ) (Figure 3a, 3b, 3c).

#### *Effect of Kaempferol and Isorhamnetin on Prefrontal and Hippocampal BDNF levels*

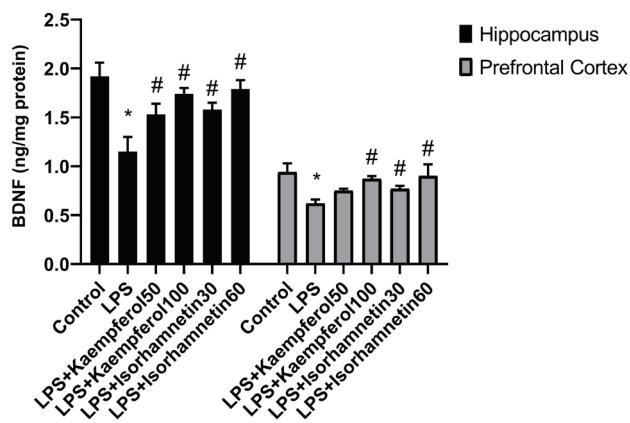
The BDNF level in the hippocampal area was statistically significantly lower in the LPS group than in the control group ( $p<0.05$ ). However, both treatment groups statistically increased the hippocampal level of BDNF ( $p<0.05$ ). On the other hand, BDNF level in the prefrontal area was statistically significantly lower in the LPS group than in the control, LPS+Kaempferol100, LPS+Isorhamnetin30, and LPS+Isorhamnetin60 groups ( $p<0.05$ ) (Figure 4).



**Figure 2:** The effect of Kaempferol and Isorhamnetin on oxidative stress in the prefrontal cortex and hippocampus. The changes of TOS (a), MDA (b), and TAS (c) levels. \* $p<0.05$  compared to Control group, # $p<0.05$  compared to LPS group using one way ANOVA post hoc Tukey test



**Figure 3:** The effect of Kaempferol and Isorhamnetin on inflammation in the prefrontal cortex and hippocampus. The changes of TNF- $\alpha$  (a), IL-1 $\beta$  (b), and IL-6 (c) levels. \* $p<0.05$  compared to Control group, # $p<0.05$  compared to LPS group using one way ANOVA post hoc Tukey test



**Figure 4:** The effect of Kaempferol and Isorhamnetin on BDNF levels in the prefrontal cortex and hippocampus. \* $p<0.05$  compared to Control group, # $p<0.05$  compared to LPS group using one way ANOVA post hoc Tukey test

## DISCUSSION

The results of this study demonstrated that Kaempferol and Isorhamnetin, which have antioxidant, anti-inflammatory, and neuroprotective properties,

improved the LPS-related behavioral changes in an LPS-induced experimental depression and anxiety model. These dose-dependent anxiolytic and antidepressant-like effects were determined to have occurred

by decreasing the increase in oxidative stress and inflammation and increasing the decreased BDNF in the hippocampus and prefrontal cortex caused by LPS.

Various behavioral tests are used to evaluate anxiety and depression in rodents. The tests in common use, the OFT, Light-Dark test, and EPM test are used to determine anxiety level, and the TST and the FST to determine the level of depression (Rammal et al., 2008; Mesfin et al., 2014; Sulakhiya et al., 2016). As there is a high frequency of anxiety and depression seen together and usually the same group of drugs are used in treatment, animal experimental apparatus for both anxiety and depression were used in this study. As a result of the tests applied, anxiety and depression-like behaviors were determined in the mice that were administered LPS, which was a result consistent with the findings of several studies (Jangra et al., 2014; Sulakhiya et al., 2014).

It is known that flavonoids obtained from several natural plants show an antidepressant and anxiolytic-like effect in experimental animal depression and anxiety models (Zhang et al., 2012; Ko et al., 2020). Although there are a few studies related to the anxiolytic and antidepressant effects of Kaempferol, there are insufficient studies of Isorhamnetin. In the current study, Kaempferol and Isorhamnetin were determined to have reduced anxiety and depression-like behaviors. This finding demonstrates that Kaempferol and Isorhamnetin could have a potential healing effect on anxiety and depression symptoms.

For many years the hippocampus has been known to be associated with the regulation of learning and memory abilities, and it has an important role in mood regulation (Gould et al., 1992; Cameron and Gould, 1994). The findings in the hippocampus have also been reported to provide a gateway to the prefrontal cortex, which is another important brain region associated with stress and stress-related behaviors, and is important for working memory, executive function, and self-regulatory behaviors, and which responds to stress (McEwen and Morrison, 2013; McEwen et al., 2016). These two regions of the brain are areas affected by many psychiatric diseases, especially anxiety and depression (Liu et al., 2017). In this study, it was determined that MDA and TOS levels increased and TAS levels decreased in the hippocampus and prefrontal areas after the application of LPS. This result shows an increase in oxidative stress in the hippocampal and prefrontal areas due to LPS.

Previous studies have reported that oxidative stress plays an important role in the physiopathology of anxiety and depression (Bouayed et al., 2009; Grases et al., 2014). It has also been reported that while there is an increase in lipid peroxidation in anxiety and depression, there is a decrease in DNA damage, glutathione and antioxidant enzyme activity (Bouayed et al., 2009; Grases et al., 2014). In a mouse model of chronic social defeat stress (CSDS), Kaempferol increased the levels of SOD, GPx, CAT, and GST which were reduced in the prefrontal cortex, and decreased the MDA level (Gao et al., 2019). In a diabetic rat model, Isorhamnetin was shown to reduce the increased oxidative stress in the brain (Jamali-Raeufy et al., 2019). In addition, Isorhamnetin has been reported to suppress the increasing LPS-induced oxidative stress in BV2 microglia cell line (Xu et al., 2012). In the current study, Kaempferol and Isorhamnetin were determined to have reduced the oxidative stress formed in the hippocampus and prefrontal cortex with LPS. The use of Kaempferol and Isorhamnetin together can be evaluated as a promising approach to provide protection against anxiety and depression by suppressing oxidative stress.

There are studies in the literature showing that proinflammatory cytokines play an important role in the physiopathology of mood disorders (Sulakhiya et al., 2016; J. Zhao et al., 2016). In experimental studies to investigate the inflammatory mechanism of anxiety and depression, lipopolysaccharide (bacterial endotoxin) is one of the most widely used substances (O'Connor et al., 2009; Mesfin et al., 2014; Sulakhiya et al., 2016). It has been reported that anxiety and depression-like behaviors could be formed by the administration of LPS causing the expression of proinflammatory cytokines, such as TNF- $\alpha$  and IL-1 $\beta$  in the periphery and in the brain in experimental animals (Jangra et al., 2014; Sulakhiya et al., 2014). It has been reported high concentrations of proinflammatory cytokines, such as TNF- $\alpha$  and IL-1 $\beta$  in the serum or plasma of depressive patients (Hannestad et al., 2011).

By triggering an increase in oxidative stress and proinflammatory cytokines, LPS may also cause activation of the hypothalamus-hypophyseal-adrenal axis, consequently causing the expression of corticotropin-releasing hormone (CRH) related to the behavioral response such as anxiety and depression in animals (Sah et al., 2011). In the current study, LPS was determined to have increased the levels of

proinflammatory cytokines (TNF- $\alpha$ , IL-1 $\beta$ , and IL-6) in the hippocampus and prefrontal cortex. In an LPS-induced neuroinflammation mouse model study by Cheng *et al.*, it was reported that Kaempferol decreased levels of IL-1 $\beta$ , IL-6, TNF- $\alpha$ , MCP-1, and COX-2 in brain tissue (Cheng *et al.*, 2018).

In an experimental diabetic rat model, Isorhamnetin reduced increasing TNF- $\alpha$  levels in the brain (Xu *et al.*, 2012). In another study of ischemic brain damage in mice, isorhamnetin reduced the increasing IL-1 $\beta$ , IL-6, and TNF- $\alpha$  levels in the ipsilateral cortex (Zhao *et al.*, 2016). In the current study, Kaempferol and Isorhamnetin were observed to reduce the increasing proinflammatory cytokine levels due to LPS. When these results are examined, inflammation is significant in the pathophysiology of anxiety and depression.

Neurotrophins are proteins strongly associated with the survival, development and helping with the function of neurons. BDNF is one of the neurotropic factors found most abundantly in the central nervous system. Studies have shown the importance of BDNF on the pathogenesis of depression and anxiety (Chen *et al.*, 2006; Kalueff *et al.*, 2006). Clinical studies have also reported that BDNF concentration is reduced in the blood of depressive patients, but returns to normal levels with antidepressant treatment (Karege *et al.*, 2002; Shimizu *et al.*, 2003; Brunoni *et al.*, 2008). There are also studies in literature showing that LPS application suppresses hippocampal BDNF production and that this shows a negative correlation with depression (Satomura *et al.*, 2011; Jangra *et al.*, 2014; Lin and Wang, 2014). In the current study, LPS was determined to have reduced BDNF levels in the hippocampus and prefrontal cortex. Kaempferol and Isorhamnetin were determined to have increased the reduced BDNF levels. These results demonstrate that neurotropic factors such as Kaempferol and Isor-

hamnetin, which increase the molecules of BDNF, could play a key role in improving anxiety and depression symptoms.

The fact that the additive effects of Kaempferol and Isorhamnetin have not been investigated is a limitation of this study. In addition, the fact that molecular techniques were not used in this study could be a limitation of the study.

## CONCLUSIONS

The results of this study not only showed the importance of oxidative stress, inflammation, and plasticity in the etiopathogenesis of anxiety and depression, but also the efficacy of Kaempferol and Isorhamnetin in protecting against these two disorders could occur by increasing the antioxidative and anti-inflammatory effects and neuronal plasticity. Nevertheless, there is a need for further studies to show by which effect mechanism this protective effect is provided and that it could be a potential therapeutic agent.

## ACKNOWLEDGMENTS

The authors would like to thank from bottom of their hearts all participants and their parents. This study was supported by a grant from Scientific Research Project Coordination Unit of Cumhuriyet University (Project ID no. T-804). ME, HG, and DGM contributed substantially to the conception and design of the study. ME, HG, and DGM acquired the data, and all authors contributed to its analysis and interpretation. ME drafted the manuscript and HG and DGM critically revised it for important intellectual content. All the authors approved the final version submitted for publication in the published article.

## CONFLICT OF INTEREST

The authors have no conflicts of interest to declare.

## REFERENCES

Bouayed J, Rammal, H, Soulimani, R (2009) Oxidative stress and anxiety: relationship and cellular pathways. *Oxid Med Cell Longev* 2(2): 63–67.

Brenes GA (2007) Anxiety, depression, and quality of life in primary care patients. *Prim Care Companion J Clin Psychiatry* 9(6): 437–443.

Brunoni AR, Lopes M, Fregni F (2008) A systematic review and meta-analysis of clinical studies on major depression and BDNF levels: implications for the role of neuroplasticity in depression. *Int J Neuropsychopharmacol* 11(8):1169–1180.

Cameron HA, Gould E (1994) Adult neurogenesis is regulated by adrenal steroids in the dentate gyrus. *Neuroscience* 61(2): 203–209.

Chen ZY, Jing D, Bath KG, Ieraci A, Khan T, Siao CJ, Herrera DG, Toth M, Yang C, McEwen BS, Hempstead BL, Lee FS (2006) Genetic variant BDNF (Val66Met) polymorphism alters anxiety-related behavior. *Science* 314(5796): 140–143.

Cheng X, Yang YL, Yang H, Wang YH, Du GH (2018) Kaempferol alleviates LPS-induced neuroinflammation and BBB dysfunction in mice via inhibiting HMGB1 release and down-regulating TLR4/MyD88 pathway. *Int Immunopharmacol* 56: 29–35.

Chi G, Zhong W, Liu Y, Lu G, Lü H, Wang D, Sun F (2016) Isorhamnetin protects mice from lipopolysaccharide-induced acute lung injury via the inhibition of inflammatory responses. *Inflamm Res* 65(1): 33–41.

Gao W, Wang W, Peng Y, Deng Z (2019) Antidepressive effects of kaempferol mediated by reduction of oxidative stress, proinflammatory cytokines and up-regulation of AKT/β-catenin cascade. *Metab Brain Dis* 34(2): 485–494.

Gould E, Cameron HA, Daniels DC, Woolley CS, McEwen BS (1992) Adrenal hormones suppress cell division in the adult rat dentate gyrus. *J Neurosci* 12(9): 3642–3650.

Grases G, Colom MA, Fernandez RA, Costa-Bauzá A, Grases F (2014) Evidence of higher oxidative status in depression and anxiety. *Oxid Med Cell Longev* 2014: 430216.

Hannestad J, DellaGioia N, Bloch M (2011) The effect of antidepressant medication treatment on serum levels of inflammatory cytokines: a meta-analysis. *Neuropsychopharmacology* 36(12): 2452–2459.

Hussein RM, Mohamed WR, Omar HA (2018) A neuroprotective role of kaempferol against chlorpyrifos-induced oxidative stress and memory deficits in rats via GSK3β-Nrf2 signaling pathway. *Pestic Biochem Physiol* 152: 29–37.

Jamali-Raeufy N, Baluchnejadmojarad T, Roghani M, Keimasi S, Goudarzi M (2019) Isorhamnetin exerts neuroprotective effects in STZ-induced diabetic rats via attenuation of oxidative stress, inflammation and apoptosis. *J Chem Neuroanat* 102: 101709.

Jangra A, Lukhi MM, Sulakhiya K, Baruah CC, Lahkar M (2014) Protective effect of mangiferin against lipopolysaccharide-induced depressive and anxiety-like behaviour in mice. *Eur J Pharmacol* 740: 337–345.

Kalueff AV, Avgustinovich DF, Kudryavtseva NN, Murphy DL (2006) BDNF in anxiety and depression. *Science* 312(5780):1598–1599.

Karege F, Perret G, Bondolfi G, Schwald M, Bertschy G, Aubry JM (2002) Decreased serum brain-derived neurotrophic factor levels in major depressed patients. *Psychiatry Res* 109(2); 143–148.

Ko YH, Kim SK, Lee SY, Jang CG (2020) Flavonoids as therapeutic candidates for emotional disorders such as anxiety and depression. *Arch Pharm Res* 43(11); 1128–1143.

Lin YL, Wang S (2014) Prenatal lipopolysaccharide exposure increases depression-like behaviors and reduces hippocampal neurogenesis in adult rats. *Behav Brain Res* 259: 24–34.

Liu W, Ge T, Leng Y, Pan Z, Fan J, Yang W, Cui R (2017) The Role of Neural Plasticity in Depression: From Hippocampus to Prefrontal Cortex. *Neural Plast* 2017; 6871089.

McEwen BS, Morrison JH (2013) The brain on stress: vulnerability and plasticity of the prefrontal cortex over the life course. *Neuron* 79(1); 16–29.

McEwen BS, Nasca C, Gray JD (2016) Stress Effects on Neuronal Structure: Hippocampus, Amygdala, and Prefrontal Cortex. *Neuropsychopharmacology : Official Publication of the American College of Neuropsychopharmacology* 41(1); 3–23.

Mesfin M, Asres K, Shibeshi W (2014) Evaluation of anxiolytic activity of the essential oil of the aerial part of *Foeniculum vulgare* Miller in mice. *BMC Complement Altern Med* 14; 310.

Mormède C, Palin K, Kelley KW, Castanon N, Dantzer R (2004) Conditioned taste aversion with lipopolysaccharide and peptidoglycan does not activate cytokine gene expression in the spleen and hypothalamus of mice. *Brain Behav Immun* 18(2); 186–200.

O'Connor JC, Lawson MA, André C, Moreau M, Lestage J, Castanon N, Kelley KW, Dantzer R (2009) Lipopolysaccharide-induced depressive-like behavior is mediated by indoleamine 2,3-dioxygenase activation in mice. *Mol Psychiatry* 14(5); 511–522.

Ohkawa H, Ohishi N, Yagi K (1979) Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Anal Biochem* 95(2); 351–358.

Parrella E, Del Gallo F, Porrini V, Gussago C, Benarese M, Fabene PF, Pizzi M (2022) Age-Dependent Neuropsychiatric Symptoms in the NF-κB/c-Rel Knockout Mouse Model of Parkinson's Disease. *Front Behav Neurosci* 11(16); 831664.

Peng J, Liu Y, Umpierre AD, Xie M, Tian DS, Richardson JR, Wu LJ (2019) Microglial P2Y12 receptor regulates ventral hippocampal CA1 neuronal excitability and innate fear in mice. *Mol Brain* 12(1); 71.

Rammal H, Bouayed J, Younos C, Soulimani R (2008) Evidence that oxidative stress is linked to anxiety-related behaviour in mice. *Brain Behav Immun* 22(8); 1156–1159.

Sah SP, Tirkey N, Kuhad A, Chopra K (2011) Effect of quercetin on lipopolysaccharide induced-sickness behavior and oxidative stress in rats. *Indian J Pharmacol* 43(2); 192–196.

Saki K, Bahmani M, Rafieian-Kopaei M (2014) The effect of most important medicinal plants on two important psychiatric disorders (anxiety and depression)-a review. *Asian Pac J Trop Med* 7S1; S34–42.

Satomura E, Baba H, Nakano Y, Maeshima H, Suzuki T, Arai H (2011) Correlations between brain-derived neurotrophic factor and clinical symptoms in medicated patients with major depression. *J Affect Disord* 135(1–3); 332–335.

Shimizu E, Hashimoto K, Okamura N, Koike K, Komatsu N, Kumakiri C, Nakazato M, Watanabe H, Shinoda N, Okada S, Iyo M (2003) Alterations of serum levels of brain-derived neurotrophic factor (BDNF) in depressed patients with or without antidepressants. *Biol Psychiatry* 54(1); 70–75.

Steimer T (2002) The biology of fear- and anxiety-related behaviors. *Dialogues Clin Neurosci* 4(3); 231–249.

Sulakhiya K, Keshavlal GP, Bezbarua BB, Dwivedi S, Gurjar SS, Munde N, Jangra A, Lahkar M, Gogoi R (2016) Lipopolysaccharide induced anxiety- and depressive-like behaviour in mice are prevented by

chronic pre-treatment of esculetin. *Neurosci Lett* 611; 106–111.

Sulakhiya K, Kumar P, Jangra A, Dwivedi S, Hazarika NK, Baruah CC, Lahkar M (2014). Honokiol abrogates lipopolysaccharide-induced depressive like behavior by impeding neuroinflammation and oxi-do-nitrosative stress in mice. *Eur J Pharmacol* 744; 124–131.

Xu SL, Choi RCY, Zhu KY, Leung K-W, Guo AJY, Bi D, Xu H, Lau DTW, Dong TTX, Tsim KWK (2012) Isorhamnetin, A Flavonol Aglycone from *Ginkgo biloba* L., Induces Neuronal Differentiation of Cultured PC12 Cells: Potentiating the Effect of Nerve Growth Factor. *Evid Based Complement Alternat Med* 2012; 278273.

Zhang L-M, Yao J-Z, Li Y, Li K, ChenH-X, Zhang Y.-Z, Li Y-F (2012) Anxiolytic effects of flavonoids in animal models of posttraumatic stress disorder. *Evid Based Complement Alternat Med* 2012; 623753.

Zhao JJ, Song JQ, Pan SY, Wang K (2016) Treatment with Isorhamnetin Protects the Brain Against Ischemic Injury in Mice. *Neurochem Res* 41(8); 1939–1948.