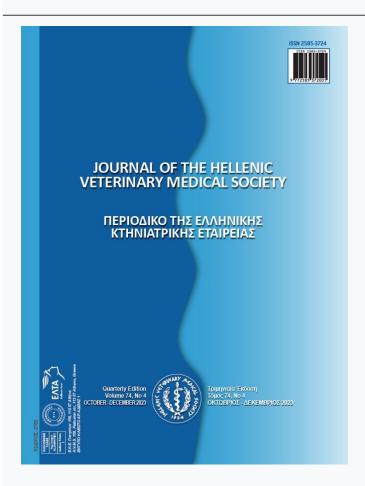




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First isolation of trh positive Vibrio alginolyticus from Engraulis encrasicolus in **Turkey**

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ABSTRACT: Engraulis encrasicolus is the most popular fish species among consumers in Turkey especially during winter months. E. encrasicolus, which is generally found in public markets, is a small fish that takes a long time to clean and therefore is often marketed without cleaning. Although large fish are exclusively handled by sellers, more people come into contact with smaller fish such as E. encrasicolus during home cleaning. This means that a zoonosis in fish such as E. encrasicolus could pose a widespread public health concern. Vibrio alginolyticus, which has virulence genes, can particularly cause wound infection through wounds and abrasions on people's hands while cleaning fish. It can lead to severe food poisoning when it enters the body orally from hands contaminated with pathogens or from the environment. Thermostable direct hemolysin (tdh) and tdh-related hemolysin (trh) are the gene regions that significantly increase the pathogenicity of Valginolyticus. In this study, a total of 200 (20 from each sale point) E. encrasicolus purchased from 10 different sale points in 4 different public markets were investigated bacteriologically. V. alginolyticus was isolated from the internal organs (spleen, kidney, liver) of 14 fish (7%) from 4 sale points in two public markets and was identified by biochemical methods. The isolates were then confirmed by PCR and trh-tdh virulence genes were investigated. While the trh was detected in 6 of 14 isolates (42.8 %), tdhwas not found in any of them (0%). In this study, trh positive V. alginolyticus was isolated from E. encrasicolus for the first time in Turkey. It should be emphasized that zoonotic agents in E. encrasicolus may cause widespread public health problems. However, there is need for more comprehensive studies to be conducted especially in the Çanakkale province and Black Sea region where *E. encrasicolus* are extensively harvested.

Key Words: Aegean region; Engraulis encrasicolus; Thermostable direct hemolysin (tdh); tdh -related hemolysin (trh), Vibrioalginolyticus

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INTRODUCTION

Ingraulis encrasicolus, of the Engraulidae family, L is partially found in temperate waters and is generally caught in the Black Sea and Canakkale Straits in Turkey (Türksönmez and Diler, 2019). According to the fisheries statistics for 2020, 291.910 tons of E. encrasicolus wereharvested from Turkish waters and it is the pelagic sea fish species which is caught the most (TUIK, 2020). It has been reported that the total sum of E. encrasicolus catch in 2021 was 171.253 tons (TUIK, 2021). Again, in numerous scientific studies conducted in many cities in Turkey, E. encrasicolus ranks first among the most consumed marine fish (Selvi et al., 2022; Kuşat and Şahan, 2021; Deniz and Sarıözkan, 2020; Yüksel and Diler, 2019). The consumption of E. encrasicolus has spread from the Black Sea region to the rest of the country despite low consumption rate of other fish. E. encrasicolus may bethe only fish species consumed by the majority in Turkey.

Vibrioalginolyticus is the major pathogen of fish and bivalves; it is rare in humans and does not cause a serious infection except to immunosuppressed people. However, it can cause serious food poisoning to humans, when it has virulence genes such as trh and tdh. Therefore, V. alginolyticus is considered an emerging pathogen (Xie et al., 2005; Smolikova et al., 2001; W. Winn Jr., et al., 2006). In Turkey, large fish such as sea bass, sea bream, mullet, bonito, trout and carp are generally sold after they are cleaned by the sellers. However, small fish such as *E. encrasicolus* are often sold without cleaning, as they are cheaper and harder to clean. That's why only sellers come into contact with large fish, while smaller fish, such as E. encrasicolus, are handled by more people in their homes. V. alginolyticus can infect the eyes and ears of people who handle the fish through contact and wounds on the hands.It can cause food poisoning directly by taking the hand to the mouth, and indirectly by splatter of the fish parts to the greens (arugula, cress, parsley) served as a side dish to the fish (Rubin and Tilton, 1975; Pien et al., 1977; Lee et al., 2008; Reilly et al., 2011; Neill and Carpenter, 2010).

The first *trh* positive *V. alginolyticus* in the world was reported in Alaska and Tunisia (Narjol et al., 2006; Ben kahla-Nakbi et al., 2006). The first *trh* positive *V. alginolyticus* isolation in Turkey was reported by Avsever from bivalve molluscs in 2016 (Avsever, 2016). Both in Turkey and in the world, *trh* and *tdh* genes have mostly been studied in *Vibrio parahae*-

molyticus (Terzi et al., 2009; Türk et al., 2011; Leoni et al., 2016; Raghunath, 2015; Terzi-Gulel and Martinez-Urtaza, 2023). There are fewer studies investigating tdh and trh genes in V. alginolyticus (Gonzales-Escolana et al., 2006; Avsever, 2016, 2022) as tdh and trh are important virulence factors in the pathogenesis of mainly V. parahaemolyticus. However, in recent studies, V. alginolyticus was also seen to rapidly increase its pathogenicity and was reported to transfer V.parahaemolyticus virulence genesto itself (Gonzales-Escolana et al., 2006).

Although *V. alginolyticus* infections have been reported from humans (Baran et al., 2016; Citil et al., 2015) and aquatic organisms (Demir, 2012; Dolgun, 2022) in Turkey, *trh*, *tdh* genes were often uninvestigated. This makes it difficult to understand the prevalence of *V.alginolyticus* positive for virulence genes in Turkey. The presence of *trh-tdh* genes of *V.parahae-molyticus* were investigated in *Sarda sarda*, *Merlangius merlangus* and frozen *E. encrasicolus* samples in Turkey (Doğruer et al., 2022). However, there are no reports of *trh* (+) *V. alginolyticus* in *E. encrasicolus* from Turkey.

The aim of this study is to report *trh* positive *V. alginolyticus* isolation in *E. encrasicolus* for the first time in Turkey and to draw attention to the fact that home-cleaning *E. encrasicolus* may cause widespread public health problems.

MATERIAL AND METHODS

Sampling

In this study, 200 fish (11 cm +-1) (20 from each sale point) of *E. encrasicolus* purchased from 4 different public markets and 10 different sale points in Izmir were used. The average air temperature in December, the month when sampling took place, was 12 (+-2) °C. Each sample was placed in a separate small sterile bagand transported to the laboratory under cold chain.

Pathogen Isolation and Identification

Internal organs (liver, spleen and kidney) of each fish were cut into parts with sterile scissors and inoculated in Alkaline Peptone Water (APW). APW were incubated at 37 °C for 24 hours. Followed by subculture into TCBS (Thiosulfate Citrate Bile Sucrose-Merck) agar. Petri dishes were incubated at 37 °C for 48 hours. Gram-negative, oxidase-positive, motile colonies of 2-3 mm diameter were transferred to TSA (Triptic Soy Agar) and biochemical identification was

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made from the colonies on TSAafter incubation for 24 hours at 37 °C. 14 isolates identified biochemically (TS/TS ISO 8914, 1998; Austin and Austin, 2016).

Molecular Identification of Pathogens

These isolates were confirmed with PCR(Polimerase Chain Reaction). DNA isolation was carried out from these isolates using a commercial DNA extraction kit (High Pure, Germany). Isolates were thereby confirmed as described by Luo and Hu (2008) with primers targeting the gyrB gene in V. alginolyticus. V. alginolyticus ATCC 17749 was used as a positive control. The primer sequence used in the study was 5'-TCA GAG AAA GTT GAG CTA ACG ATT-3' (AlgF1, forward) and 5'- CAT CGT CGC CTG AAG TCG CTG T -3' (AlgR1, reverse). The total volume for the PCR reaction was 25 μl; 0.4 μM AlgF1 (2 μl), 0.4 μM AlgR1 (2 μl), 5 μl genomic DNA, Taq DNA polymerase (5 units / μl) (0.40 μl) (MBI, Fermantas), 10xPCR buffer (2.50 μl), 50 mM MgCl2 (1.25 μl), 10 mM dNTPs (dCTP, dATP, dTTP, dGTP) (0.63 µl), and 11.22 µl nuclease free water. The PCR was performed as follows; an initial denaturation at 94 ° C for 4 min; 32 cycles of denaturation at 94 ° C for 30 s, annealing at 64 ° C for 30 s, extension at 72 ° C for 1 min, and a final extension at 72 ° C for 8 min. After PCR amplification, 4 µl of amplification product was loaded into a 1.0% agarose gel for electrophoresis. DNA size marker 100 DNA Ladder Plus (MBI Fermentas) was used. After electrophoresis, bands were visualized with designated equipment.

Investigation of trh and tdh genes

Primers used were the forward and reverse primers. (PCR) primers, targets and amplification sizes are shown in Table 1.

Vibrio parahaemolyticus DNA samples (Terzi et al.2009) were used as positive control, while negative control consisted of distilled sterile water. For PCR (trh), reaction volume consisted of; 5 μl genomic DNA, 5 μM TRH-L primer (2.0 μl), 5 μM TRH-R primer (2.0 μl), Taq DNA polymerase (5 units / μl) (0.40 μl) (MBI, Fermantas), 10xPCR buffer (2.50 μl), 50 mM MgCl2 (1.25 μl), 10 mM dNTPs (dCTP,

dATP, dTTP, dGTP) (0.63 µl) and 11.22 µl nuclease free water (Total 25 µl). For PCR (tdh), the master mix consisted of; 5 µl genomic DNA, 5 µM TDH-L primer (1 µl), 5 µM TDH-R primer (1 µl), Taq DNA polymerase (5 units / μl) (0.40 μl) (MBI, Fermantas), 10xPCR buffer (2.50 μl), 50 mM MgCl2 (1.25 μl), 10 mM dNTPs (dCTP, dATP, dTTP, dGTP) (0.63 µl) and 13.22 µl nuclease free water (Total 25 µl). PCR was performed as follows: initial denaturation at 94 $^{\circ}$ C for 5 min., 40 cycles of denaturation at 94 ° C for 30 sec., annealing at 58 ° C for 45 sec., and primer extension at 68 ° C for 75 sec. Final extension was at 68 ° C for 7 min. PCR products were evaluated with electrophoresis on a 2% (w/v) agarose gel (1 hour, 75 volts). DNA size marker 100 DNA Ladder Plus (MBI Fermentas) was used. Visualisation of bands was carried out with designated equipment.

RESULTS

A total of 200 E. encrasicolus (20 from each sale point) purchased from 10 different sale points in 4 different public markets in Izmir province were investigated bacteriologically. V. alginolyticus was isolated from the internal organs (spleen, kidney, liver) of 14 fish (7%) from 4 sale points in two public markets. Hemorrhages on the head, operculum, abdomen (Fig. 1) and enlarged internal organs (liver/spleen) were overall observations. Enlargment of the liver-spleen was observed in all 14 fish from which V. alginolyticus was isolated. However, hemorrhages on the body surface and enlargement of the liver-spleen were also noted in some fish from which V. alginolyticus could not be isolated. Isolates were identified by biochemical methods. All isolates showed the same biochemical patterns. Biochemical identification results are provided in Table 1. After the isolates were confirmed by PCR (Fig.2), trh and tdh virulence genes were investigated (Fig. 3-4). While the trh gene region was detected in 6 of the 14 isolates (42.8 %), tdh gene was not found in any of them (0%). Trh positivity rate in V. alginolyticus isolates were 33.33%, 33.33%, 0%, 80% and trh positive V. alginolyticus were found in 2.5%, 1.66%, 0%, 4% of the samples from four markets (A, B, C,D), respectively (Table 2).

Table 1. (PCR) primers, targets and amplification sizes										
PCR Primers	Targets	Amplification sizes	Literature							
GGCTCAAAATGGTTAAGCG -CATTTCCGCTCTCATATGC	<i>Trh</i> gene	250 bp	Cohen et al. (2006)							
CCATCTGTCCCTTTTCCTGC	<i>Tdh</i> gene	373 bp	Cohen et al. (2006)							

Table 2. Biochemical propertie	es of <i>V. alginolyticus</i> isolates
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Gram	+	Urease	_	LDC	+	Growth in 22°C	+	MR/VP	+/+
Oxidase	+	ONPG	_	ADH	_	Growth in 37°C	+	Growth in %0 NaCl	_
Motility	+, Flagellar	Indole	+	10μg Ο/129	Resistant	Mannitol	+	Growth in %3 NaCl	+
Colony color in TCBS	Green	Hemolysis in Blood Agar	+	150 μg Ο/129	Sensitive	ODC	+	Growth in %8 NaCl	+
Glucose	+	Lactose	_	Oxidation/ Fermentation	+/+	H ₂ S/Gas			

Table 3: Mo	olecular	analyzes	by mar	ket p	lace and	sales	points

Public Markets	A		В			С			D					
Sale Points	a	1	b	С	d	e	f	g	h	i			j	
Isolate No:	1	2	3	Not isolated	4, 5	6	Not isolated	7, 8, 9	Not isolated	10	11	12	13	14
gyrB	+	+	+		+	+		+		+	+	+	+	+
Trh	+	-	-		-	+		-		+	+	+	-	+
Tdh	-	-	-		-	-		-		-	-	-	-	-

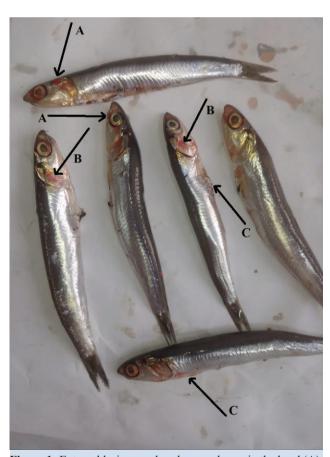


Figure 1: External lesions such as haemorrhages in the head (A), operculum (B) and the abdominal area (C) of fish infected with *V. alginolyticus*

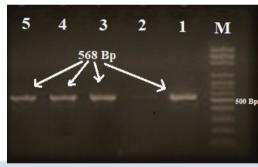


Figure 2: Results from *gyrB* PCR: Marker, 100 bp. Line 1: Positive control *V. alginolyticus* ATCC 17749, 568 bp. Line 2 Negative control distilled water. Line 3, 4, 5: Isolates (No 1,2,3), 568 bp (*gyrB*).

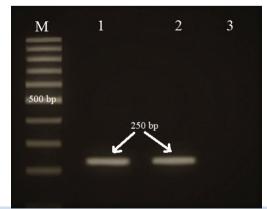


Figure 3: PCR results for virulence gene *trh*. M: Marker 100 bp. Line 1: *trh* positive control (Terzi et al, 2009), 250 bp. Line 2: *trh* positive isolate sample (No 1), 250 bp. Line 3: Negative control distilled water.

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DISCUSSION

Avsever (2022) had previously reported the isolation of trh positive V. alginolyticus from black mussels in Izmir and Çanakkale regions. In this study, trh positive V. alginolyticus was isolated from anchovies caught from the Çanakkale province and Black Sea region. According to these studies, trh-positive V. alginolyticus appears to be present in mussels from the Aegean Sea, the Dardanelles and the Black Sea. On the other hand, Terzi and Martinez (2016) detected trh positive V.parahaemolyticus in Merlangius merlangus and Sarda sarda samples from the Black Sea between 2006 and 2010. Thus, the trh gene may have been transferred from V.parahaemolyticus to V. alginolyticus. The findings of another report by Türk et al., (2011) also supports this possibility in which the trh or tdh gene was not detected in V. parahaemolyticus isolates from bivalve molluscs in the Aegean region in 2011. In conclusion, these studies show that the trh gene may have originated from the Black Sea. Since E. encrasicolus is mostly caught from the Black Sea, the presence of trh gene may be common in the Vibrios of this fish species. Similarly, in an unpublished study investigating the mucilage problem in the Marmara Sea (Anonymous, 2021), it was stated that the dominant pathogen genus that decomposes mucilage was Vibrio and with V. alginolyticus being the most common species within the genus. However, all this is an assumption and should be investigated in more regions and more fish species in order to be proven.

It should also be noted that some fish species such as anchovy and sardines (*Sardina pilchardus*) are sold in Turkish markets without being cleaned (Anonymous, 2021). This practice causes more people to come into contact with it during cleaning at home. This study as well as other similar ones may lead to the requirement of anchovy and sardines to be precleaned in the market. A legislative action is considered by the authors to be beneficial in terms of public health. There are no reports found on *trh* positive *V. alginolyticus* isolates in sardines as in anchovies. Dogruer et al., (2022) detected *V. parahaemolyticus* in frozen sardine products, but did not investigate the presence of *trh-tdh* gene.

The biochemical structure of the isolates were similar to other studies (Ardıç and Özyurt 2002; Demir, 2012; Baran et al., 2022). However, there were also differences in the types of biochemical tests performed in these studies. However, biochemical properties do not affect the pathogenicity of pathogens as

much as virulence gene regions. Serological tests, which are more deterministic on pathogenicity than the biochemical properties were not performed in this study. This was due to the imcreased cost of serological studies and the recent prominence of molecular studies. On the other hand, PCR detects the pathogen instead of a serological history of the infection and is usually able to detect its presence much earlier in the course of an infection.

We isolated V. alginolyticus from all four of the four public markets (A,B,C,D) (100%), while there was no isolation of trh positive V. alginolyticus in one (C). On the other hand, in one market, our trh positive V. alginolyticus isolation rate was 80% among V. alginolyticus isolates. There was no consistent response from the fishermen in the markets C and D when asked if they obtained their fish from different regional sources (Western Black Sea, East Black Sea etc.) However, we were informed that the fish were purchased from wholesale fish sales points in Izmir. This difference in the two markets is thought to be coincidental. To clarify this, it is necessary to repeat the study at least a few times and the origin of the fish in wholesale markets should be investigated through new research projects or by other researchers. Thus, E. encrasicolus health status in different section of the Black Sea can be elucidated.

Clinical symptoms seen in fish were overall compatible with infections caused by Vibrio spp. However, the fact that we detected hemorrhaging fins, skin and enlarged internal organs (liver/spleen) in fish negative for V. alginolyticus shows that there may be other pathogens in the collected samples. On the other hand, since the aim was to search for Vibrio species carrying the trh-tdh gene in these samples, the isolates that were not found to be Vibrio by TCBS and O 129 tests were not included in the evaluation. Therefore, different pathogens may have played a role in the lesions in fish. Vibrio spp. other than V. alginolyticus were not isolated in the samples used in the study. This may be due to the fact that *V.parahaemolyticus* is rare and fish infected with more pathogenic pathogens agents such as Vibrioanguillarum may have been eliminated because of the high mortality rate.

In Turkey, *V. alginolyticus* isolation from humans (Ardıç and Özyurt, 2002; Citil et al., 2015; Baran et al., 2022) was also reported. In order to determine whether the cause of these human cases is related to handling of fish such as *E. encrasicolus* at home, a full investigation should be carried out with evalu-

ating patient history, including possible contact with sea food followed by confirmatory phylogenetic analysis. This can be accomplished through joint multidisciplinary studies of both veterinarians and medical doctors.

CONCLUSION

As a result, in this study, trh positive V. alginolyt-

icus was isolated for the first time in *E. encrasicolus* in Turkey and potential public health problems which may arise after handling and cleaning *E. encrasicolus* at home are emphasized.

CONFLICT OF INTEREST

On behalf of all authors, the corresponding author states that there is no conflict of interest.

REFERENCES

- Anonymous (2021). Marmara İzleme Projesi (MAREM), https://www.evrensel.net/haber/442690/musilaji-parcalayan-bakteri-denizle-rde-salgina-ve-balik-populasyonlarında-azalisa-neden-olabilir
- Anette B, Ellingsen AB, Jaran S. Olsen JS, Per E. Granum PE, Liv M. Rørvik LM, González-Escalona N (2013)Genetic characterization of trh positive Vibrio spp. isolated from Norway Front Cell Infect Microbiol 25:3-107.
- Austin B and Austin DA (2016)PathogensFish Pathogens: Disease of Farmed and Wild Fish.
- Avsever ML (2016) First report of trh positive *Vibrio alginolyticus* isolates from bivalve molluscs in Turkey. Revue Méd Vét 167: 65-70
- Avsever ML (2022) Determination of tdh and trh Positive Vibrio alginolyticus Isolates from Black Mussel (Choromytilus meridionalis) in Aegean Sea coast of Turkey. Acta Aquatica Turcica 18 (3): 295 302.
- Baran B, Acar A, Genç Y, Aksu N (2016) Olgu raporu: Vibrio alginolyticus'a bağlı bir eksternal otit olgusu. TurkHijyen 73 (1): 49-54
- Ben Kahla-Nakbi A, Chaieb K, Besbes A, Zmantar T, Bakhrouf A (2006) Virulence and enteropathogensl repetitive intergenic consensus PCR of Vibrio alginolyticus strains isolated from Tunisian cultured gilthead sea bream and sea bass outbreaks. Vet Microbiol 117 (2-4):321-327
- Cohen N, Ennaji H, Hassar M, Karib H (2006) The Pathogensl quality of red meat and offal in Casablanca(Morocco). Mol Nutr Food Res 50: 547-562
- Citil BE, Derin S, Sankur F, Sahan M, Citil MU (2015) Vibrio alginolyticus Associated Chronic Myringitis Acquired in Mediterranean Waters of Turkey. Case Rep Infect Dis. Article ID 187212, pp 3.
- Demir Y (2012)Turkey de ihraç edilen Bivalvia türlerinden Vibrio türlerinin izolasyon ve identifikasyonu Etlik Vet Mikrobiyol Derg 23: 1-8.
- Doğruer Y, Telli AE, Telli N, Biçer Y (2022) Dondurulmuş Deniz Ürünlerinde Vibrio spp. 'nin İlmiğe Dayalı İzotermal Amplifikasyon (Loop Mediated Isothermal Amplification, LAMP) Yöntemiyle Belirlenmesi. Dicle Üniv Vet Fak Derg 15(1):14-19
- Dolgun O. (2022). Levreklerden (Dicentrarchus Labrax) İzole Edilen Vibrio Türlerinin Moleküler Tiplendirilmesi Ve Antibiyotik Dirençliliklerinin Belirlenmesi. Doktora tezi.
- Feoni F, Talevi G, Masini L, Ottaviani D, Rocchegiani E (2016)*Trh* (*tdh* –/ *trh* +) gene analysis of clinical, environmental and food isolates of *Vibrio parahaemolyticus* as a tool for investigating pathogenicity. International Journal of Food Microbiology 225: 43-53
- González-Escalona N, Blackstone GM, DePaola A (2006) Characterization of a Vibrio alginolyticus Strain, Isolated from Alaskan Oysters, Carrying a Hemolysin Gene Similar to the Thermostable Direct Hemolysin-Related Hemolysin Gene (trh) of Vibrio parahaemolyticus, Applied and Environmental Eicrobiology AEM. 72 (12): 7925-7929.
- Gulel TG, Urtaza JM (2023) Molecular Characterizations of Vibrio parahaemolyticus in Seafood from the Black 2 Sea, Turkey. Letters in Applied Microbiology 62 (6):494-500.
- Kuşat M, Şahan M (2021). Su Ürünleri Tüketim Tercihleri Üzerine Uşak İlinde Bir Anket Çalışması. Acta Aquatica Turcica 17 (3): 376-385.
- Lee DY, Moon SY, Lee SO, Yang HY, Lee JJ, Lee MS (2008) Septic shock due to *Vibrio alginolyticus* in a cirrhotic patient: the first case in Korea," Yonsei Medical Journal 49 (2): 329-332.
- Luo, P and Hu C (2008) Vibrio alginolyticus gyrB sequence analysis and gyrB-targeted serol identification in environmental isolates. Diseases

- of Aquatic Organisms 82: 209-216.
- Mustapha S, Ennaji MM, Cohen O (2013) Vibrio Alginolyticus: An Emerging Pathogen of Foodborne Diseases, Maejo International Journal of Science and Technology 2 (4):302-309
- Neill A and Carpenter CJ (2010) Other pathogenic vibrios, in Mandell, Douglas and Bennett's: Principles and Practice of Infectious Diseases, G. L. Mandell, J. E. Bennett, and R. Dolin, Eds., pp. 2787-2791, Churchill Livingstone, Elsevier, Philadelphia, Pa, USA, 7th edition.
- Nurittin A, Mustafa Ö (2002) Olgu raporu: Vibrio alginolyticus'a bağlı otit. Mikrobial Bulten, 38:145-148
- Pien F, Lee K, Higa H (1977)Vibrio alginolyticus infections in Hawaii. J Clin Microbiol. 5(6): 670-672.
- RaghunathR(2015)Rolesofthermostabledirecthemolysin(*Tdh*)andTdh-related hemolysin (*Trh*) in *Vibrio parahaemolyticus*. Front. Microbiol., Sec Food Microbiology Vol 5.
- Reilly GD, Reilly CA, Smith EG, and Baker-Austin C (2001) Vibrio alginolyticus-associated wound infection acquired in British waters, Guernsey, Eurosurveillance, 16 (42).
- Rubin SJ, Tilton RC (19759 Isolation of Vibrio alginolyticus from wound infections. J Clin Microbiol 2(6):556-558
- Selvi K, Kaya B, Özdikmenli Tepeli S, Kandemir G (2022) Çanakkale'nin Yenice ve Bayramiç ilçelerinde su ürünleri tüketiminin değerlendirilmesi. Acta Aquatica Turcica18 (1): 93 - 108
- Smolikova LM, Lomov LM, Khomenko TV, Murnachev GP, Kudriakova TA, Fetsaĭlova OP, Sanamiants EM, Makedonova LD, Kachkina GV, Golenishcheva EN (2001)Studies on halophilic vibrios causing a food poisoning outbreak in the city of Vladivostok. Zh Mikrobiol Epidemiol Immunobiol 6:3-7
- Terzi G, Büyüktanır Ö, Yurdusev N(2009)Detection of the *tdh* and *trh* genes in *Vibrio parahaemolyticus* isolates in fish and mussels from middle black sea coast of Turkey. Lett Appl Microbiol 49: 757-763.
- Terzi Gulel G, Martinez-Urtaza J (2016)Molecular characterizations of *Vibrio parahaemolyticus* in seafood from the Black Sea, Turkey, Letters in Applied Microbiology62 (6):494-500
- TÜİK (TurkeyStatistical Institute), (2021) Su Ürünleri İstatistikleri 2021, Ankara.
- Türk N, Avsever ML, Ün C (2011)Investigation of *tdh* and *trh* genes in *Vibrio parahaemolyticus* isoletes from isolated *Bivalve molluscs*. Bor Vet Bil Der 33 (47): 29-36.
- Türk N, Yabanlı M, Avsever ML, Aydın MA (2011) Çift kabuklu yumuşakçalardan Vibrio parahaemolyticus izolasyonu. Bor Vet Bil Der 33 (47).
- Türksönmez Ç, Diler A. (2019) Seasonal Determination of Heavy Metal Levels of Anchovy (*Engraulis encrasicolus* L.,1758) Obtained from the Marmara Sea. Acta Biologica Turcica, 32(4): 242-247.
- TS/TS ISO 8914 (1998) Microbiology-General Guidance for the detection of vibrio parahaemolyticus.
- Winn W, Allen S, Janda W, Koneman E, Procop G, Schreckenberger P, Woods G (2006) Curved Gram negative bacilli and oxidase positive fermenters: Campylobacteraceae and Vibrionaceae. In: Color Atlas and Textbook of Diagnostic Microbiology. 6th ed. Lippincott-Raven Publishers, Philadelphia, 321-361.
- Yüksel E, Diler A (2019) Ankara İlinde su ürünleri tüketim tercihlerinin belirlenmesi. Aydın Gastronomy, 3(1): 11-21.